



# TIF1 $\beta$ is phosphorylated at serine 473 in colorectal tumor cells through p38 mitogen-activated protein kinase as an oxidative defense mechanism



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## ARTICLE INFO

### Article history:

Received 18 August 2017

Accepted 27 August 2017

Available online 31 August 2017

### Keywords:

TIF1 $\beta$  phosphorylation

Chromatin

Oxidative stress

MAPKs

Colorectal cancer

## ABSTRACT

TIF1 $\beta$  is a pleiotropic regulator of a diverse range of cellular processes such as DNA repair or gene repression in stem cells. This functional switch depends on phosphorylation at serine residue 473 and multiple pathways exist to accomplish this. However, the effects of exogenous reactive oxygen species (ROS) generated by bacterial flora and dietary metabolites in the colonic lumen or chemotherapy on TIF1 $\beta$  have not been determined. We report here that exposure of colorectal cancer (CRC) cell lines DLD-1 and HCT116 to hydrogen peroxide specifically induces TIF1 $\beta$  Ser473 phosphorylation. Hydrogen peroxide also induces primarily p38 MAPK and some p42/44 MAPK phosphorylation. Chemical inhibition of p38 MAPK and p42/44 MAPK reduced phosphorylation of TIF1 $\beta$  serine 473 and increased CRC cell death upon peroxide exposure. Taken together, this suggests that it is primarily peroxide-induced p38 MAPK that mediates Ser473 phosphorylation and activation of TIF1 $\beta$  to enable more efficient DNA repair to assist in tumor cell survival against exogenous ROS.

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## 1. Introduction

The protein Transcription Intermediary Factor 1 beta (TIF1 $\beta$ ) is an epigenetic modulator that represses errant gene expression via complexing with heterochromatin protein 1 (HP1), allowing it to interact with the Krüppel-associated box (KRAB) repressor domain found in zinc-finger transcription factors [1]. It is highly expressed in hematopoietic stem cells (as a repressive maintenance factor) [2] as well as several types of cancers [3–5] and high expression of TIF1 $\beta$  is associated with poor prognosis in colorectal cancer patients [6]. A mediator of multiple homeostatic processes, TIF1 $\beta$  has another key function where it influences DNA repair through HP1 proteins [7] and mediates cellular survival after DNA damage in

MCF-7 and HCT116 cells [8]. Whether TIF1 $\beta$  serves as a repressor or a repair modulator seems to be site-dependent, as the phosphorylation site of TIF1 $\beta$  at serine 473 (TIF1 $\beta$ -S473) has been found to regulate TIF1 $\beta$ -HP1 interaction [9] and activate its role in efficient DNA repair and cell survival [10].

Bacterial flora and dietary metabolites produce ROS in the colon and rectum [11] and inadequate antioxidant defense could result in loss of tissue homeostasis and subsequent tumor formation. To this end, several reports have been published that point to ROS involvement in the pathogenesis of colorectal cancer (CRC) [12–14]. The ROS hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) can modify proteins and nucleic acids and it is thought to work at low levels as a signaling molecule regulating cell survival and growth through modulation of transcription factors [15]. However, even if compensatory mechanisms are activated, microsatellite instability from excessive H<sub>2</sub>O<sub>2</sub> would induce DNA damage [16] associated with colorectal cancer development. Therefore, peroxides are implicated in both the regulation of and progression of diseases and the role they play is dependent on their concentration [17,18]. Although the cellular damage caused by ROS in colorectal cancer

*Abbreviations:* TIF1 $\beta$ , Transcription Intermediary Factor 1 beta; ROS, reactive oxygen species; CRC, colorectal cancer; MAPK, mitogen-activated protein kinase; HP1, heterochromatin protein 1.

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has been well studied, less well known is the role of these potential second messengers in regulating gene transcription via phosphorylation of repressor proteins.

In the case of TIF1 $\beta$ , several factors have been reported to induce phosphorylation on the primary activation site Ser473. For example, mitogen-activated protein kinase-activated protein (MAPKAP) kinase 2 (MK2) regulates phosphorylation of TIF1 $\beta$ -S473 in Kaposi's sarcoma-associated herpes virus (KSHV) infection [19]. In early S-phase under normal culture conditions, protein kinase C $\delta$  (PKC $\delta$ ) regulates TIF1 $\beta$ -S473 phosphorylation for cell cycle progression and cellular proliferation [9]. Temporally dynamic TIF1 $\beta$ -S473 phosphorylation has been found upon growth factor stimulation [20]. On the other hand, TIF1 $\beta$ -S473 phosphorylation may also be induced by nutrient depletion in breast cancer [21]. This diverse pool of regulatory elements establishes Serine 473 as the common activation hub for TIF1 $\beta$  to fulfill its roles of chromatin regulation (DNA damage response) and cell survival and growth. However, little is known about the effect of oxidative stress as an additional modulator of phosphorylation on this crucial site and if activation of the DNA repair mechanism by basal or exogenous ROS could upregulate TIF1 $\beta$  to protect tumor cells by increasing efficiency of DNA repair.

In this study, we hypothesized that, at physiologically relevant concentrations, TIF1 $\beta$  phosphorylation will increase, blunting peroxide-induced cell damage. We found that, in fact, H<sub>2</sub>O<sub>2</sub>-induced CRC cell death was intensified in TIF1 $\beta$  knockdown conditions. We then discovered that H<sub>2</sub>O<sub>2</sub> dose induce

phosphorylation of TIF1 $\beta$ -S473. In addition, activation of p38 MAPK and secondary activation of p42/44 MAPK by ROS may regulate TIF1 $\beta$ -S473 phosphorylation in response to oxidative stress to protect CRC cells from ROS-mediated death.

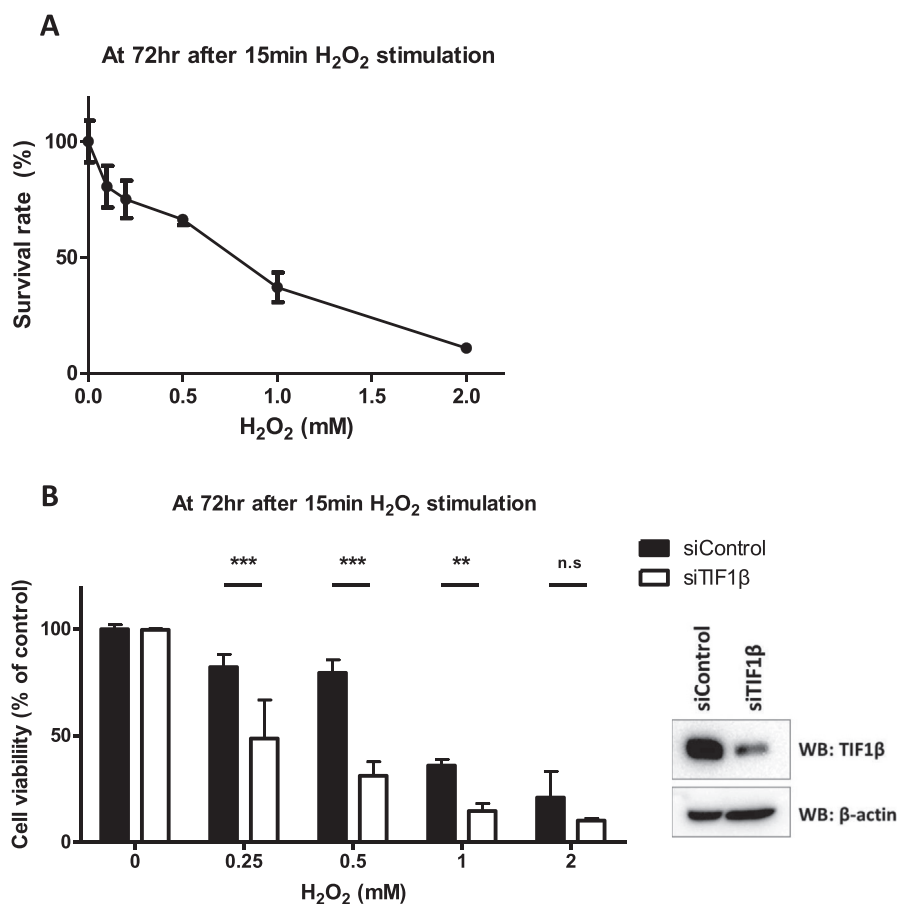
## 2. Material and methods

### 2.1. Cell lines and cell culture

Human colorectal adenocarcinoma HCT116 cells and human embryonic kidney (HEK) 293T cells were maintained in Dulbecco's modified Eagle's medium (DMEM, Invitrogen) supplemented with 10% fetal bovine serum and 1% of a penicillin-streptomycin solution (Gibco) at 37 °C in a 5% CO<sub>2</sub>-humidified atmosphere. Human colorectal adenocarcinoma cells (DLD-1) were cultured in RPMI 1640 medium (Invitrogen) supplemented with 10% fetal bovine serum and 1% of a penicillin-streptomycin solution at 37 °C in a 5% CO<sub>2</sub>-humidified atmosphere. Multiple frozen aliquots were established upon the acquisition and all experimental cells were passaged for fewer than 30 passages after reviving from liquid N<sub>2</sub>. Cells were washed once after cell growth to a subconfluent state and then made quiescent by incubation in serum- and supplement-free medium for 18 h (hr) before further experiments.

### 2.2. H<sub>2</sub>O<sub>2</sub> treatment

A 1 M stock solution of H<sub>2</sub>O<sub>2</sub> was initially made by diluting a 30%



**Fig. 1.** H<sub>2</sub>O<sub>2</sub>-induced cell death is enhanced by TIF1 $\beta$  knockdown in DLD-1 cells. (A) DLD-1 cells were treated for 15 min with differing concentrations of H<sub>2</sub>O<sub>2</sub> as indicated. Cell viability was determined at 72 h after H<sub>2</sub>O<sub>2</sub> treatment by MTT assay. Data are presented as mean  $\pm$  SD, n = 3. (B) DLD-1 cells were transfected with siRNA (control and TIF1 $\beta$ ). After 48 h, cells were challenged with H<sub>2</sub>O<sub>2</sub> for 15 min. Cell viability was determined at 72 h after H<sub>2</sub>O<sub>2</sub> treatment by MTT assay. Data are presented as mean  $\pm$  SD, n = 3. \*\*P < 0.01, \*\*\*P < 0.001. Right panel shows Western blot data indicating total TIF1 $\beta$  after siRNA transfection.

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