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RELEVANT VARIATIONS AND NEUROPROTECIVE EFFECT OF

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HYDROGEN SULFIDE IN A RAT GLAUCOMA MODEL
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COH rats. Furthermore, ¹ING UANG, ^a SHUO XU, ^a

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- 17 Abstract-Glaucoma is an irreversible and blinding neurodegenerative disease of the eye, and is characterized by progressive loss of retinal ganglion cells (RGCs). Since endogenous hydrogen sulfide (H₂S) was reported to be involved in neurodegeneration in the central nervous system, the authors aimed to develop a chronic ocular hypertension (COH) rat model simulating glaucoma and therein test the H₂S level together with the retinal protein expressions of related synthases, and further investigated the effect of exogenous H₂S supplement on RGC survival. COH rat model was induced by cross-linking hydrogel injection into anterior chamber, and the performance of the model was assessed by intraocular pressure (IOP) measurement, RGC counting and retinal morphological analysis. Endogenous H₂S level was detected along with the retinal protein expressions of H₂S-related synthases cystathionine β -synthase (CBS), cystathionine γ -lyase (CSE) and 3-mercaptopyruvate sulfurtransferase (3-MST) in the COH rats. Retinal H₂S level and RGC survival were evaluated again after NaHS (a H₂S donor) treatment in the COH rats. The results showed that the COH model succeeded in simulating glaucoma features, and retinal H₂S level decreased significantly when the retinal protein expressions of CBS, CSE and 3-MST were downregulated generally in the

COH rats. Furthermore, the decrease of retinal H₂S level and loss of RGCs were both improved by NaHS treatment in experimental glaucoma, without obvious variation of IOP. Our study revealed that the intracameral injection of cross-linking hydrogel worked efficiently in modeling glaucoma, and H₂S had protective effect on RGCs and might be involved in the pathological mechanism of glaucomatous neuropathy. © 2016 IBRO. Published by Elsevier Ltd. All rights reserved.

Key words: chronic ocular hypertension, glaucomatous neuropathy, retinal ganglion cells, hydrogel, hydrogen sulfide.

INTRODUCTION

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Hydrogen sulfide (H₂S), long known for its characteristic 20 odor of rotten eggs and toxic nature, was recently 21 recognized as the third member of endogenous 22 gasotransmitter family, along with carbon monoxide and 23 nitric oxide (Abe and Kimura, 1996; Lowicka and 24 Beltowski, 2007; Qu et al., 2008). This novel gaseous 25 signaling molecule was reported to be implicated in 26 physiological and pathological processes as diverse as 27 neuroregulation, vasodilation, internal secretion, inflam-28 mation. etc. (Kaneko et al., 2006; Yang et al., 2008; 29 Zhang and Bhatia, 2008; Papapetropoulos et al., 2009; 30 Kimura, 2010; Tan et al., 2010; Tay et al., 2010). The 31 endogenous H₂S in humans and other mammalians is 32 mainly produced by cystathionine β -synthase (CBS), 33 cystathionine γ -lyase (CSE) and 3-mercaptopyruvate sul-34 furtransferase (3-MST) along with cysteine aminotrans-35 ferase, which localized in various tissues with different 36 distributions (Levonen et al., 2000; Meier et al., 2001; 37 Sen et al., 2012; Kimura, 2014). So far as the ocular tis-38 sues were concerned, CBS, CSE and 3-MST were 39 reported to be expressed in the retina of human as well 40 as other mammalians, and take part in various retinal 41 physiopathological processes (Persa et al., 2006; Pong 42 et al., 2007; Mikami et al., 2011; Markand et al., 2013; 43 Si et al., 2013). 44

Nowadays, the cytoprotective effect of H₂S is drawing 45 accumulative interests of researchers to explore its 46 therapeutic potentials. It was reported that H₂S played a 47 role in neurodegenerative diseases in the central 48 nervous system (CNS) (such as Alzheimer's disease 49 and Parkinson's disease), presenting a decreased level 50 in brain tissue and blood plasma, and the disease-51 associating symptoms and the biomolecular features 52

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Abbreviations: 3-MST, 3-mercaptopyruvate sulfurtransferase; CBS, cystathionine β-synthase; CNS, central nervous system; COH, chronic ocular hypertension; CSE, cystathionine γ-lyase; DAPI, 4',6-diamidino-2-phenylindole; Dil, 1,1'-dioctadecyl-3,3,3',3'-tetramethy-indocarbocya nin perchlorate; D-PBS, Dulbecco's phosphate-buffered saline; GCL, ganglion cell layer; H₂S, hydrogen sulfide; HCCS, HyStem Cell Culture Scaffold kit; INL, inner nuclear layer; IOP, intraocular pressure; IPL, inner plexiform layer; RGCs, retinal ganglion cells; TUNEL, terminal deoxynucleotidyl transferase-mediated deoxyuridine triphosphate nick end labeling.

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were alleviated by application of exogenous supply of H₂S 53 (Hu et al., 2010; Kamat et al., 2015). In the eye, it was dis-54 covered that the endogenous H₂S level in plasma and 55 retina decreased in streptozotocin-induced diabetic 56 retinopathy in rats, and treatment with exogenous H₂S 57 donor, which increased the internal H₂S level, improved 58 the impaired retinal structure and function reasonably 59 60 (Si et al., 2013). Furthermore, supplement of H₂S was found to mediate therapeutic effect on retinal ischemia/ 61 reperfusion injury and NMDA-induced neuronal lesion 62 (Biermann et al., 2011; Sakamoto et al., 2014). 63

Glaucoma is a leading cause of irreversible blindness 64 65 worldwide, characterized by gradually impairing the visual 66 field due to loss of retinal ganglion cells (RGCs) over time. in which the elevation of intraocular pressure (IOP) plays 67 an essential role (Gupta et al., 2006; Quigley and 68 Broman, 2006). Although the pathogenesis of glaucoma 69 is still not fully understood yet, it is generally accepted that 70 the apoptosis of RGCs and the secondary damage to the 71 neighbor neurons lead to the impairment of visual func-72 tion. Glaucoma is recognized as a neurodegenerative dis-73 ease, and the oxidative stress, mitochondrial dysfunction 74 75 and immune-inflammatory response were confirmed to be 76 implicated in the pro-apoptotic mechanisms of RGCs in 77 glaucoma (Pinazo-Duran et al., 2013), which were also 78 the targets for the therapeutic potentials of H₂S. In the 79 present study, we explored a new type of rat model of 80 chronic ocular hypertension (COH) for evaluating the variations of endogenous level of H₂S together with the retinal 81 protein expressions of CBS, CSE and 3-MST, and further 82 assessed the protective effect of exogenous supplement 83 of H₂S on RGCs in experimental glaucoma, trying to 84 investigate the role of H₂S in the mechanism of glaucoma-85 tous neuropathy. 86

Animals

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All experimental procedures described here conformed to 89 the National Institutes of Health guidelines for the Care 90 and Use of Laboratory. The animal procedures were in 91 accordance to the ARVO Statement for the Use of 92 Animals in Ophthalmic and Vision Research, and with 93 approval from the institutional review board of Ruijin 94 Hospital, Shanghai, China. Male adult Sprague-Dawley 95 rats weighing approximately 200-250 g were obtained 96 from Shanghai Slack Laboratory Animals Ltd (Shanghai, 97 China). The rats were fed ad libitum and maintained in 98 an air-conditioned room at approximately 23 °C and 99 100 60% humidity, in a 12-h light-dark cycle for the duration 101 of the experiments. All surgeries were performed under systemic anesthesia, and all efforts were made to 102 minimize suffering. The details of number of animals 103 used in separate experimental procedures are listed in 104 Table 1. 105

MATERIALS AND METHODS

106 Induction of IOP elevation

Rats were anesthetized by intraperitoneal injection of
xylazine 10 mg/kg (Sigma–Aldrich, St. Louis, MO) and
ketamine hydrochloride 25 mg/kg (Sigma–Aldrich),

Table	1.	Number	of	animals	used	in	separate	procedures
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Procedure	Ν	Procedure	Ν
Identification of purified RGCs	54	H ₂ S level measurement	24
CCK-8 assay	12	Immunohistochemistry staining	24
TUNEL assay	108	Western blotting	24
Intracameral injections for model building and IOP profile	60	H ₂ S level measurement (drug treated)	24
Retrograde labeling of RGCs	24	Retrograde labeling of RGCs (drug treated)	24
Hematoxylin–Eosin staining	24		

respectively. Topical anesthesia to each eye was 110 conducted by a drop of 0.5% proparacaine 111 hydrochloride (Bausch & Lomb, Tampa, FL). The COH 112 was induced in the right eyes of the rats. The IOP was 113 elevated by injecting a pre-mixed in situ cross-linking 114 hydrogel, HyStem Cell Culture Scaffold kit (HCCS; 115 Sigma-Aldrich), into the anterior chamber. HCCS 116 consisted of HyStem (a thiol-modified carboxymethyl 117 hyaluronic acid) and Extralink (a thiol-reactive 118 polyethylene glycol diacrylate), both dissolved in 119 decassed water according to the manufacturer's 120 instructions and mixed at the ratio of 4:1 immediately 121 before the injection. The cross-linking gelation of the 122 liquid mixture occurred in situ about five minutes in the 123 anterior chamber. The anterior chamber puncture was 124 performed in cornea from the peripheral area toward the 125 central to create a long enough tunnel incision with a 126 31-gauge needle. Then a volume of 7 µL fresh liquid 127 mixture of HyStem and Extralink was immediately 128 injected into the anterior chamber through the incision 129 with a Hamilton syringe (Hamilton Bonaduz AG, 130 Switzerland). However, the left eyes were not 131 determined as control eyes to avoid potential 132 inflammatory reactions caused by contralateral COH 133 eyes (Rojas et al., 2014) and to reduce the suffering of 134 animals in consideration of humane care. Sham-135 operations, which were similar to the COH-inducing oper-136 ations except for the injection of an equal volume of saline 137 solution substitute for Hystem and Extralink mixture, were 138 performed on the right eyes of else rats to be determined 139 as control group and employed in all following experi-140 ments. A drop of 0.5% levofloxacin hydrochloride (Santen 141 Pharmaceutical, Japan) was used for infection prevention 142 after operations. 143

Drug administration

NaHS(5.6 mg/kg;Sigma–Aldrich)wasinjected145intraperitoneally to treat rats, being started 3 days prior146146to the induction of COH and maintained for 4 weeks on147a daily basis till the executions were done. Saline of the148equivalent volume was used as treatment of control.149

IOP measurement

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IOP	m	easurements	;	were	dor	ne u	nder	brief	1	51
system	ic	anesthesia	by	isoflu	urane	inhala	tion	(2–4%;	1	52

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