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# Modulation of the NF-κB signaling pathway by the HIV-2 envelope glycoprotein and its incomplete BST-2 antagonism



François E. Dufrasne<sup>a,\*</sup>, Mara Lucchetti<sup>a</sup>, Anandi Martin<sup>a</sup>, Emmanuel André<sup>a,b</sup>, Géraldine Dessilly<sup>a</sup>, Benoit Kabamba<sup>a,b</sup>, Patrick Goubau<sup>a</sup>, Jean Ruelle<sup>a</sup>

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#### ABSTRACT

The HIVs have evolved by selecting means to hijack numerous host cellular factors. HIVs exploit the transcription factor NF- $\kappa$ B to ensure efficient LTR-driven gene transcription. However, NF- $\kappa$ B is primarily known to act as a key regulator of the proinflammatory and antiviral responses. Interestingly, retroviruses activate NF- $\kappa$ B during early stages of infection to initiate proviral genome expression while suppressing it at later stages to restrain expression of antiviral genes. During HIV-1 infection, diverse viral proteins such as Env, Nef and Vpr have been proposed to activate NF- $\kappa$ B activity, whereas Vpu has been shown to inhibit NF- $\kappa$ B activation. It is still unclear how HIV-2 regulates NF- $\kappa$ B signaling pathway during its replication cycle. Here we confirm that human BST-2 and HIV-1 Env proteins can trigger potent activation of NF- $\kappa$ B. Importantly, we demonstrate for the first time that the HIV-2 Env induces NF- $\kappa$ B activation in HEK293T cells. Furthermore, the anti-BST-2 activity of the HIV-2 Env is not sufficient to completely inhibit NF- $\kappa$ B activity.

#### 1. Introduction

The transcription factor NF-κB plays pivotal roles in the innate immune and antiviral responses by inducing expression of proinflammatory cytokines, type I interferons (IFN-α and IFN-β), interferonstimulated genes (ISGs) as well as expression of genes involved in cell growth and survival (Chan and Greene, 2012; Gosh and Hayden, 2012; Sen and Baltimore, 1986). In absence of stimuli, NF-κB is maintained in an inactive form in the cell cytoplasm by the inhibitors of κB (IκBs). A variety of stimuli promote degradation of IkB proteins: viral antigens, cytokines and specific cell surface receptors such as interleukin 1 receptor, IFN receptors, T cell TCR-CD3 complex, Toll-like receptors (TLRs) or tumor necrosis factor (TNF) receptors (Chan and Greene, 2012; Pfeffer, 2011; Vallabhapurapu and Karin, 2009). Furthermore, human BST-2/Tetherin acts as an innate sensor of viral assembly and budding. A dityrosine motif (Y<sub>6</sub>xY<sub>8</sub>) in the BST-2 cytoplasmic tail recruits Syk kinase to prime the NF-kB signaling pathway (Arias and Evans, 2014; Cocka and Bates, 2012; Galao et al., 2012, 2014; Tokarev et al., 2013). The induction of the NF-κB signaling pathway requires activation of the IKK complex (IKKα, IKKβ and NEMO subunits). This complex phosphorylates IkB regulatory domains, thus allowing its ubiquitination and degradation. Consequently, NF-κB is translocated into the cell nucleus and binds to specific DNA sequences located in the promoters of NF-κB-dependent target genes (Chan and Greene, 2012; Heusinger and Kirchhoff, 2017; Kanarek and Ben-Neriah, 2012; Pfeffer, 2011; Vallabhapurapu and Karin, 2009). Interestingly, most of the HIV long terminal repeats (LTRs) include kB binding sequences, typically one in HIV-2 and two or three in HIV-1 (Bachu et al., 2012; Chen-Park et al., 2002; Hiebenthal-Millow et al., 2004; Jeeninga et al., 2000; Stroud et al., 2009). Although NF-kB is a key regulator of the host immune and antiviral responses, it is manipulated by retroviruses for promoting effective transcription of viral genes (Chan and Greene, 2012; Hiscott et al., 2001). However, even though NF-κB activation is beneficial for initial viral replication, this leads to rapid antiviral immune responses against the retroviruses. HIVs overcome this challenge by activating NF-κB at early stages of the viral cycle while inhibiting this transcription factor at later stages, restraining therefore expression of various antiviral genes (Heusinger and Kirchhoff, 2017; Hiscott et al., 2001).

While many studies attempted to identify the proteins used by HIV

a Université catholique de Louvain, Experimental and Clinical Research Institute (IREC), Medical Microbiology Unit (MBLG), AIDS Reference Laboratory, Avenue Hippocrate 54, B-1200 Brussels, Belgium

b Cliniques Universitaires Saint-Luc, Clinical Biology Department, Microbiology Unit, B-1200 Brussels, Belgium

Abbreviations: HIV, human immunodeficiency virus; Env, envelope glycoprotein; BST-2, Bone Marrow Stromal Cell Antigen 2; NF-κB, nuclear factor kappa-light-chain-enhancer of activated B cells; IκBs, inhibitors of κB proteins; IKKs, IκB kinases

<sup>\*</sup> Corresponding author.

E-mail addresses: francois.dufrasne@uclouvain.be (F.E. Dufrasne), anandi.martin@uclouvain.be (A. Martin), emmanuel.andre@uclouvain.be (E. André), geraldine.dessilly@uclouvain.be (G. Dessilly), benoit.kabamba@uclouvain.be (B. Kabamba), patrick.goubau@uclouvain.be (P. Goubau), jean.ruelle@uclouvain.be (J. Ruelle).

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to fine-tune NF- $\kappa$ B activity throughout the replication cycle, it is still unclear and controversial if the HIV-1 envelope glycoprotein (Postler and Desrosiers, 2012), the HIV-1 and HIV-2 Nef proteins (Mangino et al., 2011; Sauter et al., 2015) or the Vpr proteins (Liang et al., 2015; Liu et al., 2013; Roux et al., 2000) activate the NF- $\kappa$ B signaling pathway. At later stages of infection, it is well defined that the HIV-1 Vpu protein - which is not encoded by HIV-2 - inhibits the NF- $\kappa$ B activity by three means: internalization and degradation of BST-2/Tetherin (Galao et al., 2012; Tokarev et al., 2013), impeding the I $\kappa$ B degradation through sequestration of  $\beta$ -TrCP proteins (Akari et al., 2001; Bour et al., 2001) and blocking the NF- $\kappa$ B nuclear translocation (Sauter et al., 2015).

To date, no study investigated neither the effects of the HIV-2 Env protein in the NF- $\kappa$ B activation, nor the impacts of the Env-mediated anti-BST-2 antagonism in the modulation of the NF- $\kappa$ B signaling pathway. In the present study, we tested the potential impacts of HIV-2 Env in the NF- $\kappa$ B activity.

#### 2. Results

#### 2.1. The HIV-2 envelope glycoprotein induces NF-κB activation

Since it has been described that the cytoplasmic domain of the HIV-1 glycoprotein gp41 enhances NF- $\kappa$ B activation (Postler and Desrosiers, 2012), we first sought to determine whether the HIV-2 envelope glycoprotein was capable of inducing the NF- $\kappa$ B pathway. To test this hypothesis, we generated a derivative of HEK293T cells stably expressing firefly luciferase gene under the control of an NF- $\kappa$ B-dependent promoter (HEK293T-NF- $\kappa$ B cells). Transfections of the transduced cells with plasmids encoding viral Env or Nef proteins of either HIV-1 or HIV-2, followed by a luciferase reporter assay allowed to assess the effect of these proteins on NF- $\kappa$ B activity. We observed that HIV-1 Env activated NF- $\kappa$ B  $\sim$  12- to 20-fold compared to the negative control (Fig. 1A). These results confirmed that the HIV-1 Env protein stimulates NF- $\kappa$ B activity in HEK293T cells, in agreement with published data (Postler and Desrosiers, 2012). Interestingly, expression of HIV-2 Env protein potently enhanced NF- $\kappa$ B activation with a  $\sim$  17- to 28-fold

increase compared to the negative control (Fig. 1A). Therefore, we showed for the first time that the HIV-2 envelope glycoprotein may activate the NF- $\kappa$ B signaling pathway in HEK293T cells.

A recent study conducted by Sauter et al. (2015) assigned the NF-κB activation ability to the viral accessory Nef protein in both HIV-1 and HIV-2. We partly reproduced these experiments and we tested the potential activation of NF-κB pathway by the Nef proteins. Transfections with plasmids encoding HIV-1 or HIV-2 Nef both showed a weak and non-significant induction of NF-κB activity, as previously reported (Fig. 1A) (Heusinger and Kirchhoff, 2017; Sauter et al., 2015).

As previously mentioned, human BST-2 can activate the NF- $\kappa$ B pathway, triggering thereby proinflammatory responses. This restriction factor acts as an innate immune sensor of the assembly and budding of mammalian enveloped virions (Cocka and Bates, 2012; Galao et al., 2012, 2014; Tokarev et al., 2009, 2013). In our experiments, overexpression of BST-2 indicated, as expected, that this protein induced the NF- $\kappa$ B signaling pathway (Fig. 1A).

To validate the specificity of the NF- $\kappa$ B pathway activation by BST-2 and HIV-2 Env proteins, we used a dominant negative form of IKK $\beta$ . Dominant negative IKK $\beta$  (DN-IKK $\beta$ ) is unable to phosphorylate the IkB protein, therefore preventing its proteasomal degradation and nuclear translocation of NF- $\kappa$ B subunits. Thus, NF- $\kappa$ B is consistently maintained in its inactive form in the cytoplasmic compartment (Cocka and Bates, 2012). Firstly, co-transfection of expression vectors encoding BST-2 and DN-IKK $\beta$  revealed an extensive and significant inhibition of NF- $\kappa$ B activity initially induced by BST-2. Secondly, co-transfection of plasmids encoding HIV-2 Env and DN-IKK $\beta$  showed a similar inhibition (Fig. 1B). These results demonstrated that the HIV-2 Env-mediated NF- $\kappa$ B activation observed was specific of the NF- $\kappa$ B pathway and ruled out the potential involvement of another transcription factor.

## 2.2. Differential modulation of the NF- $\kappa B$ activation by the anti-BST-2 HIV-1 Vpu and HIV-2 Env

HIV-1 utilizes the accessory Vpu protein to overcome BST-2 host restriction factor by removing BST-2 from the cell surface and facilitates therefore the budding of nascent virions from infected cells (Dube et al.,

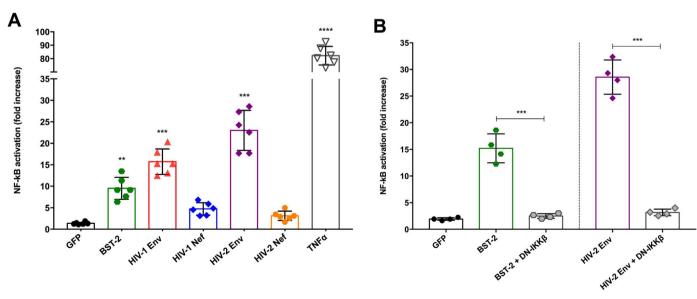


Fig. 1. Stimulation of NF-κB activity by human BST-2 and lentiviral proteins. (A) Transduced HEK293T cells were transfected with expression vectors encoding GFP (as a negative control), human BST-2 or viral proteins of HIV-1 and HIV-2 (Env or Nef proteins). HEK293T cells were lysed and luciferase activities were determined 24 h post transfection. As a positive control, TNFα was added into the cell culture medium (25 ng/ml) to induce a potent activation of NF-κB pathway (Chan and Greene, 2012). (B) Specificity of the NF-κB pathway activation by BST-2 and HIV-2 Env proteins. Transduced HEK293T cells were co-transfected with BST-2 or HIV-2 Env plasmids alongside with a dominant negative form of IKKβ (DN-IKKβ). NF-κB activity is represented as fold increase, being the ratio between the luciferase activity (RLU) of the corresponding measure and that of transduced but non-transfected HEK293T cells. These results were derivated from six independent experiments (n = 6) in (A) and four independent experiments (n = 4) in (B). Data were analyzed by one-way ANOVA statistical test followed by a multiple comparison test which compares each mean with that of the negative control (GFP) in (A) or compares each mean with all other means in (B) (\*\* p-value < 0.01, \*\*\* < 0.001 and \*\*\*\* < 0.0001). Error bars indicate mean  $\pm$  standard deviation (SD).

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