



Hydrogen gas prolongs the shelf life of kiwifruit by decreasing ethylene biosynthesis



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ABSTRACT

The application of hydrogen-rich water was recently shown to inhibit senescence process of kiwifruit, but it is unclear if hydrogen gas (H₂) exerts a similar function. The present results show that the production of endogenous H₂ in kiwifruit decreases during ripening. Fumigation with H₂ not only increased endogenous H₂ concentrations but delayed the softening and cell wall disassembly in flesh. Ethylene (C₂H₄) production was also inhibited by H₂, which was supported by a decrease in 1-aminocyclopropene-1-carboxylate (ACC) concentration, ACC synthase, and ACC oxidase activities, and the downregulation of the corresponding gene transcripts. Further, ACC-induced C₂H₄ production and ripening were blocked by H₂ and the incidence of natural decay and disease incidence were reduced. Overall, the results suggested that the H₂-induced delay of kiwifruit ripening resulted from an inhibitory effect on C₂H₄ biosynthesis.

1. Introduction

Kiwifruit is classified as a climacteric fruit with two widely known species: *Actinidia deliciosa* and *Actinidia chinensis*. In China, *A. deliciosa* 'Xuxiang' is known for its relatively large fruit size, green flesh, and good flavor (Li et al., 2017a), and has achieved commercial popularity. Despite these attractive fruit qualities, this cultivar is known for its rapid softening, which leads to a short shelf-life and greater susceptibility to fungal infection during ripening.

Ethylene (C₂H₄) is a key regulator of fruit senescence. For most climacteric fruits, C₂H₄ production continues at a slower rate during storage (Mworio et al., 2012). However, in kiwifruit, C₂H₄ production is almost zero at temperatures below 11–14 °C (Antunes et al., 2000); substantial amounts of C₂H₄ could only be detected after long periods of storage (Kim et al., 1999). Kiwifruit are highly sensitive to C₂H₄, and small amounts of C₂H₄ can accelerate fruit softening. As reported by McDonald and Harman (1982), 0.005–0.01 μL L⁻¹ C₂H₄ could induce the ripening and softening of kiwifruit. The control of C₂H₄ level is therefore crucial in postharvest handling of kiwifruit. Some techniques, including temperature management and controlled atmosphere storage, are often applied to minimize the effects of C₂H₄ on kiwifruit (Hertog et al., 2016; Antunes and Sfakiotakis, 2002). 1-Methylcyclopropene (1-MCP), a potent inhibitor of ethylene perception, delays kiwifruit ripening during shelf life period (Koukounaras and Sfakiotakis, 2007).

Several plant hormones, such as acetylsalicylic acid (Yin et al., 2013) and polyamines (Jhalegar et al., 2012), and certain gaseous molecules, including nitric oxide (NO) (Zhu et al., 2010) and ozone (Minas et al., 2014), also delayed the softening of kiwifruit stored at ambient temperature, mainly through the suppression of ethylene biosynthesis. These results indicate that the softening of kiwifruit at ambient temperature is dependent on C₂H₄.

Molecular hydrogen (H₂) has recently been proposed as a multi-functional signalling gas. In animals, this gas exerts beneficial effects on many physiological and pathological processes, including the protective effects of antioxidant activity, anti-inflammation, and antiapoptosis (Hong et al., 2010). Hence, it acts as a therapeutic agent in biomedical fields, clinical, and in experimental models of many diseases (Hong et al., 2010). In plants, older studies reported the evolution of H₂ in several species (Torres et al., 1984) and the seed germination of Winter rye is induced under an atmosphere of H₂ (Renwick et al., 1964). However, the physiological roles of H₂ have only recently been characterized. For example, by using hydrogen-rich water, H₂ was revealed to orchestrate a wide range of processes in plants, such as the regulation of stomatal closure (Jin et al., 2016) and root development (Zhu and Liao, 2016). H₂ was also proposed to be a broad-spectrum anti-stress molecule, because hydrogen-rich water confers protection against salt (Xu et al., 2013), drought (Xie et al., 2014), paraquat (Jin et al., 2013), and heavy metal stresses (Chen et al., 2014). These results indicate that

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the protective function of H₂ is mainly dependent on the increased antioxidant ability exerted through various regulatory mechanisms. Previous work also suggested that hydrogen-rich water attenuated the senescence of kiwifruit through a decrease in oxidative damage (Hu et al., 2014). The possible role of hypoxia in the abovementioned actions conferred by hydrogen-rich water could not easily be excluded (Xie et al., 2014).

To gain a better understanding of the role of H₂ in increasing the shelf life of kiwifruit, fruit were fumigated with H₂ gas. Endogenous H₂, the relationship between H₂ and C₂H₄, and disease incidence during ripening of kiwifruit were examined. The results reveal a new functional mechanism of H₂ when acting as a positive regulator of the ripening process in kiwifruit by decreasing C₂H₄ production.

2. Materials and methods

2.1. Plant materials

Kiwifruit (*Actinidia deliciosa* cv. 'Xuxiang') were obtained from a commercial orchard (33°59'–34°19'N, 107°39'–108°00'E) in Mei County, Baoji City, Shaanxi Province, China. Fruit used for experiment I were picked on October 30, 2015. At this stage, the initial firmness was 40–60 N. The fumigation method of H₂ in experiment I was repeated in experiment II and III in the following season, with except that harvest occurred on October 20, 2016. At this stage, the initial firmness was 50–70 N. The fruit used for experiments were selected by uniform size, shape, and the absence of mechanical injuries and diseases.

2.2. Experimental design

2.2.1. Experiment I: effects of fumigation with different concentrations of H₂

In total, 660 kiwifruit were selected for the experiment and divided into 11 treatment groups with 60 fruit in each group. The fruit were fumigated with different concentrations of H₂ (0, 2.25, 4.5, and 45 μL L⁻¹) in a sealed plastic container (21 L, Lock & Lock). Different H₂ concentrations were obtained from a cylinder that contained 1000 ± 50 μL L⁻¹ H₂ in nitrogen (Nanjing Special Gas Factory Co., Ltd. Nanjing, China), and injected into the container by a syringe through an injection port on the container. Fruit were fumigated in an atmosphere containing H₂ for 24 h at ambient temperature (25 ± 1 °C). Control fruit were placed in a similar container for an identical incubation duration without any H₂. During fumigation, the accumulated CO₂ in each chamber was absorbed by 15 g CO₂ absorbent (Calcium hydroxide, purchased from Shanghai Guang shao Metallurgical Technology Co., Ltd. Shanghai, China).

To optimize the treatment conditions, given the above restraints, the effects of two different treatment modes [continuous fumigation for 24 h (with 24 h defined as a one-stage treatment) and continuous fumigation for 12 h, treated again, and continuous fumigation for another 12 h (12 h + 12 h, defined as a two-stage treatment)] on the quality of the fruit were simultaneously compared.

Fumigation with 1.0 μL L⁻¹ 1-methylcyclopropene (1-MCP) was used as the positive control, owing to its established beneficial effect in delaying the senescence of kiwifruit (Koukounaras and Sfakiotakis, 2007), and exogenous C₂H₄ fumigation treatment at different concentrations (0.5, 1.0, and 1.5 μL L⁻¹) was used as the negative control. The concentration of required exogenous ethylene was obtained through the injection of an appropriate amount of pure ethylene (99.99%, Nanjing Special Gas Factory Co., Ltd. Nanjing, China) into the experimental container. After the gas within the chamber was circulated using a small fan, the ethylene concentration inside the container was determined by gas chromatography (Li et al., 2017b).

All treated fruit were stored at 25 ± 1 °C. After a shelf life of 9 d, 60 fruit subjected to each treatment (three replicates of 20 fruit each) were sampled, and flesh firmness was determined to choose the most

suitable concentration and mode of H₂ gas treatment.

2.2.2. Experiment II: effects of postharvest H₂ fumigation on C₂H₄ biosynthesis and kiwifruit quality

To confirm the roles of H₂ on kiwifruit C₂H₄ biosynthesis, a separate experiment was conducted. As described in experiment I, fruit were exposed to 4.5 μL L⁻¹ H₂ (two-stage, based on above result) in an air-tight 21-L chamber for 24 h at 25 ± 1 °C. Fruit sealed in the same containers for 24 h were used as the control. After treatments, the fruit were stored at 25 ± 1 °C to allow ripening to proceed. At 3 d intervals, 36 fruit from each treatment (each replicate contained 12 fruit) were randomly sampled and stored at -70 °C prior to various experimental assays, and fresh samples were used for the determination of firmness, C₂H₄ production, and cellular structure.

2.2.3. Experiment III: treatment of exogenous C₂H₄ (ACC) or H₂, and co-treatment of C₂H₄ + H₂ and ACC + H₂

The combination of C₂H₄ + H₂: Fruit were sealed in air-tight containers for 24 h and exposed to H₂ (4.5 μL L⁻¹, two-stage), C₂H₄ (1.0 μL L⁻¹), or a co-treatment of C₂H₄ (1.0 μL L⁻¹) + H₂ (4.5 μL L⁻¹, two-stage). Control fruit were placed in a similar container for an identical incubation duration in the absence of H₂ or C₂H₄, as described in experiment I.

The combination of ACC + H₂: Fruit were submerged in 5 mmol L⁻¹ ACC and distilled water (control) for 5 min and the fruit were air-dried. The combined treatment consisted of fruit that were either treated by ACC or distilled water and fumigated with H₂.

Subsequently, fruit were stored at 25 ± 1 °C and 85–90% relative humidity (Testo 177H1, Germany) for 9 d to simulate the ripening behavior during shelf life. At 3 d intervals, 36 fruit from each treatment (each replicate contained 12 fruit) were randomly sampled and used for the determination of C₂H₄ production and flesh firmness.

2.3. H₂ production

The endogenous H₂ level from fresh pulp tissue was determined by a previously described chromatographic system (Gas chromatography 7890, Agilent Technologies Inc., USA) (Jin et al., 2013), with minor modifications. The gas chromatography was equipped with a thermal conductivity detector (TCD) and a column containing a Molecular Sieve 5 Å stationary phase. Fresh pulp samples were pulverized in liquid nitrogen by using a cryomill (IKA A11, Germany). Approximately 0.5 g of the frozen powder was homogenized with 7 mL distilled water for 2 min and placed in a vial; 5 μL octanol and 0.5 mL 5 mol L⁻¹ H₂SO₄ were added. Afterwards, pure N₂ gas (99.99%) was bubbled into the vial by a nitrogen blowing instrument (An Jian Beijing Co., Ltd. Beijing, China) to fully displace the air and the vial was capped and shaken vigorously for 2 min. Subsequently, the vial was heated at 70 °C for 1 h to liberate H₂ from plant tissues and allowed to cool at room temperature before the analysis of a 100-μL sample of the headspace gas.

2.4. Firmness

Flesh firmness was determined on two opposite sides at the equator of each fruit with a TMS-Touch texture analyzer fitted with 10-mm flat probe (Food Technology Corporation, USA). The compression test was performed by using a trigger force of 0.7 N, a test speed of 1 mm s⁻¹, and a penetration depth of 8 mm.

2.5. Transmission electron microscopy (TEM)

The morphological examination of flesh tissues by TEM was conducted as described by Hu et al. (2014).

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