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# Kinetic investigations on alkaline fading of malachite green in the presence of micelles and reverse micelles

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## 36 1. Introduction

The chemistry of dyes and their applications to other scientific fields 37get immense popularity in the modern era. They have played an impor-38 tant role as an effective indicator in the analysis of some elements such 39 as ruthenium (Ru), iodine (I) and osmium (Os) [1]. The recognition of 40DNA by dye molecules is of paramount importance in biochemistry 41 and molecular biology [2]. Dye sensitization of semiconductors is 42 becoming a promising strategy for solar energy conversion [3]. The prin-43 cipal application of dyes is in the coloration of substrates, typically 44 45 textiles, where a great quantity of water and dyes are used, with about 50% of the dyes discharged into wastewater, thereby polluting the 46 environment [4]. The wastewater, discharged from textile industries, 47 contains high concentrations of reactive dyes that exert serious threats 48 49 to sustainability of natural ecosystems [5]. Moreover, the contamination of reactive dyes may present a risk to the aquatic living organisms 50through bioaccumulation thus entering into their food chain. Toxicity 5152of reactive dyes has been reported at concentrations as low as 5.2 mg  $L^{-1}$  [6]. Recently, several methods for dealing with the treat-53 ment of textile wastewater like physico-chemical treatment [5], biolog-5455ical oxidation [7], adsorption and advanced oxidation processes (AOPs), e.g. ozonation, photolysis, electrochemical, and sonolysis [8] have been 5657investigated. Usually, these processes lead to the release of more toxic 58 products than the parent compound that prove fatal for the living

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ABSTRACT

The kinetics of alkaline hydrolysis of malachite green (MG<sup>+</sup>) have been studied spectrophotometrically in the 22 presence of cetyltrimethylammonium bromide (CTAB) under pseudo-first-order condition at buffered of 23 pH 11 and 298 K. The rate increases slightly up to the critical micelle concentration of CTAB then increases 24 rapidly for surface catalysis of the micelles. The reaction has also been studied in the water pools of the CTAB/ 25 1-butanol/heptane/water reverse micelles and found 4–8 times faster over its rate in aqueous phase but the 26 rate decreases exponentially with its water pool (*w*) size. The CTAB micellar medium has been characterized 27 using steady state emission spectroscopy and tensiometry at the reaction condition for better explanation of 28 the experimental findings. The thermodynamic activation parameters for the hydrolysis reaction have also 29 been determined for comparison of the kinetic behavior in different environments. 30

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creatures [9]. Hydrolysis is one of the prime detoxification mechanisms 59 for organic compounds as hydrolysis by-products are normally less 60 toxic to the environment than the parent compound [10]. 61

A number of investigations over the last three decades [11-13] have 62 revealed that the rates of a variety of organic reactions are altered in the 63 presence of dilute aqueous solution of surfactants [14]. One of the most 64 important properties of the aqueous surfactant solution is their ability 65 to solubilize a wide variety of organic molecules, thereby influencing 66 their reactivity. The variation of rate can be explained with the specific 67 interactions between surfactant molecules and reactant species, viz. 68 Coulombic, hydrophobic and charge transfer. All of the interactions 69 can play a significant role in altering the rate of the reaction [15,16]. 70 The surfactant molecules mutate the reaction rate through the forma-71 tion of aggregates, called micelles. These micelles are capable in chang-72 ing the rate of the reaction by several ways [17]-decreasing the entropy 73 of reactant species due to their binding with micelles, relative stabiliza-74 tion or destabilization of the reactants due to their preferential interac-75 tion with surfactants and promoting the approach of reactants towards 76 the micellar core. The catalytic activity of micelles has been studied for 77 different organic reactions, especially the hydrolysis of esters [18], Schiff 78 bases and carbocationic dyes [19]. That means the micellar effects on Q3 the reactivity have frequently been a matter of study. However, a lot 80 of interests have been placed on the study with reverse micelles because 81 of its similarity with the microenvironment in the biological cells 82 [20]. Although the majority of kinetic studies have been carried out in 83 the ternary systems formed by AOT (sodium bis(2-ethylhexyl) 84 sulfosuccinate)/alkane/water microemulsions [21]; nevertheless, only 85 few systematic quantitative studies have been carried out in four 86 component microemulsions [22-24]. Zakharova et al. [25] have shown 87

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the catalytic effect of reverse micellar systems on the alkaline hydrolysis 88 89 of phosphonic acid esters. Microemulsions, also called reverse micelles, are pseudo-heterogeneous mixtures of water-insoluble organic com-90 91pounds, water and a surfactant/co-surfactant mixture. They are transparent, isotropic and thermodynamically stable dispersion systems of 92'oil' and 'water'. Two types of reverse micelles are recognized-oil-in-93 water (o/w) reverse micelles, where the 'oil' resides at the center of 9495the droplet and is surrounded by surfactant and co-surfactant, whereas 96 'water' is at the droplet center in the water-in-oil (w/o) reverse micelles. 97 The size of liquid drop in the dispersed phase is in the range of 8–80 nm. 98 Due to the dynamic nature of reverse micelles, they can exchange their constituents including water, surfactants or other content [26]. The 99 charged head groups of the reverse micelles make the liquid confined 100 101 in it different in their physical properties from that of the bulk phase [27,28]. The liquids, entrapped in a very small cavity of nanometer 102 range, play the fundamental role as a medium that controls the struc-103 tures, functions and thermodynamics of the system which is akin to 104 that of the biological membrane system. One of the most striking 105features of reverse micelles is the "water pool" which is usually 106 expressed by the molar ratio of water to surfact i.e.,  $w = [H_2O]/$ 107 [Surf.] [29–31]. The size of water pool can precisely be controlled by 108 taking different values of 'w'. The parameter 'w' is believed to be the 109 110 key factor in controlling the most important structural features of the reverse micellar system. In most instances, water-in-oil systems (w/o)111 have been used and the water droplets have been proven useful as 112 'minireactors' for various types of reactions. 113

Malachite green (di[4-dimethylaminophenyl]phenyl cation) is an 114 115important water soluble dye. The name of the dye just comes from the similarity of its color. These dyes represent a class of dyes of commercial 116 and analytical importance [32]. The high solubility of the dye in water 117 makes it potential water pollutants. In the textile industry, the removal 118 119 of the dyes from effluents in an economic fashion is a major problem. 120 Since these wastewaters contain substances that modify the medium, and a study of the dyes fading in the presence of surfactants, which 121 are already present in such water, can contribute to the development 122of new decontaminating procedures. The kinetic studies of alkaline 123fading of malachite green thus have extensively been developed [33]. 124

125 In this article, we have presented some of our interesting results on the study of kinetics of alkaline fading of malachite green in micellar 126medium and reverse micellar medium of the cationic surfactant CTAB. 127All of the experimental works have been carried out in buffer medium 128of pH = 11 at 298 K. The reaction of malachite green with hydroxide 129ions is a one-step reaction [34] and follows pseudo-first-order kinetics. 130 Rationalizations of the kinetic results with proper explanations have 131 been attempted. 132

#### 133 2. Experimental

#### 2.1. Reagents & materials 134

Malachite green (MG<sup>+</sup>), CTAB, sodium carbonate and bicarbonate 135136salts have been used as received from Merck, India. Pyrene has been 137 procured from Sigma-Aldrich and used without further purification. N-Cetyl pyridinium chloride (CPC) has been purchased from Merck, 138India and recrystallized from methanol/ethanol mixture before use. 139Highly pure water from Milli-Q Ultra has been used for preparation of 140 141 aqueous solutions throughout the experiment. Fresh aqueous solution of malachite green has been used for day to day works and it is stored 142in a dark place. The concentration of malachite green solution has 04 been determined spectrophotometrically using its molar extinction co-144 efficient,  $\epsilon_{616}\,(MG^+)=148424$  L mol $^{-1}$  cm $^{-1}$  [35]. The aqueous solu-145tion of CTAB and CPC have been prepared with 0.02 mol  $L^{-1}$ 146  $Na_2CO_3 = 0.02 \text{ mol } L^{-1} NaHCO_3$  buffer solution of  $pH = 11 (\pm 0.02)$ . 147 Ethanolic solution of pyrene has been used as a probe. All fluorimetric, 148 tensiometric and kinetic experiments have been carried out at a 149150constant temperature of 298 K and buffer medium of pH = 11.

# 2.2. Methods

### 221 Fluorimetry

Steady-state fluorescence measurements have been performed on a 153 Shimadzu RF-5000 spectrofluorimeter thermostated at 298 K ( $\pm 0.1$  K). 154 For the determination of critical micelle concentration (CMC) and effec- 155 tive dielectric constant ( $\epsilon$ ) on CTAB micellar surface, a pyrene solution of 156 concentration  $1 \times 10^{-6}$  mol L<sup>-1</sup> has been used as a probe. Pyrene has 157 been excited at 332 nm, and its emissions recorded at 375 and 158 386 nm corresponding to the first and third vibronic peaks respectively, 159 with the use of both excitation and emission slits of 5 nm. The formation 160 of micelle has been attributed by the change in the intensity ratios of the 161 first and third peaks  $(I_{\rm I}/I_{\rm III})$  from the emission spectra of pyrene. The 162 aggregation number (N) of CTAB in buffer (pH = 11) medium has 163 been determined using the steady-state fluorescence quenching 164 (SSFQ) method where N-Cetyl pyridinium chloride (CPC) has been 165 employed as guencher (Q). For the determination of N, the concentra- 166 tion of CTAB has been kept well above its CMC throughout the 167 experiments. 168

## 2.2.2. Tensiometry

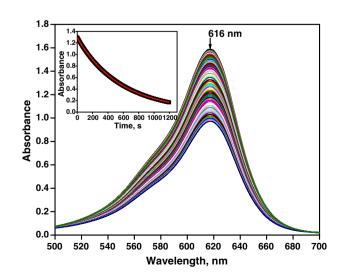
Tensiometric measurement has been carried out on a calibrated du 170 Nouv tensiometer (Jencon, India) employing ring detachment tech- 171 nique [36]. A buffer solution of 10 mL has been placed in a thermostated 172 298 K ( $\pm$ 0.1 K) double walled jacketed glass container where the CTAB 173 solution of desired concentration has been added gradually in a number 174 of steps with the aid of a micro-pipette from Finnpipette. After each ad- 175 dition of surfactant, the mixture has been stirred well using a magnetic 176 stirrer, allowed the mixture to equilibrate and finally the measurement 177 of surface tension ( $\gamma$ ) was carried out subsequently. The measured 178 values of surface tension have been found to be reproducible within 179  $\pm 0.02$  dyn cm<sup>-1</sup>. 180

# 2.2.3. Kinetic measurements

The alkaline hydrolysis reactions have been carried out spectropho- 182 tometrically on a UV-visible Spectrophotometer (Shimadzu, UV-1800) 183 equipped with a Peltier temperature controller, TCC-240A maintaining 184 a constant temperature of 298 K ( $\pm$  0.1 K). The kinetics of hydrolysis re- 185 actions have usually been performed up to 2-3 half-lives of the reaction. 186 The decrease in absorbance (Fig. 1) at 616 nm has been recorded as a 187 function of time. All the kinetic runs have been carried out under 188 pseudo-first-order conditions with excess amount of alkali over MG<sup>+</sup>. 189 The observed pseudo-first-order rate constants  $(k_{obs})$  have been 190

Fig. 1. UV-vis Spectra of MG<sup>+</sup> and <sup>-</sup>OH reaction mixture in aqueous buffer medium at different time intervals. [Inset: Change of absorbance as a function of time at 616 nm.]  $[MG^+]_0 = 5.77 \times 10^{-5} \text{ mol } L^{-1}, \text{ pH} = 11, \mu = 4 \times 10^{-3} \text{ mol } L^{-1} \text{ at } 298 \text{ K}.$ 

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