

Aflatoxin B₁-lysine adduct in dried blood spot samples of animals and humans



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ARTICLE INFO

Article history:

Received 6 September 2016

Received in revised form

30 October 2016

Accepted 1 November 2016

Available online 2 November 2016

Keywords:

Aflatoxin

Dried blood spots

ABSTRACT

Dried blood spots (DBS) were proposed as potentially viable method for exposure assessment of environmental toxicants in infant and young children. For this study, we validated an experimental protocol to quantify AFB₁-lysine adduct in DBS samples of AFB₁-treated F344 rats, as well as samples from human field study. Significant dose-response relationships in AFB₁-lysine adduct formation were found in DBS samples of rats treated with single- and repeated-dose AFB₁. AFB₁-lysine levels in DBS samples were highly correlated with corresponding serum sample levels. The Person coefficients were 0.997 for the single-dose exposure, and 0.996 for the repeated-dose exposure. Levels of AFB₁-lysine adduct had also good agreement between DBS and serum samples as shown by Bland-Altman plot analysis. For human field study samples (n = 36), a Pearson correlation coefficient of 0.784 was found between AFB₁-lysine adduct levels of DBS and corresponding serum samples. Bland-Altman plots showed the distribution of the log differences between DBS and serum AFB₁-lysine levels are within 95% confidence intervals. These results showed AFB₁-lysine adduct levels in DBS cards and serum samples from animals and human samples are comparable, and the DBS technique and analytical protocol is a good means to assess AFB₁ exposure in infant and children populations.

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1. Introduction

Aflatoxins, produced mainly by *Aspergillus flavus* and *Aspergillus parasiticus*, have been one of the most well-known risk factors to liver cancer worldwide, and aflatoxin B₁ (AFB₁) has been classified as group 1 human carcinogen by International Agency for Research on Cancer (IARC, 1993, 2012). Toxic effects of AFB₁ have been widely explored in both animal studies and human epidemiological studies. Acute exposure to high levels of AFB₁ can lead to lethal aflatoxicosis, while chronic low level exposures have been linked with high incidence of hepatocellular carcinoma as well as growth stunting (Asim et al., 2011; Chao et al., 1994; Etzel, 2014; Groopman et al., 1996; Henry et al., 2002; Kensler et al., 2011; Knipstein et al., 2015; Tillett, 2010; Van Rensburg et al., 1985; Wang et al., 1996, 2001). As a ubiquitous contaminant among groundnuts- and corn-based foods, the primary route of exposure has been through

oral consumption of food contaminated with AFB₁, which has been a major food safety issue and public health concern for decades, particularly in developing countries. Studies in recent years have suggested a strong link between childhood stunting, a developmental disorder with various short-term and long-term adverse health outcome, and chronic dietary exposure to mycotoxins, particularly AFB₁ (de Onis and Branca, 2016; Groopman et al., 2014; IARC, 2015; Watson et al., 2015). These increased concerns over AFB₁ exposure and its adverse health effect have placed new demands on the efficiency, viability, and convenience of the exposure assessment methods among the young populations.

Serum AFB₁-lysine adduct is considered to be the most reliable biomarker for assessing chronic exposure to AFB₁. As shown by Fig. 1, this adduct is formed as a part of metabolic pathway of AFB₁. Following absorption, AFB₁ is activated by CYP1A2 and 3A4 in human liver into reactive AFB₁-8,9-epoxides, which subsequently react with DNA to form AFB₁-N⁷-guanine adduct, a highly unstable DNA adduct which underwent depurination readily and are excreted as the guanine base adduct in urine (Guengerich et al., 1998; Kensler et al., 2011). By-products of epoxides can also bind to albumin, the only protein shown to bind AFB₁ on a significant level, with lysine the primary amino acid group for the covalent

Abbreviations: AF, aflatoxins; AFB₁, aflatoxin B₁; AFB₁-lysine, aflatoxin B₁-lysine; CYP450, cytochrome P-450; DMSO, dimethyl sulfoxide; SPE, solid phase extraction.

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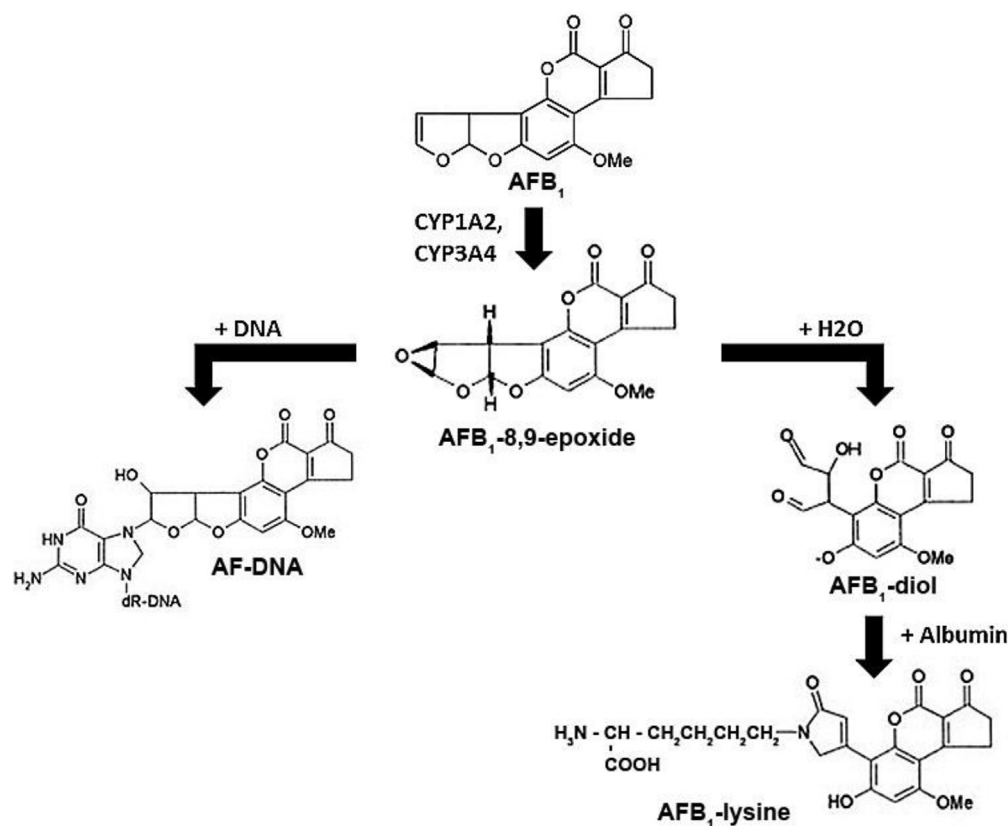


Fig. 1. Metabolic pathway and major DNA and protein adducts of AFB₁. Depicted here including phase I metabolic activation and formation of DNA adduct, AFB₁-N⁷-guanine, and the albumin adduct, AFB₁-lysine.

binding of AFB₁-diol (Sabbioni et al., 1987). Due to the longer half-life as compared to AFB₁-N⁷-guanine, AFB₁-lysine adduct has been considered as a reliable biomarker for assessment of chronic AFB₁ exposure (Kensler et al., 2011). Upon enzymatic digestion, AFB₁-lysine adducts were released from albumin in blood and can be quantified using various methods, including immunoassays, high-performance liquid chromatography (HPLC) with fluorescence detection, and isotope dilution mass spectrometry (IDMS) (McCoy et al., 2008; Sabbioni et al., 1987, 1990). Studies have found significant association between AFB₁ exposure and serum AFB₁-lysine levels in both experimental animals and human studies (Qian et al., 2013; Sabbioni et al., 1990; Wild et al., 1992, 1996). This biomarker has also been used to evaluate various intervention strategies (Groopman et al., 2014; Kang et al., 2015; Kensler et al., 2011; Leroy et al., 2015; Mitchell et al., 2016; Tang et al., 2008; Xu et al., 2010).

Dried blood spots (DBS) have emerged as an attractive alternative to serum sample collection. This technique, popularly used in new-born congenital disease screening first developed by Robert Guthrie (Sharma et al., 2014), was proposed as a potential means for screening biomarkers for exposure assessment and epidemiology, due to its less invasive nature, reduced health hazards, and ease of storage and shipment (Lehmann et al., 2013; McDade, 2014). However, to our knowledge, there are currently few studies on assessing aflatoxin exposure using DBS samples in young populations. The aims for the current study is thus to validate a previously developed method for quantification of AFB₁-lysine levels in DBS samples, using both samples from animals treated with single- and repeated-doses of AFB₁, as well as human DBS samples of known AFB₁ exposure.

2. Materials and methods

2.1. Reagents and chemicals

Unlabelled aflatoxin B₁ (>98% purity), dimethyl sulfoxide (DMSO), albumin determination reagent (bromocresol purple), and normal rat serum were purchased from Sigma Aldrich Chemical Company (St. Louis, MO). ELISA kit for human albumin analysis (ab108788) was obtained from Abcam (Cambridge, MA). Pronase (25 kU, Nuclease-free) was purchased from Calbiochem (La Jolla, CA). Protein assay dye reagent concentrate and protein standards were from Bio-Rad Laboratories Incorporation (Hercules, CA). Mixed mode solid phase extraction (SPE) cartridges were purchased from the Waters Corporation (Milford, MA). All other chemicals and solvents were of highest grade and purity available. Normal human whole blood purchased from Biological Specialty Corporation (Colmar, PA, 18915). AFB₁-lysine standard used in this study was synthesized as previously described by Sabbioni et al. (1987). AFB₁-lysine has been confirmed via both UV spectrophotometry and LC/MS. Confirmation and quantification of AFB₁-lysine standard is established using UV spectrum, where absorbance at 400 nm was used and adjusted with published molar extinction coefficient of 30,866 (Lmol⁻¹cm⁻¹) as described by Sabbioni (1990). The contents are further confirmed via LC/MS, with selective reaction monitoring transition of 457 (m/z)–394 (m/z) for AFB₁-lysine, as described in figure caption of supplemental Fig. 1.

2.2. Sample preparation and extraction

Seven commercial DBS cards from different commercial sources

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