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Q2 Nano-sized $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$ facilitate anaerobic transformation of 2 hexavalent chromium in soil–water systems

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ABSTRACT

The purpose of this study is to investigate the effects of nano-sized or submicro $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$ on 17 the bioreduction of hexavalent chromium (Cr(VI)) and to evaluate the effects of nano-sized 18 $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$ on the microbial communities from the anaerobic flooding soil. The results 19 indicated that the net decreases upon Cr(VI) concentration from biotic soil samples amended 20 with nano-sized Fe_2O_3 (317.1 ± 2.1 mg/L) and Fe_3O_4 (324.0 ± 22.2 mg/L) within 21 days, which 21 were approximately 2-fold of Cr(VI) concentration released from blank control assays ($117.1 \pm$ 22 5.6 mg/L). Furthermore, the results of denaturing gradient gel electrophoresis (DGGE) and 23 high-throughput sequencing indicated a greater variety of microbes within the microbial 24 community in amendments with nano-sized $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$ than the control assays. Especially, 25 *Proteobacteria* occupied a predominant status on the phylum level within the indigenous 26 microbial communities from chromium-contaminated soils. Besides, some partial decrease of 27 soluble Cr(VI) in abiotic nano-sized $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$ amendments was responsible for the 28 adsorption of nano-sized $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$ to soluble Cr(VI). Hence, the presence of nano-sized 29 $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$ could largely facilitate the mobilization and biotransformation of Cr(VI) from 30 flooding soils by adsorption and bio-mediated processes. 31

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Introduction

48 Chromium (Cr) has been listed as one of the top 20 contami-
49 nants in the superfund priority list of hazardous substances
50 (Dhal et al., 2013). For centuries, the anthropic activities, e.g.,
51 mining, processing, and smelting activities have greatly
52 contaminated the soil and water resources, resulting in an
53 unavoidable introduction of Cr-contaminants (Wang et al.,
54 2007). Geochemical processes and weathering, acting upon
55 metallurgical wastes and by-products, initiate transport of Cr
56 from contaminated areas and redistribution to surrounding
57 soils, streams, and groundwater. Thus, Cr will be accumulated

in soils. Its presence could be regarded as one of potential 58 largest environmental risks due to the deposit/precipitation of 59 Cr-contaminants, especially emerged in the soils close to Cr 60 mines areas (Desjardina et al., 2002). Therefore, there was an 61 urgent need to pay the efforts on investigating the transforma- 62 tion of Cr from soil–water systems as seeking the feasible 63 remediation on Cr pollution in the environment (Teng et al., 64 2013). 65

In general, the main species of Cr were presented with 66 trivalent (Cr(III)) and hexavalent (Cr(VI)). Cr(VI) is highly soluble 67 and toxic. However, the reduced form, Cr(III), is relatively in 68 soluble and in a low toxicity (Arias and Tebo, 2003). In alkaline 69

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soils, such as the soil from Cr mine, the predominant state is hexavalent Cr(VI) and existed with the form of CrO_4^{2-} and $\text{Cr}_2\text{O}_7^{2-}$ (Døssing et al., 2011). In soil-water systems, microorganisms play important roles in the transformation of Cr(VI) and Cr(III). Recently, microbial mediation on the transformation of Cr(VI) in soil-water systems has been well studied (Samuel et al., 2012; Wang and Shen, 1995). Many microorganisms could directly reduce Cr(VI), such as *Proteobacteria*, *Shewanella* alga, *Bacillus* spp., and several new isolated bacterial (Basu et al., 1997; Garavaglia et al., 2010; Shen and Wang, 1993). Guha et al. (2001) have found that oxygen could also serve as electron acceptors and subsequently competed with Cr(VI) in the systems containing high Cr(VI) concentrations. Moreover, Cr(VI) in estuarine soils was reported to be permanently reduced to Cr(III) under anaerobic conditions (Wadhawan et al., 2013). The regarding surveys illustrated that some limited factors, such as oxygen and electron donors, affected the bio-mediated process on the speciation and mobilization of Cr(VI) from soil.

Some materials, such as carbon sources, electron shuttles, and iron minerals affect Cr(VI) transformation under anaerobic conditions (Field et al., 2013). Further, given the extensive use of nanoparticles (Xu et al., 2012) recently, large amounts of nanoparticles have now been released into aquatic environments and soil systems. It will lead to an unexpected ecological and environmental outcome because of the unique physical and chemical properties of nanoparticles (Jiang et al., 2013; Zhu et al., 2016). Therefore, it is critical to explore how nanoparticles affect soil microbial communities, and the potential effects on Cr(VI) transformation. Additionally, the recent studies have demonstrated that nanoparticles facilitated extracellular electron transfer in microbial fuel cells and soil systems. The addition of magnetite nanoparticles into soils seemed to increase the activity of methanogens through shifting the microbial abundances of acetate-oxidizing bacteria, propionate-oxidizing bacteria, and methanogenic archaea (Yamada et al., 2015; Cutting et al., 2010). Because nanoparticles might potentially influence the microbes directly (e.g., via serving as electron shuttles to transport electrons), the possible mediating effects derived from iron oxide nanoparticles on Cr(VI) reduction should deserve to be studied. For instance, Rao et al. (2013) has demonstrated that a more significant modification to the yeast cells in presence of phyto-inspired $\text{Fe}^0/\text{Fe}_3\text{O}_4$ nanoparticles during Cr(VI) reduction than in absence of $\text{Fe}^0/\text{Fe}_3\text{O}_4$ nanoparticles. In addition, nanoscale zero-valent iron and biogenetic nano-magnetite were capable of removing aqueous Cr(VI) from alkaline groundwater (Watts et al., 2015; Liu et al., 2010; Li et al., 2011). Although there have been studies showing that iron oxide nanoparticles mostly affected Cr(VI) reduction process, there was no clear evidence regarding roles of nanoparticles during Cr(VI) reduction in soil (Singh et al., 2012). Exactly, our previous study demonstrated that the addition of nano-sized Fe_2O_3 , Fe_3O_4 and SiO_2 could potentially stimulate bacterial growth, and then changed the arsenic transformation in soil (Dong et al., 2014). Furthermore, Kato et al. (2010) reported that nano-sized Fe_3O_4 and nano zero valent iron oxide could also be used as electron conduits to dramatically improve microbial extracellular electron transfer in soil. Therefore, iron oxide nanoparticles, for example nano-sized Fe_3O_4 , may influence microbial respiratory of Cr(VI) in soil-water systems. However, there was severely poor studies could powerfully illustrate how nano-sized Fe_2O_3 /

Fe_3O_4 shift the microbial community composition during the process of Cr(VI) transformation.

Hence, the study presented the aims to investigate Cr(VI) transformation from Cr-contaminated soil-water systems in presence of nano-sized or submicro $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$, particularly emphasizing on the role of nano-sized $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$. Additionally, microbiota in soil sample was analyzed by denaturing gradient gel electrophoresis (DGGE) and using pyrosequencing after treatment of nano-sized $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$. Finally, the involving mechanisms regarding nano-sized $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$ affecting Cr(VI) transformation and microbial communities in soil were committed to be illuminated.

1. Experimental

1.1. Test site

Cr-contaminated soil samples were collected from a tailings at a Cr-enriched mine area in Qujing, Yunnan Province, China. The total Cr content in the experimental samples used was 12,202.5 mg/kg (Granadillo et al., 1994). The initial water-extraction Cr(VI) concentration in soil was 750 mg/kg and the initial water-extraction Cr(VI) concentration of each reactor was 325 mg/kg (Padaravskas et al., 1998). Content of other elements were negligible in soil. The pH of the soil was 8.9.

Soil samples from tailings were carefully collected and stored in sterile polyethylene bags and transported to the laboratory. Those not-dried samples to permitted for the microbiological analysis and used for the batches of soil microcosms culture incubation. The remained moist soil samples from tailings were stored in polyethylene vinyl containers at 4°C to maintain the original environment and survival of indigenous bacteria.

1.2. Sample preparation and treatment

Nano-sized Fe_2O_3 (30 nm)/ Fe_3O_4 (20 nm) and submicro $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$ were purchased from Aladin company. The morphology of the above particles was characterized by scanning electron microscopy (SEM) (Appendix A Fig. S1). The specific surface area of nano-sized Fe_2O_3 , nano-sized Fe_3O_4 submicro Fe_2O_3 and submicro Fe_3O_4 were 108.44m²/g, 8.92m²/g, 1.05m²/g and 0.04m²/g, respectively. All experimental operations were conducted under obligate anaerobic conditions. The treatment conditions were set up as follows: (1) anaerobic, 20 ± 1 g soil, treated with 0.2 g of submicro $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$ and 24 mL deionized water; (2) anaerobic, 20 ± 1 g soil, treated with 0.2 g of nano-sized $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$ and 24 mL deionized water; and (3) anaerobic, 20 ± 1 g soil, treated with 24 mL deionized water. In order to clarify if Cr(VI) could be abiotically reduced, another triplicate anaerobic samples with same amendments were incubated under the abiotic condition. The sterile treatments were autoclaved at 120°C for 20 min. The mixture of soil/water slurries were placed in 105 mL serum bottles, bubbled with N_2 for 30 min, fitted with a butyl rubber stopper, sealed with an aluminum clamp under an N_2 atmosphere, and finally incubated at 30°C in the dark. Soil microcosms were subsampled at discrete time points.

In the other experiments exploring the adsorption of Cr(VI) to nano-sized or submicro $\text{Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$, 100 mg/L Cr(VI)/(K₂CrO₄)

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