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Short communication

Genetic conservation of potentially immunogenic proteins among Brazilian isolates of *Babesia bovis*

Carlos A.N. Ramos^{a,b,*}, Flábio R. Araújo^b, Leucio C. Alves^a, Ingrid Ieda Fernando de Souza^{b,c}, Daniel S. Guedes Jr^d, Cleber Oliveira Soares^b

^a Laboratório de Doenças Parasitárias dos Animais Domésticos, Departamento de Medicina Veterinária – UFRPE, Recife, PE 52171-900, Brazil

^b Embrapa Gado de Corte, Campo Grande, MS 79002-970, Brazil

^c UFMS, Campo Grande, MS 79070-900, Brazil

^d FIOCRUZ, Bio-Manguinhos, Rio de Janeiro, RJ 21040-360, Brazil

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ABSTRACT

Bovine babesiosis caused by *Babesia bovis* remains an important constraint for the development of cattle industries worldwide. Effective control can be achieved by vaccination with live attenuated phenotypes of the parasite. However, these vaccines have a number of drawbacks, which justifies the search for better, safer vaccines. In recent years, a number of parasite proteins with immunogenic potential have been discovered. However, there is little information on the genetic conservation of these proteins among different parasite isolates, which hinders their assessment as immunogens. The aim of the present study was to evaluate the conservation of the genes *ama*-1, *acs*-1, *rap*-1, *trap*, *p*0 and *msa*2c among five Brazilian isolates of *B. bovis*. Through polymerase chain reaction, genetic sequencing and bioinformatics analysis of the genes, a high degree of conservation (98–100%) was found among Brazilian isolates of *B. bovis* and the T2Bo isolate. Thus, these genes are worth considering as viable candidates to be included in a recombinant cocktail vaccine for cattle babesiosis caused by *B. bovis*.

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1. Introduction

Bovine babesiosis is a tick-borne disease that imposes important constraints on livestock health and economic development in tropical and subtropical regions throughout the world (McCosker, 1981). In Brazil, economic losses due to tick fever are on the order of 500 million dollars annually (Grisi et al., 2002) and most losses are likely due to infection by *Babesia bovis*, as this species is the aetiological agent of most outbreaks of babesiosis (Rodrigues et al., 2005; Almeida et al., 2006; Antoniassi et al., 2009; Câmara et al., 2009). *B. bovis* infection is characterised by fever, anaemia, jaundice, sequestration of infected erythrocytes in the host microvasculature, hypotensive shock and often the death of infected animals (Wright and Goodger, 1988).

Vaccination against *B. bovis* is a widespread method used to diminish the impact of clinical disease (Nari et al., 1979; Kessler et al., 1987; Callow et al., 1997; De Vos and Bock, 2000; Ojeda et al., 2010). However, the available vaccines are based on bovine blood infected with live attenuated organisms (Nari et al., 1979; Kessler et al., 1987). Despite inducing a strong protective immune response in vaccinated animals, these vaccines have limitations, such as the transmission of other blood-borne pathogens, failure due to vaccine breakdown (De Vos and Bock, 2000), the possibility of reversion to virulence and a short shelf life in the case of chilled vaccines. Moreover, there are important ethical implications related to the production of a live



^{*} Corresponding author at: Embrapa Gado de Corte, BR 262, Km 04, CP 154, Campo Grande, MS 79002-970, Brazil. Tel.: +55 67 33682085.

E-mail address: carlosanramos@yahoo.com.br (C.A.N. Ramos).

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vaccine due to the need for artificially infected cattle in order to obtain large volumes of infected blood (Kessler et al., 1987; Callow et al., 1997).

The search for better, safer vaccines has been based on the investigation into novel immunogenic proteins that can provide a good level of protection and better safety in comparison to current vaccines. Recombinant DNA techniques have enabled the production of many such proteins with immunogenic potential (Gaffar et al., 2004a,b; Norimine et al., 2006).

Several genes that encode *B. bovis* proteins with immunogenic potential are currently known, especially after the sequencing of the genome of this protozoan (isolate T2Bo, Texas, USA) (Brayton et al., 2007). Some of these antigens were pointed as potentially immunogenic proteins, based on fractioning of merozoite proteins or culture supernatants and testing of individual fractions tested for induction of protective immunity in cattle or animal models; antibody or T-cell-proteomic approaches; genomic approach or the combination proteomic–genomic approach, as reviewed by Brown et al. (2006).

However, information on the conservation of genes that encode remains scarce (Perez et al., 2010). Such knowledge is important, as various proteins may elicit protective responses only against homologous isolates and outbreaks of the disease may involve the challenge of heterologous isolates.

The aim of the present study was to evaluate the conservation of genes *ama*-1, *acs*-1, *rap*-1, *trap*, *p*0 and *msa*2c, that encode potentially immunogenic proteins among different Brazilian strains of *B. bovis*.

2. Material and methods

2.1. Isolates

Five Brazilian isolates of *B. bovis* from the states of Bahia, São Paulo, Rio Grande do Sul, Mato Grosso do Sul and Rondônia were used, representing all physiographic regions of Brazil: Northeast (NE), Southeast (SE), South (S), Midwest (MW) and North (N), respectively (Kessler et al., 1998).

2.2. DNA extraction

Genomic DNA from Brazilian isolates of *B. bovis* was obtained from $350 \,\mu$ L of infected bovine blood using the Easy DNA kit (Invitrogen Carlsbad, CA, USA), following the manufacturer's instructions. The integrity and concentration of DNA samples were assessed by agarose gel electrophoresis 0.8% and NanoDrop ND1000 (Thermo Scientific, Waltham, MA, USA).

2.3. Gene amplification and cloning

For the sequence analysis, whole open reading frames of the genes *ama*-1, *acs*-1, *rap*-1, *trap*, *p*0 and *msa*2c from the five Brazilian isolates were amplified by polymerase chain reaction (PCR) and cloned with pGEM-T Easy (Promega Co., Madison, WI, USA), following the manufacturer's instructions. Amplifications of the whole genes were performed in different steps using specific primer sets designed to amplify \sim 500 bp fragments with overlapping ends (Supplementary File 1).

All amplification reactions were performed in a volume of $25 \,\mu$ L, containing 10 mM of Tris–HCl (pH 8.3), 50 mM of KCl, 1.5 mM of MgCl₂, 0.2 mM of each deoxynucleoside triphosphate, 12 pmol of each primer, 100 ng of genomic DNA and 1.25 U of *Taq* DNA polymerase (Invitrogen, Carlsbad, CA, USA). Amplifications were performed in a Mastercycler thermocycler (Eppendorf, Hamburg, Germany) as follows: $94 \,^{\circ}$ C for 1 min (denaturation); 30 cycles of $94 \,^{\circ}$ C for 1 min (denaturation); and a final cycle at 72 $\,^{\circ}$ C for 4 min (extension). PCR products were analysed by electrophoresis on 1% agarose gel stained with SybrGold (Invitrogen, Carlsbad, CA, USA).

2.4. Gene sequencing and analysis

The DNA sequences of the genes *ama*-1, *acs*-1, *rap*-1, *trap*, *p*0 and *msa*2c from the five Brazilian isolates of *B. bovis* were obtained using the BigDye Terminator kit and ABI3130 sequencing analyzer (Applied Biosystems, Foster City, CA, USA). For each gene studied at least five different amplicons, obtained from different PCR reactions, were sequenced. Sequences were assembled using the Sequencher v.4.1.4 software program (Gene Codes, Ann Arbor, MI, USA) and submitted to BLASTn search (http://www.ncbi.nlm.nih.gov) to determine the sequence identity. Multiple sequence alignment was performed with the ClustalW algorithm (www.ebi.ac.uk/Tools/clustalw2/index.html).

3. Results and discussion

The gene sequences of the Brazilian isolates of *B. bovis* were deposited in the Genbank under the accession numbers shown in Table 1.

The multiple alignment revealed levels of identity from 98% to 100% among the Brazilian isolates and between these isolates and the T2Bo isolate, the genome of which is available in the Genbank under the accession number NZ_AAXT00000000. The most conserved genes were *ama*-1, *p*0, *acs*-1 and *msa*2c, which exhibited 99–100% identity among the Brazilian isolates and between the American (T2Bo) and Brazilian isolates. The percentages of genetic identity among the Brazilian and American isolates of *B. bovis* are displayed in Table 2.

The alignment of the *acs-1* gene with the mRNA sequence (Genbank AF331454) allowed the identification of three introns between nucleotides 113 and 149, 232 and 265 and between 393 and 427.

On the amino acid level, proteins encoded by the genes analysed exhibited identities higher than 96.8%, with *p*0 exhibiting the highest identity (100%) among the Brazilian and T2Bo isolates. The deduced amino acid sequence of the *trap* protein exhibited the lowest identity (96.8%) among the isolates analysed.

The research and development of subunit vaccines have reached significant levels in recent years, including encouraging results, as in the case of protozoan *Leishmania chagasi* Download English Version:

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