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Experimental and Toxicologic Pathology

journal homepage: www.elsevier.de/etp



Napsin A is possibly useful marker to predict the tumorigenic potential of lung bronchiolo-alveolar hyperplasia in F344 rats



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ARTICLE INFO

Article history: Received 27 June 2013 Accepted 5 November 2013

Keywords: Napsin A DHPN NNK Rat Lung Carcinogenesis

ABSTRACT

There are 2 types of bronchiolo-alveolar hyperplasia found in rat lungs. One is 'inflammatory hyperplasia' with a potential to recover in future with removal of the stimulating insult and the other is 'latent tumorigenic hyperplasia' as an independent preneoplastic lesion for adenocarcinoma. In the present experiment, we focused on rat lung bronchiolo-alveolar hyperplasia induced by 4-(methyl-nitrosamino)-1-(3-pyridyl)-1-butanone (NNK), which decreases with time after induction and reverts to normal, or by N-bis(2-hydroxypropyl)nitrosamine (DHPN), with tumorigenic potential to progress to adenoma and adenocarcinoma. Though NNK is a typical carcinogen inducing lung adenocarcinoma in female A/J mice, the tumorigenic potential by NNK in rats is weak. Differences between hyperplasias induced by DHPN and by NNK were here examined immunohistochemically.

Formalin fixed paraffin embedded lung samples with hyperplastic and inflammatory lesions were obtained from rats exposed to DHPN or NNK and from lung inflammation models induced with fine particles like CuO, NiO and quartz. The 19 markers were examined immunohistochemically.

Napsin A, in the inflammatory lesions and hyperplasia induced by NNK, was positive for macrophages and secretions in the alveoli spaces but less so in the walls of the alveoli. In the proliferative lesions including hyperplasia induced by DHPN, strong positive staining for napsin A was observed in the walls of the alveoli. Thus high expression was suggested to be possibly useful for detecting tumorigenic potential of rat lung hyperplasia.

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1. Introduction

Lung cancer is one of the most common cancers in the world, with cigarette smoking regarded as the predominant cause. The risk of lung cancer development remains elevated even after giving up smoking and second-hand environmental tobacco smoke from others is also considered to be a problem (Mitchell and Sanders, 2002). Therefore, identification of potential chemopreventive agents in animals models is important. In rodents, 4-(methyl-nitrosamino)-1-(3-pyridyl)-1-butanone (NNK) and N-bis(2-hydroxypropyl)nitrosamine (DHPN) are well known lung carcinogens, but their effects differ between rats and mice. Previously, in order to establish an appropriate bioassay for detection

of promoting potential to lung tumor after intratracheal instillation of fine particle in rats, sequential histopathological changes were examined after initiation of lung tumorigenesis by 3 intraperitoneal injections (IPs) of 10 mg NNK or 0.1% DHPN in drinking water for 2 weeks and intratracheal instillation (IT) to quartz, as a typical lung toxic agent (Yokohira et al., 2005, 2007, 2009b), into F344 male rats (Yokohira et al., 2009a). Lung proliferating lesions were seen by NNK and by DHPN in rats. However, whereas DHPN induced hyperplasias whose size, number and malignancy increased with time, NNK induced hyperplasia with inflammation proved reversible. Thus their multiplicity was 3.2 ± 2.6 (20 rats) at completion of treatment, but only 0.2 ± 0.4 (20 rats) and 0.8 ± 0.8 (20 rats) at 23 and 30 weeks, respectively. This is in line with the earlier finding that tumorigenic potential of NNK in rats is weak (Yokohira et al., 2009a). In contrast, NNK is well known as a strong lung carcinogen when administered to mice, especially to the A/J strain with high susceptibility to lung tumor induction (Takeuchi et al., 2003, 2006, 2009; Yokohira et al., 2008b). From our data there appear to be 2 types of bronchiolo-alveolar hyperplasia in the rat lungs. One is

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Table 1Antibody for immunostaining in Experiments 1 and 2.

| Antibody | Product code | Company | Antigen retrievala | Dilution | Reaction time (min) |
|----------------------------|----------------|---|--------------------|---------------|---------------------|
| Cell proliferation | | | | | |
| PCNA(PC10) | sc-56 | Santa Cruz Biotechnology, Inc., CA, USA | D | 1:100 | 15 |
| Ki67 | NCL-Ki67p | Leica Microsystems Newcastle Ltd., Newcastle Upon Tyne, United Kingdom | Α | 1:2000 | 15 |
| EGF-R | M 3563 | Dako North America, Inc., CA, USA | C | 1:200 | 15 |
| Cell cycle | | | | | |
| Cyclin D1 | 413531 | Nichirei Biosciences Inc., Tokyo, Japan | В | 1:100 | 15 |
| p27 | M 7202 | Dako Denmark A/S, Glostrup, Denmark | A | 1:400 | 15 |
| p53 | M 7001 | Dako Denmark A/S, Glostrup, Denmark | A | 1:50 | 15 |
| p16 | CINtec p16 | Roche mtm laboratories AG, Heidelberg, Germany | Α | Fully diluted | 15 |
| Tumor producing | | | | | |
| CEA | A 115 | Dako Denmark A/S, Glostrup, Denmark | D | 1:100 | 15 |
| Alveolar epithelium | | , , , , , , , , , , , , , , , , , , , | | | |
| Napsin Å | NCL-L-napsin A | Leica Microsystems Newcastle Ltd., Newcastle Upon Tyne, United Kingdom | D | 1:100 | 15 |
| TTF-1 | M 3575 | Dako North America, Inc., CA, USA | В | 1:200 | 15 |
| SP-A | M 4501 | Dako Japan Co., Ltd., Kyoto, Japan | A | 1:500 | 15 |
| Cell membrane | | | | | |
| Cytokeratin 7 | NCL-CK7-OVTL | Novocastra Laboratories Ltd., Newcastle Upon Tyne, United Kingdom | A | 1:500 | 15 |
| Cytokeratin 20 | NCL-CK20 | Leica Microsystems Newcastle Ltd., Newcastle Upon Tyne, United Kingdom | Α | 1:75 | 15 |
| Endocrine receptor | | · · | | | |
| Estrogen receptor α | NCL-ER-6F11 | Leica Microsystems Newcastle Ltd., Newcastle Upon Tyne, United Kingdom | A | 1:100 | 15 |
| Progesterone receptor | NCL-PGR-312 | Leica Microsystems Newcastle Ltd., Newcastle Upon Tyne, United Kingdom | A | 1:600 | 15 |
| Chromogranin A | A 0430 | Dako Denmark A/S, Glostrup, Denmark | A | 1:100 | 15 |
| Synaptophysin | NCL-SYNAP-299 | Leica Microsystems Newcastle Ltd., Newcastle Upon Tyne, United Kingdom | A | 1:100 | 15 |
| Squamous cell | | - | | | |
| Cytokeratin 34βE12 | M 0630 | Dako Denmark A/S, Glostrup, Denmark | A | 1:50 | 15 |
| Cytokeratin 5/6 | M 7237 | Dako Denmark A/S, Glostrup, Denmark | A | 1:50 | 15 |

^a Antigen retrieval: A: heat treatment with pH7 reagent; B: heat treatment with pH 9 reagent; C: enzymatic treatment; D: non-treatment.

'inflammatory hyperplasia' with the potential for reversion to normal and the other is 'latent tumorigenic hyperplasia' considered as a preneoplastic precursor for adenocarcinoma.

In the present experiment, we focused on comparing rat lung hyperplasias induced by NNK and DHPN immunohistochemically. Experiments were conducted to find suitable marker(s) for prospective tumorigenic and malignant potential of early stage lung hyperplasia. To choose probable markers, preliminary staining of hyperplasias, adenomas and adenocarcinomas induced by DHPN after 30 weeks in F344 rats was performed (Experiment 1). Selected examples were then examined with hyperplasias induced by NNK or DHPN (Experiment 2). Additionally, expression of napsin A was examined in inflammatory lesions in lung induced by fine particles for comparison. Previously, lung toxicity of fine particles from various materials was examined in our in vivo bioassay using the IT method (Yokohira et al., 2007, 2008a, 2009b). With the same dose of 2 mg/rat IT, fine particles of quartz, CuO and NiO all caused severe toxicity with severe inflammatory changes featuring neutrophil infiltration and edema.

2. Materials and methods

2.1. Animals

Experimental animals (Experiments 1 and 2) were maintained in the Division of Animal Experiments, Life Science Research Center, Kagawa University, according to the Institutional Regulations for Animal Experiments. All of the animals were housed in polycarbonate cages with white wood chips for bedding and given free access to drinking water and a basal diet, CE-2 (CLEA Japan Inc.,

Tokyo, Japan), under controlled conditions of humidity ($60 \pm 10\%$), lighting (12-h light/dark cycle) and temperature (24 ± 2 °C).

2.2. Experiment 1. Analysis of various markers for the lung proliferative lesions

2.2.1. Tissue samples

Formalin fixed paraffin embedded (FFPE) lung samples including neoplastic lesions (hyperplasia, adenoma and adenocarcinoma) were obtained with the rat DHPN induced lung carcinogenesis model (Yokohira et al., 2009a). Briefly, male 6-week old F344/DuCrlCrlj rats (Japan Charles River, Inc., Kanagawa, Japan) were given 0.1% DHPN (Nacalai Tesque Inc., Kyoto, Japan) in drinking water for 2 weeks, and sacrificed at week 30. At autopsy, the lungs were removed and the lungs, including the trachea and heart, were infused from the trachea with 10% phosphate buffered formalin and then rinsed in 10% phosphate buffered formalin. The lungs were immersed in 10% phosphate buffered formalin for approximately 48 h, and slices were routinely processed for embedding in paraffin for histopathological examination. The method to develop FFPE lung samples from harvested lungs was the same in Experiments 1 and 2.

2.3. Experiment 2. Validation of the selected markers from Experiment 1

2.3.1. Tissue samples

FFPE lung samples with hyperplastic lesions and inflammatory lesions were obtained with rat DHPN or NNK (Toronto Research Chemicals, ON, Canada) induced lung carcinogenesis models on week 12 and 30 (Yokohira et al., 2009a), respectively, and with

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