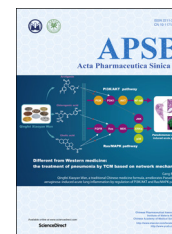




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ORIGINAL ARTICLE

Two new phenylpropanoid glycosides from the aerial parts of *Lespedeza cuneata*

Chuangfeng Zhang^a, Jian Zhou^{a,b}, Jingzhi Yang^a, Chuangjun Li^a,
Jie Ma^a, Dan Zhang^a, Dongming Zhang^{a,*}

^aState Key Laboratory of Bioactive Substance and Function of Natural Medicines, Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100050, China

^bDepartment of Pharmacy, The First Affiliated Hospital of Nanchang University, Nanchang 330006, China

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Cuneataside F

Abstract Two new phenylpropanoid glycosides named cuneataside E (**1**) and cuneataside F (**2**), were isolated from the aerial parts of *Lespedeza cuneata* (Dum. Cours.) G. Don, whose structures were *E* and *Z* isomer, respectively. Their structures were elucidated on the basis of comprehensive spectroscopic analysis (UV, IR, HR-ESI-MS, 1D and 2D NMR). In *in vitro* bioassays at 10 μmol/L, compound **1** showed moderate hepatoprotective activity against *N*-acetyl-*p*-aminophenol (APAP)-induced toxicity in HeG2 cells.

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*Corresponding author.

E-mail address: zhangdm@imm.ac.cn (Dongming Zhang).

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1. Introduction

Lespedeza cuneata (Dum. Cours.) G. Don, an annual herbaceous plant, is distributed in China, Korea, India, Australia and USA¹, which named “ye guan men” in Chinese, is a very important traditional medicine, and has been used in the treatment of diabetes², hematuria, insomnia and malnutrition³. Previous phytochemical studies have revealed flavonoids, sterols, triterpenoids⁴⁻⁶ and phenylpropanoid glycosides⁷ as chemical constituents of the plant, which showed antioxidant effects⁸⁻¹², anti-inflammatory effects¹³ and antibacterial activities¹⁴. Among them, flavonoids were the main components of *L. cuneata*. In our continuing effort in studying constituents from this important medicinal plant, two new phenylpropanoid glycosides (Fig. 1) were isolated. Their structures were elucidated by various spectroscopic methods (UV, IR, HR-ESI-MS, 1D and 2D NMR). The isolation and structural elucidation of the new compounds were described in this paper.

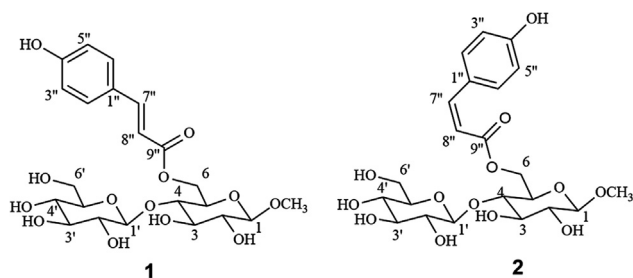


Figure 1 Structures of compounds **1** and **2**.

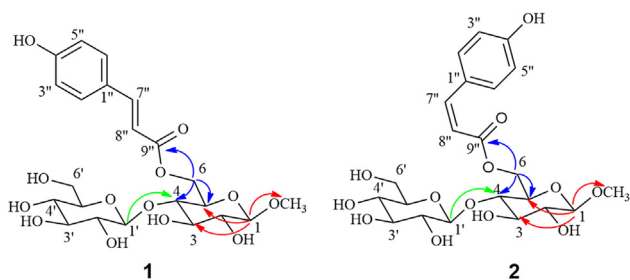


Figure 2 Key HMBC (arrows, from ¹H NMR to ¹³C NMR) correlations of compounds **1** and **2**.

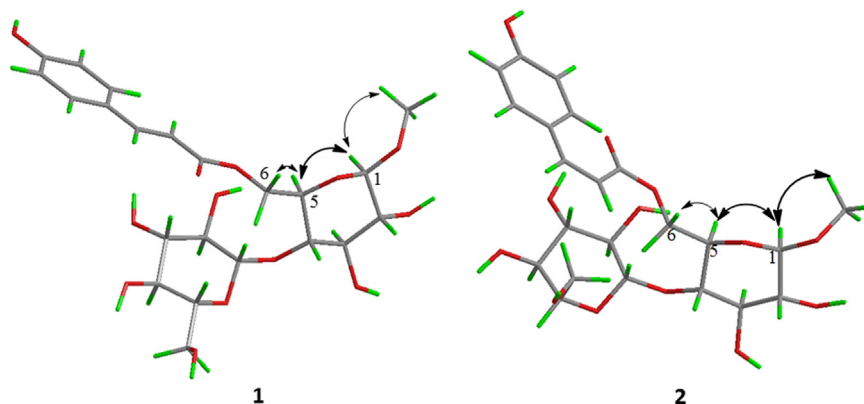


Figure 3 The NOE enhancements induced by irradiation of H-1 and H-6 for compounds **1** and **2**.

2. Results and discussion

Compound **1** was obtained as a white amorphous powder. The UV spectrum showed absorption maximums at 210, 228 and 314 nm. In the IR spectrum, absorption bands at 3375, 2901, 1604, 1515, and 1449 cm^{-1} were observed. These data indicated the presence of hydroxyl, benzene, and carbonyl groups in **1**. The molecular formula was determined to be $\text{C}_{22}\text{H}_{30}\text{O}_{13}$ on the basis of HR-ESI-MS m/z 525.1588 $[\text{M}+\text{Na}]^+$ (Calcd. for $\text{C}_{22}\text{H}_{30}\text{O}_{13}\text{Na}$ 525.1579). In the ¹H NMR spectrum of **1**, a set of AB-type signals at δ_{H} 7.56 (2H, d, $J=8.4$ Hz, H-2'', 6'') and δ_{H} 6.57 (2H, d, $J=8.4$ Hz, H-3'', 5'') were observed, which suggested the existence of a 1,4-disubstituted benzene ring. Additionally, a methoxy signal at δ_{H} 3.35 (3H, s, OMe) and two anomeric proton signals at δ_{H} 4.25 (1H, d, $J=8.0$ Hz), δ_{H} 4.16 (1H, d, $J=8.0$ Hz) with large coupling constants suggested β -glucosidic linkages. From the hydrolysate of **1**, a neutral residue containing sugars was obtained by extraction and evaporation. The sugar residue and authentic D-glucose were separately allowed to react with L-cysteine methyl ester and N-trimethylsilylimidazole (Section 4.4). Subsequent GC analysis indicated that two sugar derivatives from the sugar residue had retention time (t_{R}) identical to that of authentic D-glucose. This verified that both glycosyl units in **1** possessed the D-configuration. We can also find *trans*-disubstituted double bond at δ_{H} 7.55 (1H, d, $J=16.0$ Hz) and δ_{H} 6.43 (1H, d, $J=16.0$ Hz), which suggests that the compound is *E* isomer. The ¹³C NMR spectrum showed 22 carbon signals. An α,β -unsaturated carbonyl group was demonstrated at δ_{C} 166.5. These spectroscopic data indicates that **1** has a *trans-p*-coumaroyl and two β -glucopyranosyl groups, for which the structure was further elucidated by 2D NMR data analysis.

The proton-bearing carbon signals in the NMR spectra were assigned by cross-peaks in the HSQC spectra. HMBC correlations from H-1 to C-3, C-5, C-OCH₃; from H-6 to C-5, C-4, and C-9'' (Fig. 2); together with their chemical shifts, revealed the presence of a methoxy group at C-1 and a *trans-p*-coumaroyl group at C-6. In the NOE spectra (Fig. 3), an enhancement of the proton signal at the H-OCH₃/H-5 on irradiation of the H-1, and at the H-6 revealed that H-OCH₃ and the coumaroyl groups are linked on the same glucopyranosyl moiety. The HMBC correlations from H-1' to C-4 demonstrated that two β -glucopyranosyl groups were connected through a 1,4-linkage. Therefore, the structure of **1** was elucidated as methyl-6-*O*-[(*E*)-3-(4-hydroxyphenyl)prop-2-enyl]-4-*O*- β -D-glucopyranoside- β -D-glucopyranoside, and named cuneataside E.

Compound **2** was obtained as a white powder, whose molecular formula was determined to be $\text{C}_{22}\text{H}_{30}\text{O}_{13}$ on the basis of HR-ESI-MS. The UV spectrum showed absorption maximums at 210, 227 and

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