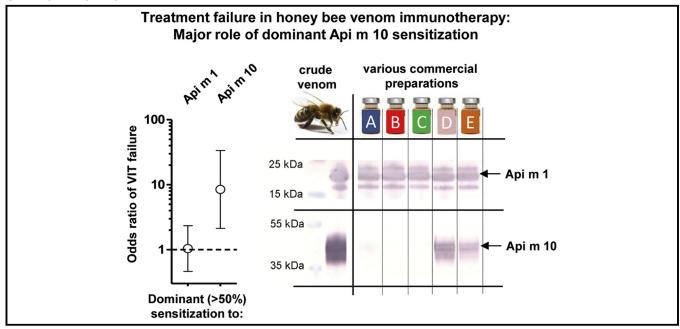
# Predominant Api m 10 sensitization as risk factor for treatment failure in honey bee venom immunotherapy



Marcel Frick, MS, a,b\* Jörg Fischer, MD,c\* Arthur Helbling, MD,d Franziska Ruëff, MD,e Dorothea Wieczorek, MD,f Markus Ollert, MD,g,h Wolfgang Pfützner, MD,i Sabine Müller, MD,b Johannes Huss-Marp, MD, MBA,b Britta Dorn,a,b Tilo Biedermann, MD,c,j Jonas Lidholm, PhD,k Gerta Ruecker, PhD,I Frank Bantleon, PhD,m Michaela Miehe, PhD,m Edzard Spillner, PhD,m‡ and Thilo Jakob, MDa,b‡ Gieβen, Freiburg, Tübingen, Munich, Hannover, and Marburg, Germany; Bern, Switzerland; Esch-sur-Alzette, Luxembourg; Odense and Aarhus, Denmark; and Uppsala, Sweden

#### **GRAPHICAL ABSTRACT**



From athe Department of Dermatology and Allergology, University Medical Center Gießen-Marburg, Justus Liebig University, Gießen; bthe Allergy Research Group, Department of Dermatology, Medical Center, University of Freiburg, Freiburg; cthe Department of Dermatology and Allergology, University Medical Center Tübingen, Tübingen; dthe Department of Internal Medicine, Spital Netz Bern, Allergy Unit Zieglerspital, Bern; ethe Department of Dermatology and Allergology, Ludwig-Maximilian-University Munich, Munich; <sup>f</sup>Department of Dermatology and Allergy, Hannover Medical School, Hannover; <sup>g</sup>Department of Infection and Immunity, Luxembourg Institute of Health (LIH), Esch-sur-Alzette; hthe Department of Dermatology and Allergy Center, Odense Research Center for Anaphylaxis, University of Southern Denmark, Odense; ithe Department of Dermatology and Allergology, University Medical Center Gießen-Marburg, Philipps University, Marburg; <sup>j</sup>the Department of Dermatology and Allergology, Technical University Munich, Munich; <sup>k</sup>Thermo Fisher Scientific, Uppsala; 1the Institute for Medical Biometry and Statistics, Medical Center -University of Freiburg, Freiburg; and mthe Department of Engineering, Aarhus University, Immunological Engineering, Aarhus.

\*These authors contributed equally to this work.

‡These authors contributed equally to this work.

This work was supported in part by a research grant of Thermo Fisher Scientific, Freiburg, to T.J.

Disclosure of potential conflict of interest: F. Ruëff has worked as an adviser for ALK Abelló Arzneimittel GmbH, Bencard, DST GmbH, and Dr. Gerhard Mann chempharm. Fabrik GmbH; has received speaker's honorarium from ALK-Abelló Arzneimittel GmbH, Astra Zeneca, Bencard, Novartis Pharma GmbH, HAL, MEDA Pharma GmbH & Co, Semperit Technische Produkte Gesellschaft m.b.H, and Stallergenes GmbH; has received consultancy fees from Novartis, HAL, and ALK-Abelló; and has received research support from Novartis. M. Ollert has received

consultancy fees from Siemens Healthcare and Hitachi Chemical Diagnostics; has received lecture fees from ThermoFisher-Phadia and Siemens Healthcare; has stock/ stock options in Siemens Healthcare; and is Scientific Cofounder of the University biotech spin-off enterprise Protein-Ligand-Structural-Design (PLS-Design) GmbH, Hamburg, Germany (2004). W. Pfützner has received consultancy fees from ALK-Abelló as an advisory board member; has received research support from the Deutsche Forschungsgemeinschaft (DFG), Philipps University Marburg, and the University Clinic of Marburg; has received lecture fees from ALK-Abelló and Novartis; has received research support from Phadia/Thermo Fisher Scientific, Freiburg, Germany. S. Müller has received lecture fees from Novartis, ALK-Abelló, and Bencard. J. Huss-Marp has been employed by, has received lecture fees from, and has stock/stock options in Phadia GmbH, ThermoFisher Scientific. T. Biedermann has received research funding and speaker's honorarium from Phadia/Thermo Fisher Scientific, Freiburg; has received consultancy fees from Phadia and ThermoFisher; has received research support from DFG. Novartis, Phadia, and ThermoFisher; and has received lecture fees from MSD, Novartis, HIPP GmbH & Co. ALK-Abelló Arzneimittel GmbH, MedComms Ltd, and Astellas Pharma GmbH. J. Lidholm is an employee of Thermo Fisher Scientific, manufacturer of the assay system used in the study. E. Spillner is cofounder of PLS-Design GmbH, Hamburg, Germany. T. Jakob has received research support, consultancy fees, and travel support from Thermo Fisher Scientific; is on the Germany Society of Allergy and Clinical Immunology Board; has received consultancy fees from ALK-Abelló, Allergy Therapeutics, Novartis, Leti GmbH, Stallergenes, and Allergopharma; is employed as Editor for Allergó Journal International, Springer, Germany; and has received lecture fees from ALK-Abello, Allergy Therapeutics, Novartis, Allergopharma, Stallergenes, and Thermo Fisher Scientific. The rest of the authors declare that they have no relevant conflicts of interest.

1664 FRICK ET AL

J ALLERGY CLIN IMMUNOL

DECEMBER 2016

Background: Component resolution recently identified distinct sensitization profiles in honey bee venom (HBV) allergy, some of which were dominated by specific IgE to Api m 3 and/or Api m 10, which have been reported to be underrepresented in therapeutic HBV preparations.

Objective: We performed a retrospective analysis of component-resolved sensitization profiles in HBV-allergic patients and association with treatment outcome.

Methods: HBV-allergic patients who had undergone controlled honey bee sting challenge after at least 6 months of HBV immunotherapy (n = 115) were included and classified as responder (n = 79) or treatment failure (n = 36) on the basis of absence or presence of systemic allergic reactions upon sting challenge. IgE reactivity to a panel of HBV allergens was analyzed in sera obtained before immunotherapy and before sting challenge.

Results: No differences were observed between responders and nonresponders regarding levels of IgE sensitization to Api m 1, Api m 2, Api m 3, and Api m 5. In contrast, Api m 10 specific IgE was moderately but significantly increased in nonresponders. Predominant Api m 10 sensitization (>50% of specific IgE to HBV) was the best discriminator (specificity, 95%; sensitivity, 25%) with an odds ratio of 8.444 (2.127-33.53; P = .0013) for treatment failure. Some but not all therapeutic HBV preparations displayed a lack of Api m 10, whereas Api m 1 and Api m 3 immunoreactivity was comparable to that of crude HBV. In line with this, significant Api m 10 sIgG<sub>4</sub> induction was observed only in those patients who were treated with HBV in which Api m 10 was detectable. Conclusions: Component-resolved sensitization profiles in HBV allowed suggests predominant IgE consistization to Api m 10 as a

Conclusions: Component-resolved sensitization profiles in HBV allergy suggest predominant IgE sensitization to Api m 10 as a risk factor for treatment failure in HBV immunotherapy. (J Allergy Clin Immunol 2016;138:1663-71.)

**Key words:** Apis mellifera, Hymenoptera venom allergy, HBV allergy, recombinant allergen, allergen-specific immunotherapy, treatment failure

Systemic allergic reaction to Hymenoptera stings affects 0.3% to 3.5% of the adult population. <sup>1,2</sup> Venom immunotherapy (VIT) protects allergic patients from systemic reactions to subsequent stings. <sup>2,3</sup> The effectiveness of VIT depends on a number of variables such as treatment duration, venom dose during maintenance therapy, and type of venom (honey bee [HB] vs vespid) used for immunotherapy. <sup>4-6</sup> Treatment failure is more frequent in HB VIT than in vespid VIT, ranging from 11% to 23% as compared with 0% to 9%. <sup>4-7</sup> A recent retrospective study on the outcome of more than 1600 sting challenges calculated an odds ratio (OR) of more than 5 for treatment failure in honey bee venom (HBV) allergy as compared with VIT in vespid venom allergy. <sup>6</sup> This increased risk of treatment failure in HBV allergy has been suggested to be

Abbreviations used

AUC: Area under the ROC curve

HBV: Honey bee venom

HB: Honey bee

LOD: Limit of detection

NLR: Negative likelihood ratio

OR: Odds ratio

PLR: Positive likelihood ratio

ROC: Receiver operating characteristic

sIgE: Specific IgE

VIT: Venom immunotherapy

associated with differences in venom composition, venom dose during natural exposure conditions, and differences in sensitization profiles. 4.7.8

Advances in proteomics and molecular biology have allowed a detailed characterization of the protein composition of HBV. The best-characterized HBV allergens are phospholipase A2 (Api m 1), hyaluronidase (Api m 2), and the basic peptide melittin (Api m 4).<sup>9,10</sup> Additional HBV allergens of lower abundance have been cloned and characterized such as acid phosphatase (Api m 3),<sup>11</sup> dipeptidylpeptidase IV (Api m 5),<sup>12</sup> icarapin (Api m 10), <sup>13,14</sup> and others as recently reviewed. <sup>15</sup> Analysis of different venom preparations have shown that Api m 3 and Api m 10, while present in the crude HBV, are absent or underrepresented in preparations used for HBV immunotherapy. 13 These findings were supported by subsequent observations that in patients with dominant sensitization to Api m 10, IgE reactivity to HBV could be inhibited by crude HBV preparations but not by therapeutic HBV preparations.<sup>8</sup> In addition, HBV-allergic patients who had undergone VIT displayed a strong induction of sIgG<sub>4</sub> to Api m 1, Api m 2, and Api m 4, whereas no or little induction of sIgG<sub>4</sub> to Api m 3 and Api m 10 could be detected.<sup>8</sup> On the basis of these 3 lines of evidence, we hypothesized that the absence or underrepresentation of Api m 3 and Api m 10 in therapeutic HBV preparations may have an impact on the treatment outcome of VIT and that distinct sensitization profiles, for example, with predominant IgE reactivity to Api m 3 and/or Api m 10, may represent a potential risk factor for treatment failure of VIT in HBV allergy. To address this issue, we here retrospectively analyzed the molecular sensitization profiles in HBVallergic patients who had undergone controlled HB sting challenge after at least 6 months of HBV.

## **METHODS**

#### **Patients**

Sera from HBV-allergic patients who had undergone controlled HB sting challenge after at least 6 months of HBV immunotherapy at a maintenance dose of  $100 \mu g$  BV were included in the study (n = 115) and classified as responder,

Received for publication February 7, 2016; Revised April 11, 2016; Accepted for publication April 22, 2016.

Available online May 24, 2016.

0091-6749

Corresponding author: Thilo Jakob, MD, Department of Dermatology and Allergology, University Medical Center Gießen-Marburg, Justus Liebig University Gießen, Gaffkystr. 14, 35395 Gießen, Germany. E-mail: thilo.jakob@derma.med.uni-giessen.de.

The CrossMark symbol notifies online readers when updates have been made to the article such as errata or minor corrections

<sup>© 2016</sup> The Authors. Published by Elsevier Inc. on behalf of the American Academy of Allergy, Asthma & Immunology. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/). http://dx.doi.org/10.1016/j.jaci.2016.04.024

### Download English Version:

# https://daneshyari.com/en/article/5646420

Download Persian Version:

https://daneshyari.com/article/5646420

<u>Daneshyari.com</u>