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# The molecular impact of omega 3 fatty acids on hepatic pro-inflammatory cytokine signaling



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#### ABSTRACT

*Purpose:* Parenteral nutrition associated liver disease (PNALD) develops in a subset of children receiving parenteral nutrition for intestinal failure. Omegaven<sup>TM</sup> is an omega-3 fatty acid ( $\Omega$ 3FA) lipid emulsion high in docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) that can lessen PNALD. Inflammatory cytokines (IL-1, TNF- $\alpha$ , TGF- $\beta$ ) are elevated in PNALD and can decrease paraoxonase 1 protein expression (PON1). We sought to determine the effect of Omegaven<sup>TM</sup>, EPA, and DHA on inflammatory cytokines TNF- $\alpha$ , IL-1, and TGF- $\beta$  via ERK1/2 and p-Smad2/3 signaling pathways as well as the changes in intracellular PON1 protein expression as a potential mechanism explaining the protective effects of Omegaven<sup>TM</sup> and  $\Omega$ 3FA.

*Methods*: HepG2 cells were cultured with each cytokine and Omegaven<sup>™</sup>, or EPA and DHA, or Intralipid<sup>™</sup>. P-Smad2/3 and PON1 protein levels were measured by Western blotting. ERK1/2 signaling was studied using homogenous time resolved fluorescence.

*Results:* Omegaven<sup>TM</sup> decreased TGF- $\beta$  mediated Smad2/3 signaling by 30% (70% of control  $\pm$  12, p < 0.03). Omegaven<sup>TM</sup> decreased IL-1 and TNF- $\alpha$  mediated ERK1/2 signaling (0.49 fold  $\pm$  0.09, p < 0.05 and 0.22  $\pm$  0.05, p < 0.05) compared to control.

*Conclusion:* Our results describe potential mechanisms by which  $\Omega$  and  $\Omega$  3FA can be hepatoprotective in the setting of PNALD by abating inflammatory cytokine signaling.

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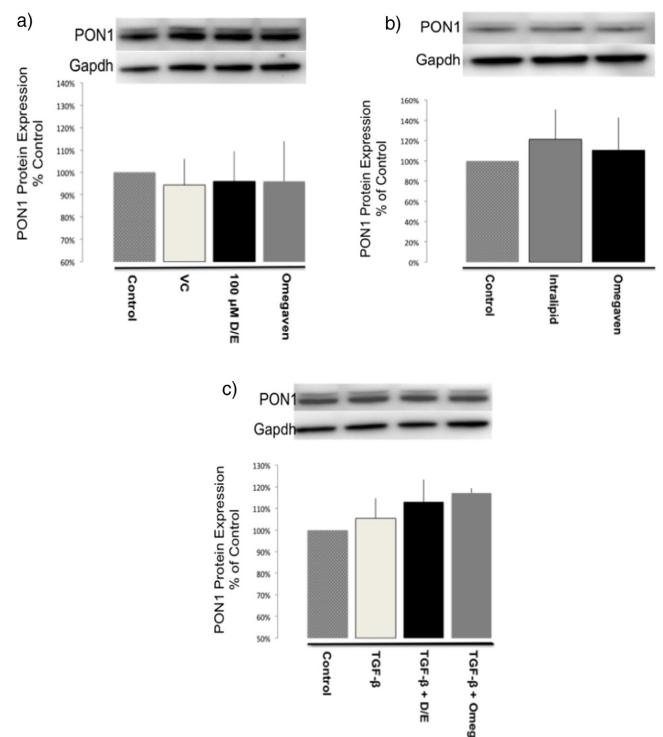
Short bowel syndrome (SBS) is a form of intestinal failure that results from inadequate length of small intestine to fully digest and absorb nutrients and calories [1]. Parenteral nutrition associated liver disease (PNALD) occurs in 40%-60% of infants and children receiving prolonged courses of life-saving parenteral nutrition (PN) for the treatment of conditions causing intestinal failure, including SBS [2]. The disease spectrum of PNALD includes steatosis, cholestasis, fibrosis, and ultimately cirrhosis [3]. The pathogenesis driving PNALD remains incompletely understood. The current FDA approved lipid emulsion, Intralipid®, is a 100% soybean oil-based lipid emulsion that contains predominantly omega-6 fatty acids, which may contribute to the progression of PNALD [3]. Treatment options for PNALD include stopping the TPN, decreasing lipid calories, reintroducing enteral feeds, and/or switching to other lipid sources such as Omegaven<sup>™</sup> [3]. Omegaven<sup>™</sup> is a fish oil-based lipid emulsion that is high in omega-3 fatty acids ( $\Omega$ 3FA), docosahexaenoic acid (DHA), and eicosapentaenoic acid (EPA) [4,5]. Several published case reports suggest Omegaven<sup>™</sup> has lessened and even reversed some stages of PNALD, however, the mechanisms of this effect are unknown [4,5].

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In the early stages of PNALD, the liver undergoes inflammatory changes. Inflammatory cytokines, such as interleukin-1 (IL-1) and tumor necrosis factor alpha (TNF- $\alpha$ ), have been shown to be elevated in PNALD and to directly contribute to liver injury [6–8]. Studies have also demonstrated that the aforementioned inflammatory pathways can activate the transforming growth factor beta (TGF- $\beta$ ) pathway, which may contribute to further hepatocellular damage and fibrosis [8,9]. Paraoxonase 1 (PON1) is an antioxidant protein that is mainly synthesized and released by the liver [10]. It possesses lactonase activity and hydrolyzes lipid peroxides [11]. Systemically, it circulates in plasma bound to high density lipoprotein (HDL) and is thought to play an important anti-inflammatory role [10,11]. Previous studies demonstrate that the inflammatory cytokines IL-1 and TNF- $\alpha$  decrease PON1 mRNA levels but these studies did not include an analysis of PON1 protein expression [12]. To date, there is a paucity of data relating inflammatory cytokine pathways to PON1 protein expression and the role that  $\Omega$ 3FA might play in these pathways and on PON1. The aim of this study is to further understand the protective effects of Omegaven<sup>™</sup>, including its key components,  $\Omega$ 3FA, DHA, and EPA on IL-1, TNF- $\alpha$ , and TGF-\beta-mediated signaling pathways and liver damage in an in vitro model of PNALD.

We hypothesize that the damaging effects of cytokines will result in a decrease in PON1 protein expression and that this effect will be prevented by Omegaven<sup>TM</sup>.

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**Fig. 1.** (a) Bar graph and representative Western blot showing no statistical difference in PON1 protein expression when treated with controls,  $\Omega$ 3FA, or Omegaven<sup>TM</sup>. (b) Bar graph and Western blot showing no statistically significant difference between treatment and control, Intralipid<sup>TM</sup>, or Omegaven<sup>TM</sup> on PON1 protein expression. (c) Bar graph and representative Western blot showing no statistical effect of TGF- $\beta$  on PON1 protein expression. VC = Vehicle control, ethanol. D/E = 100  $\mu$ M DHA + EPA. n = 3.

#### 1. Methods

#### 1.1. Cell culture

Human hepatic HepG2 cells were cultured and maintained in Dulbecco's Modified Eagle's Medium (DMEM) (Invitrogen, Carlsbad, CA) supplemented with 10% heat-inactivated FBS in 100 mm plastic plates and kept between 50% to 80% confluence. Cells were plated in six well plates at  $3 \times 10^6$  per well and allowed to adhere overnight. Cells were

then treated with DMEM supplemented with either 20% Omegaven<sup>TM</sup> (Fresenius SE & Co. KgaA, Bad Homburg, Germany) 20% Intralipid® (Fresenius SE & Co. KgaA, Bad Homburg, Germany), or a 100  $\mu$ M mixture of DHA and EPA (Cayman Chemical, Ann Arbor, MI) in DMEM for 1 h, followed by the addition of either TNF- $\alpha$  (50 ng/mL) or IL-1 (50 ng/mL) for another 16 h. TGF- $\beta$  was also used (50 ng/mL) and treated cells were incubated overnight for PON1 analysis or 60 min for p-Smad2/3 analysis as described previously [13]. Cells were harvested, lysed, and PON1 and p-Smad2/3 expression was measured by Western blot analysis.

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