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Relationship between nutrients and plankton biomass in the turbidity maximum zone of the Pearl River Estuary

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ABSTRACT

Nutrients, dissolved and particulate organic carbon and plankton (bacterio-, phyto- and zoo-) 19 were compared in the turbidity maximum zone (TMZ) and adjacent areas (non-TMZ) in the 20 Pearl River estuary. Our results showed that high levels of suspended substances had marked 21 effect on dynamics of nutrients and plankton in the TMZ. Based on the cluster analysis of total 22 suspended solids (TSS) concentrations, all stations were divided into two groups, TMZ with 23 average TSS of 171 mg/L and non-TMZ of 45 mg/L. Suspended substances adsorbed PO_4^{3-} and 24 dissolved organic carbon, resulting in higher particulate phosphorus and organic carbon (POC) 25 and lower PO₄⁻ and DOC in the TMZ, compared to the non-TMZ. However, suspended 26 substances had limited effect on nitrogenous nutrients. Phytoplankton growth was 27 light-limited due to high concentrations of suspended substances in the TMZ and a peak of 28 phytoplankton abundance appeared in the non-TMZ. In contrast, the highest bacterial 29 abundance occurred in the TMZ, which was likely partly responsible for low DOC levels. Two 30 peaks of zooplankton abundance observed in the TMZ and non-TMZ in the Pearl River estuary 31 were primarily supported by bacteria and phytoplankton, respectively. Our finding implied 32 that high levels of suspended solids in the TMZ affect the trophic balance. 33 © 2016 The Research Center for Eco-Environmental Sciences, Chinese Academy of Sciences. 34

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47 Introduction

Turbidity maximum zone (TMZ) is the dynamic region in an estuary, characterized by higher concentration of suspended matter than upstream and downstream of the estuary. The formation of the TMZ is generally attributed to gravitational circulation, tidal trapping, and sediment resuspension (Wai et al., 2004). TMZ has been observed and studied in many estuaries, including Gironde Estuary in France (Allen et al., 1977), Chesapeake Bay in USA (Sanford et al., 2001), 55 St. Lawrence Estuary in Canada (Hamblin, 1989), Fly River 56 Estuary in Papua New Guinea (Wolanski et al., 1995), Tamar 57 Estuary in England (Uncles and Stephens, 1993), and 58 Changjiang Estuary in China (Shen et al., 2008). However, the 59 formation mechanism of TMZ varies between different 60 estuaries, the location and intensity of TMZ are sensitive to 61 changes in tidal range and freshwater flow within one estuary 62 (Mitchell, 2013). 63

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The high concentrations of suspended matter play an 64 important role in regulating physical, chemical and biological 65 reactions between dissolved and particulate species as well as 66 interactions among particulate species (Gebhardt et al., 2005). 67 The tidal current causes periodic deposition and resuspension 68 of particles during tidal cycles and increases residence times 69 of particle and dissolved material (Herman and Heip, 1999), 70 which affects important physical and biological processes 71 72such as the flocculation, solubilization, sorption/desorption, 73 redox and mineralization (Goni et al., 2005). For example, the periodic resuspension of particles at the TMZ may facilitate 74 the oxidation of recalcitrant particulate organic matter by 75recharging the metal-oxide phases due to repeated exposure 76 to oxygenated water (Aller, 1994, 1998). Several studies 04 03 (Crump and Baross, 1996; Ploug et al., 2002; Servais and 78 Garnier, 2006) have shown that bacteria attached to particles 79 dominated the total bacterial activity in TMZ, and bacterial 80 mineralization of the organic carbon was enhanced by the 81 high suspended substances concentration in the TMZ. In 82 addition, the high concentrations of the suspended particles 83 in the TMZ decrease the light penetration which restricts 84 phytoplankton growth (Garnier et al., 2001). However, the 85 particulate organic matter in the TMZ can be considered as 86 87 the basis of a detrital food chain which can be consumed by zooplankton and hyperbenthos directly (Islam et al., 2006; 88 Mitchell, 2013). Furthermore, TMZ could create a predation 89 90 refuge in high turbidity water (Islam et al., 2006).

91 The Pearl River Estuary (PRE) is a very complicated large-scale estuarine system in China. Pearl River has three 92 main tributaries, West, North and East Rivers, which pass 93 94 through a complex river network and flow into the South China Sea through eight entrances. The total annual runoff 95 reaches $3.33 \times 10^{11} \text{ m}^3$ and has an annual suspended sedi-96 ment load of 8.87×10^7 tons (Zhao, 1990; Xia et al., 2004). The 97 PRE has a high loading of anthropogenic nutrients from 98 activities in agriculture, fish dike farming and sewage 99 effluents due to the high population and economic develop-100 ment in Pearl River delta region (Yin et al., 2001; Huang et al., 05 2003) The TMZ in the PRE is well developed and is primarily 102 characterized by a high TSS compared with adjacent waters. 103 Previous studies on TMZ in the PRE only focus on their 104 dynamic mechanism (Wai et al., 2004). However, little 105106 attention has been paid to biogeochemical processes in the TMZ. In this study, nutrients, organic carbon and plankton 107 (bacterio-, phyto- and zoo-) were compared between TMZ and 108 non-TMZ, in order to examine effect of turbidity maximum on 109patterns of nutrients and plankton in the PRE. 110

112 1. Materials and methods

In the present study, 16 stations were visited in the main
channel of the PRE during 6 to 10 January 2011 (during high
tide) (Fig. 1). Vertical profiles of salinity were measured *in situ*with a multi-parameter sensor YSI6600 (YSI, Yellow Springs,
Ohio). Water samples were collected at the surface (in 0.5 m)
and bottom layers (1 m above the seabed) at all stations using
a 5-L Niskin bottle.

Each water sample was immediately filtered through Whatman GF/F filters (0.7 μ m pore size), nutrient (NO₃, NO₂, NH⁴₄, and PO³⁻₄) were preserved in polyethylene bottle. Total 122 dissolved phosphorus (TDP) and dissolved organic carbon 123 (DOC) were preserved in glass bottle and immediately frozen 124 (–20°C) until analyzed. Nutrient concentrations were deter-125 mined colorimetrically following the protocols described by 126 Grasshoff et al. (2009). TDP was measured using the approach 127 described by Valderrama (1981). The concentrations of dissolved 128 organic phosphorus (DOP) were determined by subtracting the 129 dissolved inorganic phosphorus (PO³⁻₄) concentrations from the 130 TDP. Samples for DOC concentrations were analyzed with a 131 high temperature combustion method using a Liquid TOC II 132 analyzer (Knap et al., 1996). The analytical precision of nutrient, 133 DOC and TDP was <5%.

Total suspended solids (TSS) samples were all filtered on to 135 pre-weighed Whatman GF/F fiber filters (0.7 µm pore size), 136 and the filters were dried at 45°C and weighed to determine 137 the amount in mg/L of sample. Samples for particulate 138 organic matter analysis were filtered on to precombusted 139 Whatman GF/F filters and stored at –20°C. The concentrations 140 of particulate organic carbon (POC) and particulate organic 141 nitrogen (PON) were determined with a CHN Elemental 142 Analyzer (Elementar, Vario EL-III, Germany). The analytical 143 precision of POC and DOP was 6% and 7%, respectively. Total 144 particulate phosphorus (TPP) was extracted with 1 mol/L 145 hydrochloric acid (GR, Sigma) after ignition at a high 146 temperature (550°C) then determined by using the ammo- 147 nium molybdate method with ascorbic acid reduction 148 (Aspila et al., 1976). For particulate inorganic phosphorus 149 (PIP) measurements, samples were digested with 1 mol/L 150 hydrochloric acid for 16 hr at room temperature, including 151 2 hr of vibration, and then analyzed according to Aspila 152 et al. (1976). Particulate organic phosphorus (POP) was defined 153 by subtracting the PIP from TPP. The analytical precision of 154 PIP and TPP was <5%. 155

Water for bacteria enumeration was collected at the 156 surface and prefiltered through a 20 μ m mesh netting. 157 Triplicate samples were fixed with formaldehyde (2% final 158 concentration) for 15 min in 2 mL cryotubes, quick-frozen in 159 liquid nitrogen. The bacteria abundance were counted with an 160 epifluorescence microscope (OLYMPUS BX51, Japan) with a 161 100 W high-pressure mercury burner for epifluorescence 162 illumination after staining with DAPI (4,6-diamidino-2- 163 phenylindole dihydrochloride, Sigma) for at least 5 min in 164 the dark (Porter and Feig, 1980). Stained bacterial cells 165 were counted at 1000× magnification under UV excitation, 166 and at least 10–20 random fields (minimum of 400 cells) were 167 counted. 168

Water for phytoplankton enumeration was collected at the 169 surface and preserved with 1% acidified Lugol's iodine 170 solution. Phytoplankton were concentrated by settling 5– 171 25 mL aliquots by sedimentation for 48 hr and then identified 172 with a microscope (>2 μ m were counted) according to the 173 method described by Utermöhl (1958). 174

Zooplankton was sampled using a shallow-water type I net 175 (the mouth area, 0.2 m^2 ; the mesh pore size, 505μ m) by 176 towing vertically from 1 m above the bottom to the surface 177 and the volume of the filtered water was determined with a 178 flow-meter (Hydro-Bios) attached to the net. The zooplankton 179 samples were preserved immediately in 5% formaldehyde, 180 and then identified and counted with a microscope. 181

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