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A selective fluorescent receptor for the determination of nickel (II) in semi-aqueous media



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1. Introduction

Design and synthesis of a selective fluorescent probes for the sensing of ionic species important from the environmental and material science point of view, mainly heavy metal ions, is a topic of growing interest in supramolecular chemistry [1–3]. A significant effort has been dedicated to the improvement of Ni²⁺ selective fluorescent receptors in the last few decades. However, in most cases fluorescence changes can only be observed in non-aqueous solvent and as well as gets affected by the common interference of other metal ions [4–9], which bound the limits for their analytical application in real samples analysis. Therefore, the development of highly sensitive and selective fluorescent receptor for Ni²⁺ in aqueous solution is very important and challenging area for the researchers [10–13].

Nickel can be found in the plasma of the blood. The absence of nickel from the plasma or red cells of a few subjects may be regarded as evidence against the existence of a physiologic function for this element [14]. In vitro nickel activates arginase, carboxylase and trypsin, and it may inhibit acid phosphatase. There is evidence that it affects the clotting mechanism by stabilizing the labile factor [15,16]. Nickel deliberation is a potential health hazard to fauna and flora if it exceeds the normal level than the cellular needs. Nickel toxicity can cause acute pneumonitis, dermatitis, asthma, eczema, cancer of lungs, sinus, stomachaches, unfavorable effects on blood

ABSTRACT

The malonohydrazide based fluorescent probe (7*E*,8*E*)–N1',N3'-bis(1-(2-hydroxyphenyl)ethylidene) malonohydrazide (receptor **5**) was designed and synthesized. It was confirmed by spectroscopic methods and the single crystal X-ray method. The receptors **5** show strong intramolecular hydrogen bonding which is useful in host–guest complexation. The fluorescence spectra of receptor **5** have shown distinct enhancement with the addition of Ni²⁺ ion over other surveyed cations. The stability constant was obtained by Benesi–Hildebrand, Scatchard and Connor's fitting methodologies. The 1:1 stoichiometry of the host–guest complexation was confirmed by Job's continuous variation method.

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(increases red blood cells) and kidneys (increases protein in urine) other disorders of respiratory system and central nervous system [17].

In current article, we report the synthesis of (7E,8E)–N1',N3'bis(1-(2-hydroxyphenyl)ethylidene) malonohydrazide (receptor **5**) and studied its binding properties towards Cr³⁺, Mn²⁺, Fe³⁺, Co²⁺, Ni²⁺, Cu²⁺, Zn²⁺, Cd²⁺, Hg²⁺, Pb²⁺ and Bi³⁺ in DMSO/H₂O (1:1, v/v). Receptor **5** has interestingly shown significant enhancement in fluorescence emission maxima in the semi-aqueous phase with Ni²⁺ metal ion. This ability to grant fluorescent enhancement in semi-aqueous phase makes its application possible in the area of biology, environmental and material sciences.

2. Experimental

2.1. Chemical and apparatus

All commercial grade chemicals and solvents were purchased from Sigma-Aldrich and used without further purification. ¹H-NMR and ¹³C-NMR spectra were recorded on Varian NMR mercury System 300 spectrometer operating at 300 and 75 MHz in DMSO-d₆ solvent. The fluorescence spectra were recorded in DMSO/H₂O (1:1, v/v) namely on Fluoromax-4 spectrofluorometer and Shimadzu UV-24500 at room temperature using 1 cm pathlength cell.

2.2. Synthesis of (7E,8E)–N1',N3'-bis(1-(2-hydroxyphenyl) ethylidene)malonohydrazide (receptor **5**)

Malonohydrazide (3) was synthesized from condensation of one mole diethyl malonate (0.32 g, 2 mM) with two moles of

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hydrazine hydrate (0.20 g, 4 mM) by stirring for 30 min in absence of solvent. Malonohydrazide (**3**) was obtained in good yield and having appearance of white powder. Receptor **5** was obtained from one mole of Malonohydrazide (**3**) (0.26 g, 2 mM) with two moles of o-hydroxy acetophenone (0.54 g, 4 mM) in ethanol with stirring and refluxing for 90 min (Scheme 1). Receptor **5** was obtained in good yield and having white appearance, solubility in DMSO; Yield 82%; Melting Point 198–200 °C. Crystals suitable for X-ray analysis for compound **5** were obtained by slow evaporation of a solution of the compound from a methanol solution in 8 days.

¹H-NMR (300 MHz, DMSO-d₆, ppm,) δ =2.42 (s, 6H, 2CH₃), 4.31 (s, 2H, CH₂), 6.91–7.13 (4H, m, Ar–H), 7.21–7.52 (t, *J*=7.5 Hz, 2H, Ar–H), 7.54–7.85 (d, 2H, *J*=7.5 Hz, Ar–H), 11.3 (s, 2H, NH), 13.2 (s, 2H, Ar–OH). ¹³C-NMR (75 MHz, DMSO-d₆) δ =13.8, 40.5, 116.5, 118.6, 120.0, 131.3, 133.7, 159.0, and 163.25. IR (KBr, cm⁻¹): *v*=623, 663, 684, 752, 781,798, 825, 893, 1033, 1099, 2937, 3057, and 3207. MS (EI): M+H⁺ calcd. for (C₁₉H₂₀N₄O₄·2Na)=413.40; found 413.74.

2.3. Cation recognition studies

The recognition studies were performed at room temperature, and the solution was shaken for enough time before recording fluorescence and absorption spectra to ensure uniformity. The



Fig. 1. ORPET diagram of receptor 5.

Table 1

Crystal data and structural refinement for rec	otor 5	j
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Parameter	Receptor 5
Molecular formula	$C_{19}H_{20}N_4O_4 \cdot 2(H_2O)$
Formula weight	404.42
Temperature	123 K
Wavelength	0.71073 Å
Crystal system	Triclinic
Space group	P1
Unit cell dimensions	a=4.977(7) Å
	b = 5.552(3) Å
	c = 17.790(14) Å
	$\alpha = 81.33(3)^{\circ}$
	$\beta = 88.25(9)^{\circ}$
	$\gamma = 82.49(4)^{\circ}$
Volume	483.7(8) Å ³
Ζ	1
Density (calculated)	1.388 Mg/m ³
Absorption coefficient	0.11 mm^{-1}
$F(0 \ 0 \ 0)$	214
Crystal size	$0.15 \times 0.08 \times 0.01 \text{ mm}^3$
Theta range for data collection	3.5–26.0°
Index ranges	$-6 \le h \le 6, -6 \le k \le 6,$ $-21 \le l \le 31$
Reflections collected	6061
Independent reflections	3140 [R(int) = 0.051]
Refinement method	Full-matrix least-squares on F^2
Data/restraints/parameters	3514/0/287
Goodness-of-fit on F^2	108
R indices (all data)	R1 = 0.067 wR2 = 0.186
Largest difference in peak and hole	0.31 and -0.50
$(e Å^{-3})$	out and out

cation binding ability of receptor **5** in a DMSO/H₂O (1:1, v/v) by keeping the solvent ratio constant throughout the experiment, was determined by preparing standard solutions of receptor **5** $(c=1 \times 10^{-5} \text{ M})$ along with fixed amounts (0.5 equiv.) of a metal nitrates $(c=1 \times 10^{-4} \text{ M})$ in DMSO/H₂O (1:1, v/v). The cation recognition performance of receptor **5** $(c=1 \times 10^{-5} \text{ M})$ was evaluated from changes in the fluorescence spectra of the sensor upon addition of the salts $(c=1 \times 10^{-4} \text{ M})$.

2.4. Stoichiometry determination

Sequentially, to determine the stoichiometry of the receptor **5** · Ni²⁺ complexation, solutions of receptor **5** and Ni²⁺ ion were prepared at ratios of 1:9, 2:8, 3:7, 4:6, 5:5, 6:4, 7:3, 8:2, and 9:1. These solutions were allowed to stand for 1 h with frequent shaking. The fluorescence spectra were recorded with each solution. The plot of [HG] versus [H]/([H]+[G]) was used to determine the stoichiometry of the complex formed. The fluorescence intensity of emission peak maxima at 629 nm was used for stoichiometry



Fig. 2. (a) Changes in fluorescent intensity of receptor **5** upon the addition of a particular metal salts in DMSO/H₂O (1:1, v/v) solvent system, (b) fluorescence ratiometric response of receptor **5** upon the addition of a particular metal salts in DMSO/H₂O (1:1, v/v) solvent system, and (c) respective interference of metals at the time of Ni²⁺ ion detection.

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