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Letter to the Editor

Photoacoustic microscopy

Optical-acoustic objective

Multispectral imaging

Multispectral photoacoustic microscopy based on an optical-acoustic objective

ABSTRACT

We have developed reflection-mode multispectral photoacoustic microscopy (PAM) based on a novel optical-acoustic objective that integrates a customized ultrasonic transducer and a commercial reflective microscope objective into one solid piece. This technical innovation provides zero chromatic aberration and convenient confocal alignment of the optical excitation and acoustic detection. With a wavelength-tunable optical-parametric-oscillator laser, we have demonstrated multispectral PAM over an ultrabroad spectral range of 270–1300 nm. A near-constant lateral resolution of \sim 2.8 μ m is achieved experimentally. Capitalizing on the consistent performance over the ultraviolet, visible, and near-infrared range, multispectral PAM enables label-free concurrent imaging of cell nucleus (DNA/RNA contrast at 270 nm), blood vessel (hemoglobin contrast at 532 nm), and sebaceous gland (lipid contrast at 1260 nm) at the same spatial scale in a living mouse ear.

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1. Introduction

Photoacoustic microscopy (PAM) [1–3] fills the long-standing gap in high-resolution imaging of endogenous optical absorption contrasts in vivo, among which DNA/RNA [4], hemoglobin [5], and lipid [6,7] are of particular interest. Specifically, the cell nucleus is a critical organelle containing DNA genome, which strongly absorbs the ultraviolet light. Morphological changes in cell nuclei, including enlargement and envelope folding, are considered hallmarks of cancer cells [8]. Hemoglobin, a dominant absorber in the visible spectral range, is the primary oxygen carrier in the blood circulation. Angiogenesis [9,10] and hypoxia [11], which can be respectively revealed by the distribution and oxygen saturation of hemoglobin, are also core hallmarks of cancer [12]. Lipid forms a diverse group of infrared-absorbing molecules that play important roles at cellular and organismal levels [13]. Aberrant lipid metabolism is an established hallmark of cancer cells [14]. Concurrent imaging of the multiple endogenous optical absorbers at the same spatial scale holds great promise for both basic and translational cancer research. However, multispectral PAM that spans from ultraviolet to near-infrared is complicated by the chromatic aberration of the optics and not currently available.

To address this unmet challenge, we have developed a novel multispectral PAM system. Specifically, a commercial reflective microscope objective with zero chromatic aberration is employed to achieve consistent optical focusing over a broad spectral range. A customized ultrasonic transducer is attached to the dark zone of the reflective objective for convenient confocal alignment of the optical excitation and acoustic detection in reflection mode, which avoids the optical aberration and acoustic loss induced by the otherwise needed optical–acoustic beam combiners in conventional PAM systems [1,2,15,16]. The ring-shaped transducer design

provides an alternative means for the confocal alignment without a combiner, but at the expense of detection sensitivity due to the central opening [17,18]. Transmission-mode PAM [19,20] can be readily extended for multispectral measurements by using the aberration-free reflective objective; however, its application is limited by the poor accessibility to anatomical sites *in vivo*.

With a high-repetition-rate wavelength-tunable optical parametric oscillator (OPO) laser, our multispectral PAM covers an unprecedented spectral range of 270–1300 nm. A near-constant lateral resolution of ~2.8 μ m is achieved experimentally. Capitalizing on the ultrabroad spectral coverage and consistent spatial resolution, we have demonstrated concurrent PAM of cell nuclei (DNA/RNA contrast at 270 nm), blood vessel (hemoglobin contrast at 532 nm), and sebaceous gland (lipid contrast at 1260 nm) at the same spatial scale in a living mouse ear.

2. Materials and methods

2.1. Optical-acoustic objective

As shown in Fig. 1A, the optical–acoustic objective consists of a commercial reflective microscope objective (also known as Schwarzschild objective; LMM-15X-UVV, Thorlabs) and a customized ultrasonic transducer. The two hemispherical mirrors of the reflective microscope objective are coated with ultravioletenhanced aluminum, which assures zero chromatic aberration over a broad spectral range (200 nm–20 μ m). The convex primary mirror causes an obscuration in the center of the imaging system. Thus, the laser beam through this reflective objective is solid only at the focus and donut-shaped everywhere else. The ultrasonic transducer is attached to the back surface of the primary mirror, which is directly below the entrance pupil of the reflective

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Keywords:

Cell nucleus

Blood vessel

Lipid





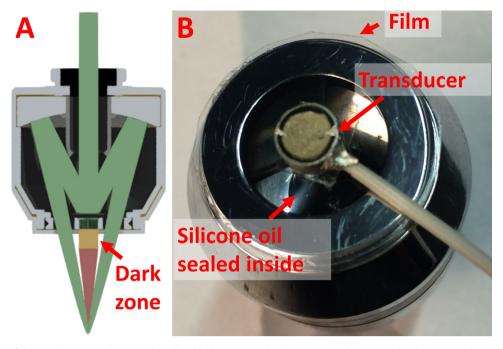


Fig. 1. (A) Sectional view of the optical-acoustic objective. The yellow block represents the ultrasonic transducer. Optical and acoustic paths are labeled in green and red, respectively. (B) Photograph of the objective showing the liquid-filling feature.

microscope objective. Positioning the transducer in the optically dark zone allows convenient alignment of the optical and acoustic foci with no interference. To ensure optimal superposition of the optical and acoustic foci, the focal length of the piezoelectric ceramic piston transducer is carefully designed and the transducer location fine tuned before permanent attachment to the reflective microscope objective. The elegant confocal alignment and easy coupling of acoustic energy to the transducer make the optical– acoustic objective ideally suited for reflection-mode PAM.

For acoustic coupling, the optical-acoustic objective needs to be immersed in a transparent liquid. The mismatch of optical refractive index at the interface between the liquid and the air cavity of the objective induces significant optical aberration [21], particularly during mechanical scan when the liquid surface is unstable. To address this issue, the objective is filled with the same liquid and sealed with a thin film (OCA8146-2, Thorlabs), which is optically transparent over 270-2000 nm (Fig. 1B). Silicone oil, the liquid we use, is non-absorbing and commonly used in oilimmersion microscope objective (e.g. UPLSAPO30XSIR, Olympus). To avoid the lensing effect induced by the surface tension of silicone oil, the entrance pupil of the objective is also sealed with a fused-silica broadband optical window (WG41050, Thorlabs). This liquid-filling feature clearly distinguishes our design from the previous PAM system based on a reflective microscope objective [21].

2.2. Multispectral PAM system

As shown in Fig. 2, our multispectral PAM system employs a wavelength-tunable OPO laser (NT242, Ekspla; wavelength coverage: 210–2600 nm; repetition rate: 1 kHz). Due to the non-uniform beam shape across the broad spectral range, the laser output is split by a flip mirror (FM; TRF90, Thorlabs) and a dichroic mirror (DM; DMLP650, Thorlabs) into three paths—ultraviolet (purple), visible (green), and near-infrared (red)—for beam reshaping. The individually reshaped and expanded beams are combined *via* another identical pair of FM and DM, spatially filtered by an iris with an 8-mm aperture (ID25, Thorlabs),

reflected by three fused-silica broadband right-angle prisms (RP; PS611, Thorlabs), and focused by the optical-acoustic objective for multispectral photoacoustic excitation. The ratios of the pulse energies after and before beam reshaping and filtering are 80.9%, 64.3%, and 60.8% for the ultraviolet, visible, and near-infrared paths, respectively. The objective is immersed in an oil tank filled with silicone oil for acoustic coupling. The bottom of tank is sealed with a thin layer of transparent polyethylene membrane to expose the object to be imaged. Commercial ultrasound gel (Aquasonic CLEAR[®], Parker Laboratories) is sandwiched between the membrane and object for acoustic coupling. The oil tank and object holder are mounted on two motorized linear stages (PLS-85, PI micos) for two-dimensional raster scanning.

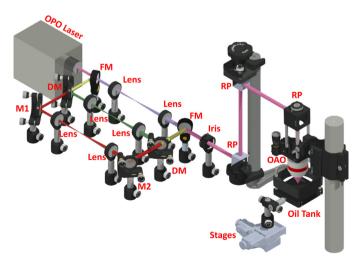


Fig. 2. Schematic of multispectral PAM. The ultraviolet, visible, and near-infrared paths are labeled in purple, green, and red, respectively. The visible and near-infrared combined path is labeled in yellow. The combined path of all three spectral ranges is labeled in pink. OPO, optical parametric oscillator; FM, flip mirror; DM, dichroic mirror; M1 and M2, mirrors; RP, right-angle prism; OAO, optical-acoustic objective.

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