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A SUBNANOMOLAR CONCENTRATION OF PITUITARY ADENYLATE CYCLASE-ACTIVATING POLYPEPTIDE (PACAP) PRE-SYNAPTICALLY MODULATES GLUTAMATERGIC TRANSMISSION IN THE RAT HIPPOCAMPUS ACTING THROUGH ACETYLCHOLINE

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- 17 Abstract—The neuropeptide PACAP modulates synaptic transmission in the hippocampus exerting multiple effects through different receptor subtypes: the underlying mechanisms have not yet been completely elucidated. The neurotransmitter acetylcholine (ACh) also exerts a well-documented modulation of hippocampal synaptic transmission and plasticity. Since PACAP was shown to stimulate ACh release in the hippocampus, we tested whether PACAP acting through ACh might indirectly modulate glutamate-mediated synaptic transmission at a pre- and/ or at a post-synaptic level. Using patch clamp on rat hippocampal slices, we tested PACAP effects on stimulationevoked AMPA receptor-mediated excitatory post-synaptic currents (EPSCs_{AMPA}) in the CA3-CA1 synapse and on spontaneous miniature EPSCs (mEPSCs) in CA1 pyramidal neurons. A subnanomolar dose of PACAP (0.5 nM) decreased EPSCs_{AMPA} amplitude, enhanced EPSC paired-pulse facilitation (PPF) and reduced mEPSC frequency, indicating a presynaptic decrease of glutamate release probability: these effects were abolished by simultaneous blockade of muscarinic and nicotinic ACh receptors, indicating the involvement of endogenous ACh. The effect of subnanomolar PACAP was abolished by a PAC1 receptor antagonist but

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not by a VPAC receptor blocker. At a higher concentration (10 nM), PACAP inhibited EPSCs_{AMPA}: this effect persisted in the presence of ACh receptor antagonists and did not involve any change in PPF or in mEPSC frequency, thus was not mediated by ACh and was exerted postsynaptically on CA1 pyramidal neurons. We suggest that a high-affinity PAC1 receptor pre-synaptically modulates hippocampal glutamatergic transmission acting through ACh. Therefore, administration of PACAP at very low doses might be envisaged in cognitive diseases with reduced cholinergic transmission. © 2016 IBRO. Published by Elsevier Ltd. All rights reserved.

Key words: PACAP, neuropeptide, synaptic transmission, glutamate, AMPA, acetylcholine.

INTRODUCTION

The neuropeptide Pituitary Adenylate Cyclase-Activating 20 Polypeptide (PACAP), initially isolated from ovine 21 hypothalamus (Mivata et al., 1989), is widely diffused in 22 the central nervous system (CNS) as well as in peripheral 23 tissues and exists in two forms with either 38 or 27 amino 24 acid residues (PACAP-38 and PACAP-27). PACAP-38 25 (PACAP) is the most abundant form in the CNS where it 26 acts as a neurotransmitter, neuromodulator and/or neu-27 rotrophic factor (Shioda, 2000; Vaudry et al., 2009) and 28 regulates numerous functions such as circadian rhythms, 29 responses to stress and brain injury, mood and learning 30 (Harmar et al., 2012). PACAP can activate two main 31 receptor types: the first is a PACAP-selective receptor 32 named PAC1, which binds PACAP with nanomolar affinity 33 and the analog peptide vasoactive intestinal peptide (VIP) 34 with a 1,000-fold lower affinity (Pisegna & Wank, 1993). 35 The second family includes VPAC1 and VPAC2 recep-36 tors, binding PACAP and VIP with equal nanomolar affin-37 ity (Ishihara et al., 1992; Lutz et al., 1993; Sreedharan 38 et al., 1993; Harmar & Lutz, 1994). PAC1, VPAC1 and 39 VPAC2 receptors are G-protein-coupled receptors that 40 potently stimulate adenylate cyclase (AC) and increase 41 intracellular cyclic AMP (cAMP) levels; activation of other 42 intracellular messengers, including calcium and/or phos-43 pholipase D, has also been reported particularly for 44 PAC1 receptors (Dickson & Finlayson, 2009). 45

Various studies indicate that PACAP modulates 46 cognitive functions inducing neurotrophic/ 47

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Abbreviations: ACh, acetylcholine; ACSF, artificial cerebrospinal fluid; AMPA, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid; CA1, Cornu Ammonis 1; CA3, Cornu Ammonis 3; cAMP, cyclic adenosine mono phosphate; D-AP5, p-(-)-2-Amino-5-phosphonopentanoic acid; EPSC, excitatory post-synaptic current; EPSCAMPA, AMPA receptormediated excitatory post-synaptic current; fEPSPs, field excitatory post-synaptic potentials; LTD, long-term depression; LTP, long-term potentiation; mEPSCs, spontaneous miniature EPSCs; NMDA, N-methyl-o-aspartate; PACAP, Pituitary Adenylate Cyclase-Activating Polypeptide; PPF, paired-pulse facilitation; TTX, Tetrodotoxin.

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neuroprotective effects as well as modulation of neuronal 48 excitability and synaptic transmission (Sauvage et al., 49 2000; Sacchetti et al., 2001; Otto et al., 2001a; Borbely 50 et al., 2013). Among brain structures playing a key role 51 in learning and memory, the hippocampus contains 52 PACAP-expressing cells and nerve fibers, as shown in 53 the rat by immunohistochemistry (Koves et al., 1991; 54 55 Piggins et al., 1996; Hannibal, 2002) and in situ hybridization (Skoglosa et al., 1999; Jaworski & Proctor, 2000; 56 Hannibal, 2002). 57

All PACAP receptor subtypes are expressed in the 58 hippocampus since early developmental stages 59 (Jaworski and Proctor, 2000). PAC1 receptors are located 60 61 on CA1-CA3 pyramidal cells (Shioda et al., 1997; Joo et al., 2004; Gupte et al., 2015), on mossy fiber terminals 62 in the dentate gyrus (Otto et al., 1999) and, as recently 63 evidenced, also on glial cells (Gupte et al., 2015). VPAC1 64 and VPAC2 receptors are expressed in all hippocampal 65 regions: in particular, VPAC2 receptors are very abundant 66 on pyramidal cells of the CA1-CA3 regions and on gran-67 ule cells of the dentate gyrus (Joo et al., 2004). 68

In hippocampal neurons, PACAP inhibits a slow 69 afterhyperpolarizing membrane ion current (I_{AHP}) (Taylor 70 71 et al., 2014) and downregulates Kv4.2 channels responsi-72 ble for I_A, a hyperpolarizing potassium current (Gupte et al., 2015), in line with previous studies showing that 73 PACAP enhances neuronal firing in the hippocampus 74 (Di Mauro et al., 2003; Liu et al., 2003). PACAP also mod-75 ulates glutamate-mediated hippocampal synaptic trans-76 mission: a variety of effects have been described 77 depending on PACAP concentration and on experimental 78 conditions. A high dose of PACAP (1 µM) induced a long-79 term inhibition of field excitatory post-synaptic potentials 80 (fEPSPs) recorded in the CA1 region of rat hippocampus 81 after stimulation of Schaffer collaterals (Kondo et al., 82 1997). In similar experimental conditions, another 83 84 research group confirmed that PACAP (1 µM) inhibited 85 fEPSP amplitude, whereas a very low subnanomolar dose (0.05 nM) of PACAP enhanced fEPSP amplitude 86 and intermediate doses (100-500 nM) exerted biphasic 87 inhibitory/excitatory effects (Roberto et al., 2001). In 88 mouse hippocampus, application of PACAP-27 at a 89 400 nM concentration induced a long-term inhibition of 90 91 fEPSPs recorded in the CA1 region (Ster et al., 2009). In our laboratories, we observed that PACAP at a 92 nanomolar dose (10 nM) reduced the amplitude of 93 glutamate-mediated excitatory post-synaptic currents 94 (EPSCs) recorded under patch clamp from rat CA1 pyra-95 midal neurons after stimulation of Schaffer collaterals 96 (Ciranna & Cavallaro, 2003). 97

98 Other studies later revealed that PACAP differentially modulates the two main components of fast glutamatergic 99 transmission, respectively mediated by a-amino-3-hydro 100 xy-5-methyl-4-isoxazolepropionic acid (AMPA) and N-101 methyl-p-aspartate (NMDA) receptors for glutamate. 102 PACAP enhances NMDA receptor-mediated synaptic 103 responses in the hippocampal CA1 region through 104 activation of PAC1, VPAC1 and VPAC2 receptors (Yaka 105 et al., 2003; Macdonald et al., 2005). On the other side, 106 the effects of PACAP on AMPA receptor-mediated cur-107

rents seem more complex (Yang et al., 2010). We 108 observed that excitatory currents induced in rat CA1 pyra-109 midal neurons by direct application of AMPA were 110 enhanced by a subnanomolar (0.5 nM) concentration of 111 PACAP and were instead inhibited by PACAP at a 112 10 nM dose (Costa et al., 2009). Interestingly, it was 113 recently demonstrated that PACAP inversely regulates 114 the phosphorylation state of two distinct sites on the 115 GluA1 subunit of AMPA receptors, acting in part through 116 an interaction with NMDA receptors (Toda & Huganir, 117 2015). 118

PACAP also modulates hippocampal long-term synaptic plasticity: as already mentioned, a PACAPinduced long-term depression (LTD) was described in the CA3-CA1 synapse (Kondo et al., 1997; Roberto et al., 2001; Ster et al., 2009). On the other side, PACAP-deficient mice display reduced long-term potentiation (LTP) in the dentate gyrus (Matsuyama et al., 2003) and learning impairment (Takuma et al., 2014). PAC1 receptor knockout mice show impaired LTP at the mossy fiber-CA3 synapse (Otto et al., 2001a; Otto et al., 2001b; Matsuyama et al., 2003) and selective deficits in associative memory (Otto et al., 2001a). Another recent study shows that PACAP infusion in the hippocampus and in the basolateral amygdala affects memory consolidation and extinction in rats submitted to contextual fear conditioning (Schmidt et al., 2015). Collectively, all these data point out that PACAP exerts multiple modulatory effects on basal glutamatergic excitatory transmission. longterm synaptic plasticity, learning and memory.

Some indication exists that PACAP interacts with the cholinergic system in peripheral tissues and in the CNS: PACAP is co-localized with acetylcholine (ACh) in presynaptic sympathetic nerve terminals in the adrenal gland (Hamelink et al., 2002). A stimulatory effect of PACAP on ACh release was observed in parasympathetic ciliary ganglion neurons (Pugh et al., 2010). In the rat hippocampus, an *in vivo* microdialysis study shows that PACAP stimulates ACh release (Masuo et al., 1993) and PACAP effect on glutamatergic transmission was partially reduced by ACh receptor antagonists (Roberto & Brunelli, 2000).

As indicated above, PACAP effects on the CA3-150 CA1 synapse are variable and dose-dependent: 151 besides, results from distinct research groups are not 152 always comparable due to different experimental 153 conditions. The mechanisms underlying the large 154 variability of PACAP-induced effects on glutamatergic 155 synaptic transmission still need to be clarified, 156 particularly with respect to: 1) differential modulation 157 of AMPAR- and NMDAR-mediated synaptic ion 158 currents; 2) multiple sites of PACAP action; 3) 159 indirect effects involving the cholinergic system. To 160 clarify these issues, we have tested the effects of 161 PACAP specifically the AMPAR-mediated on 162 component of glutamatergic transmission in the CA3-163 CA1 synapse, focusing on the site of action (pre- vs 164 post-synaptic) of PACAP effect. Secondly, we have 165 investigated which fraction of PACAP effect might be 166 indirectly mediated by ACh release. 167

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