



Effects of post-partum administration of ketoprofen on sow health and piglet growth



Elina Viitasaari^{a,*}, Laura Hänninen^a, Mari Heinonen^a, Marja Raekallio^b, Toomas Orro^c, Olli Peltoniemi^a, Anna Valros^a

^a Department of Production Animal Medicine, P.O. Box 57, University of Helsinki, Helsinki 00014, Finland

^b Department of Equine and Small Animal Medicine, P.O. Box 57, University of Helsinki, Helsinki 00014, Finland

^c Department of Animal Health and Environment, Institute of Veterinary Medicine and Animal Science, Estonian University of Life Sciences, Tartu 51014, Estonia

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ABSTRACT

The effect of the non-steroidal anti-inflammatory drug ketoprofen on the post farrowing phase of sows was studied in a randomized, blinded, placebo-controlled trial. Ketoprofen (3 mg/kg) was administered intramuscularly to 20 healthy sows for 3 days post-partum (p.p.). The control group ($n = 20$) received a saline placebo. Backfat, number of days of constipation and days before feed refusal were measured. Body condition (BCS) and shoulder sores were scored for 1 week p.p. Changes in BCS, backfat and shoulder sore scores were analysed with ANOVA. Blood was collected on days -1 , 0 , 5 and 14 with respect to medication. Aspartate aminotransferase (AST), creatinine kinase (CK), haptoglobin and serum amyloid A (SAA) were quantified and analysed with a Mann–Whitney U test.

BCS and backfat decreased less following ketoprofen administration than with the placebo (-0.08 ± 0.2 vs. -0.8 ± 0.2 , 1.0 ± 0.8 mm vs. -2.0 ± 0.9 mm, respectively; $P < 0.05$ for both) during the first 2 weeks of lactation. The shoulder sore score deterioration was milder during days 4–6 p.p. with ketoprofen than placebo ($P < 0.05$). Duration of constipation was shorter with ketoprofen than placebo (5.5 ± 0.3 vs. 6.4 ± 0.3 days p.p.; $P < 0.05$). Incidences of feed refusal occurred later in the ketoprofen group than in the placebos (9.6 ± 0.9 vs. 3.8 ± 0.8 days p.p.; $P < 0.05$). AST and SAA values were higher after ketoprofen administration than placebo on day 5 p.p. ($P < 0.05$). It was concluded that ketoprofen appeared to benefit sows during the first 2 weeks post farrowing, but caused some tissue irritation.

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Introduction

No information appears to be available in the literature on pain associated with farrowing and early lactation of sows, or on the effect of pain on health and body mass. Feed refusal has been shown to be a component of pain and sickness behaviour (McGlone et al., 1993; for review, see Weary et al., 2008) and led to deterioration in body condition. This can accelerate after 2 weeks post-partum (p.p.) (Valros et al., 2003). Sows with poor body condition score (BCS) are at increased risk of developing decubital shoulder sores (Tantasuparuk et al., 2001; Bonde et al., 2004; Maes et al., 2004). Gorecki et al. (2010) reported that human patients with a similar type of pressure ulcer experienced pain, which has not been evaluated in pigs.

Low bodyweight (BW) has been shown to indicate poor reproductive success, and is a major reason for culling sows (Tantasuparuk et al., 2001; Maes et al., 2004; Anil et al., 2008). It is therefore important to maintain sow BW during lactation. Feed

refusal can lead not only to poor BW but also to constipation, which is a major cause of abdominal pain in children (Loening-Baucke and Swidsinski, 2007), and may also cause pain in sows. Oliviero et al. (2010) reported that severe constipation is linked to prolonged farrowing. Stressful delivery also delays breast filling during early lactation, contributing to nursing difficulties in women (Chen et al., 1998). Piglet growth depends on nursing (Valros et al., 2002) and problems in lactation consequently affect piglets.

Parturition can cause external symptoms of malaise with tissue breakdown and inflammation. Tissue damage can be detected as increased serum aspartate aminotransferase (AST) and creatinine kinase (CK) activity, which are both released from damaged muscle cells (Nogueira et al., 2000). Acute phase proteins (APPs), such as haptoglobin (Hp) and serum amyloid A (SAA) concentrations increase during the inflammatory processes (Chen et al., 2003; Heinonen et al., 2010), and during the puerperium period (Verheyen et al., 2007; Papadopoulos et al., 2009).

Ketoprofen is a non-steroidal anti-inflammatory drug (NSAID), and has been approved for use in pigs (EMEA, 1996). It has antipyretic, anti-inflammatory and analgesic effects, and the

* Corresponding author. Tel.: +358 50 4156657.

E-mail address: elina.viitasaari@helsinki.fi (E. Viitasaari).

recommended dose is 3 mg/kg BW (EMEA, 1996). Ketoprofen is absorbed well and reaches peak plasma concentration rapidly after intramuscular (IM) administration (Raekallio et al., 2008; Fosse et al., 2011a). Ketoprofen products for veterinary use are racemic compounds but it is the S(+) isomer that is predominant in pigs after administration (Neirinckx et al., 2011; Mustonen et al., 2012). Ketoprofen is effective for treating pain in lame sows (Mustonen et al., 2011) and sows with respiratory infections (Swinkels et al., 1994).

Field observations suggest that sows that have farrowed within 24 h benefit from the use of analgesics. Our study investigated the effect of post-partum (p.p.) administration of ketoprofen on sow feeding, BCS, appearance of shoulder sores and duration of constipation. In addition, we studied the effect of ketoprofen on AST, CK, SAA and Hp. Our hypothesis was that when administered p.p. to sows ketoprofen helps maintain good body condition, reduces risk of shoulder sores, decreases duration of constipation, reduces inflammation and improves piglet performance.

Materials and methods

The experimental procedures were approved by the National Animal Experiment Board (ESAVI-2010-09747/Ym-23, PH22A) and FIMEA, the Finnish Medicines Agency (Vetki-nro 03/10).

Animals and housing

We performed a double blind, placebo-controlled clinical field trial on 40 sows in a commercial piglet producing farm in Western Finland. The sows were moved to farrowing crates approximately 7 days prior to expected farrowing. The crates had fully slatted floors with a heat plate and a heat lamp for the piglets. The sows received a handful of fresh straw every day. Lactating sows were given a commercial liquid feed five times daily, with an energy content starting from 17.6 MJ on day 1 of lactation, and increasing up to 128.0 MJ by day 18. Water was available freely from a water nipple. Farrowings were monitored during piggery working hours (from 06:00 to 15:00 h) and assistance was provided when needed during working hours.

When all piglets had been born and placentas expelled, the piglets were isolated from their mothers by a low fence either for approximately 1.5 h as soon as farrowing was considered over, or in the following morning at 06:00 h if farrowing had taken place at night. During that separation period the sow received an injection of 7–10 IU oxytocin to improve colostrum excretion. Litters were adjusted during the first day of life by merging equal sized piglets into a litter. A routine health check was performed daily by the carers from farrowing to weaning. Any obvious signs of disease or injury were recorded.

Medication

The sows were randomly allocated to two groups both containing 20 sows. The treatment group (KET) received 3 mg/kg ketoprofen (Ketovet 100 mg/mL, Richter Pharma) IM and the placebo group (PLAC) received an equal volume of isotonic saline. The first administration was given when the piglets were isolated by the fence, and was repeated once daily for a further 2 consecutive days.

Body condition score, backfat thickness and assessment of shoulder sores

BCS was assessed and backfat thickness measured for each sow at farrowing, on day 14 p.p. and at weaning (average 23 days). A scale of 1–5 (1 = thin, 2 = decent, 3 = good, 4 = very good, and 5 = fat) (Bonde et al., 2004) was used to gauge BCS. Backfat was measured with a digital backfat indicator (Renco Leanometer) at 70 mm behind the ribcage and from the spine. Shoulders were scored daily from day 0 to day 6 p.p. The severity of the shoulder lesions was classified on a scale of 0–3 according to Rolandsdotter et al. (2009); 0 = no mark of lesion, 1 = redness on the skin, 2 = redness of the skin and a small wound, 3 = deep and large wound, surrounding skin darkened.

Feeding behaviour and constipation

Each trough was checked every morning from farrowing to weaning. Feed refusal was recorded when the sow had left approximately half or more of her feed in the trough. The first day of feed refusal was recorded. Faeces of all sows were assessed daily from day 0 to day 7 p.p. and scored according to Oliviero et al. (2010): 0 = absence of faeces, 1 = dry, small number of unformed and pelleted

faeces, 2 = pelleted but shaped dry faeces, 3 = normal, firm and soft faeces, 4 = normal but not firm soft faeces, 5 = watery, unformed faeces. Faeces scores of 0 and 1 indicated constipation.

Piglet performance

Piglets were weighed by litter on the day of farrowing (day 0) after litter adjustment, on day 14 and at weaning. The number of piglets was recorded at each weighing and average daily weight gains (ADG) were calculated.

Blood sampling

Using an 18 G needle blood (10 mL, uncoated tubes) was collected via the saphenous, coccygeal or medial auricular veins from each sow for analysis of AST, CK, SAA and Hp on four occasions: 1 or 2 days prior to expected farrowing, at day 0 (after farrowing) and at days 5 and 14 p.p. The serum was separated by centrifugation (1900 g for 15 min), and stored at -20°C until analysed.

Laboratory analyses

Serum Hp was analysed using a haemoglobin-binding assay developed for cows (Makimura and Suzuki, 1982) with modifications, in which tetramethylbenzidine was selected as a substrate (Alsemgeest et al., 1994) and 5 μg of sample volume were used (originally 20 μg). The assay was adapted for microtitration plates and optical densities (OD) of the wells were read at 450 nm using a spectrophotometer (Multiskan MS, Labsystems). Pooled and lyophilized aliquots of porcine acute phase serum were used to create standard curves by serial dilutions. The standard curve range was 254–4060 mg/L. The assay was calibrated using a porcine serum sample of known Hp concentration provided by the European Commission Concerted Action Project (number QLK5-CT-1999-0153).

Serum concentrations of SAA were measured using a commercially available ELISA kit (Phase SAA Assay, Tridelta Development), according to the manufacturer's instructions for pigs. Initially a serum dilution of 1:500 was used for all samples. The activities of CK and AST were assessed using a clinical chemistry analyser (Konelab 30i, ThermoFisher Scientific).

Statistics

Changes in backfat and BCS were calculated between farrowing (day 0) and day 14 (0–14), day 14 and weaning (14–WEA), and from farrowing to weaning (0–WEA). Shoulder sore scores were calculated separately for changes between day 0 and all 5 consecutive days. Sows were classified according to their parity as belonging to either parity 2–5 ($n = 26$) or parity 6–9 ($n = 14$). ADG was also calculated 0–14, 14–WEA and 0–WEA.

The effect of treatment (KET and PLAC) and parity on ADG and changes in BCS, backfat and shoulder sore score, total days of constipation and first appearance (days p.p.) of feed refusal, were analysed using a two-way ANOVA. Fixed factors were treatment, parity and their interaction. Total piglet mass growth was used as a covariate for the analysis of changes in BCS and backfat, and total piglet mass growth 0–14 was used as a covariate in the models analysing the number of constipation days and the first day of feed refusal. Backfat on day 0 was used as a covariate in the models for shoulder sore score change, and BCS on day 0 in the model for analysing treatment effect on ADG.

The normality and homogeneity assumptions of the models were checked with a normal probability plot of residuals and scatter plot residuals against fitted values. Residuals in blood parameters failed to conform to a normal distribution and a Mann–Whitney U test was used to analyse the overall effect of treatment on blood parameters, separately for each sampling day, and for both parity classes. All the ANOVA results are presented as model estimates and standard errors of means (mean \pm SEM), and results for non-parametric Mann–Whitney U tests as medians (range). The upper limit for the statistically significant effect was set to $P < 0.05$. All statistical analyses were conducted using PASW Statistics 18.0.1.

Results

Changes in BCS between sampling points was smaller in the KET sows than in the PLAC sows in period 0–14 (Fig. 1) but not at any other interval. However, there was an interaction between treatment and parity ($P = 0.002$). No difference was detected in parity 2–5 sows for BCS loss between treatments, but parity 6–9 KET sows had lower losses in BCS 0–14 than PLAC sows from the same parity group (Fig. 2). KET sows gained thickness in backfat, but PLAC sows lost thickness (Fig. 3). Interaction between treatment and parity showed no effect ($P = 0.08$) for changes in backfat 0–14.

Treatment affected shoulder sore score changes between days 0–3 and 0–4 ($P = 0.006$ for both) and 0–5 ($P = 0.02$, Fig. 4). Treat-

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