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### 1 Triterpene saponins from the roots of *llex pubescens*

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#### 33 1. Introduction

Ilex pubescens Hook. et Arn., under the Chinese name 34 'Mao-dong-qing', is an evergreen shrub belonging to 35 36 Aquifoliaceae. Its roots are commonly used as Chinese 37 herbal medicine for treatment of cardiovascular disease and hypercholestaemia in south China [1]. Previous phyto-38 chemical investigation on its roots and leaves led to the 39 isolation of triterpene saponins [2–7], lignan glycosides [8], 40 41 phenylethanols [9,10], and other minor compounds [11–13]. Pharmacological evaluation proved that I. pubescens extracts 42 exhibited a variety of bioactivities, *i.e.*, to enlarge blood vessels, 43 improve microcirculation, ease the blood pressure, inhibit 44 platelet aggregation, prevent thrombosis, and reduce cardiac 45ischemia, as well as radiosensitization effect and so on [14,15]. 46 As part of systematic research on seeking the bioactive 47

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48 constituents from *llex* species [16–24], a phytochemical
49 investigation on the roots of *I. pubescens* was undertaken

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#### ABSTRACT

Five new triterpene saponins, llexpublesnins N–R (**1–5**), along with seven known analogs 14 were isolated from the root of *llex publescens*. Their structures were elucidated on the basis of 15 extensive spectroscopic analysis, including 1D and 2D NMR experiments. llexpublesnin N (**1**) 16 possessed a rare 20-hydroxyursolic acid scaffold from natural resource. These compounds 17 were evaluated *in vitro* for their cytotoxic effects on human cancer cell lines HepG2, HLE, 18 BEL7402, BEL7403, BEL7405, MCF-7, HeLa. Among them, only compounds **5** and **10** showed 19 cytotoxic potentiality against BEI-7403 and HEL cell lines [inhibition (%): 35.38 and 45.12, 20 respectively].

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previously, which led to a revelation of the absolute structure Q3 of llexpublesnins C–M [25]. As an ongoing program toward 51 the discovery of novel bioactive constituents, five new 52 triterpene saponins, namely llexpublesnins N–R (1–5), and 53 seven known ones (6–12) were isolated. This paper describes 54 the isolation and structural elucidation of these compounds 55 (Fig. 1). In addition, the cytotoxic evaluation for these Q4 compounds against the human cancer cell lines HepG2, HLE, 57 BEL-7402, BEL-7403, BEL-7405, MCF-7, and HeLa is included. 58

#### 2. Experimental

#### 2.1. General procedures

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#### 60

HRESIMS were measured on a Bruker APEX IV FT\_MS 61 (7.0T) spectrometer in positive ion mode. Optical rotations 62 were measured on an Autopol-IV polarimeter (RudoPH 63 Research analytical). IR spectra were obtained using a 64 NEXUS-470 FTIR (Nicolet) spectrometer. 1D and 2D NMR 65 spectra were recorded on Bruker Avance DRX-400 and 66 Vnmrs-500 spectrometers. Semi-preparative HPLC was per-67 formed on a Waters model 2487 (Agilent ODS column, 68  $250 \times 10$  mm i.d., 5 µm) with an Alltech evaporative light 69

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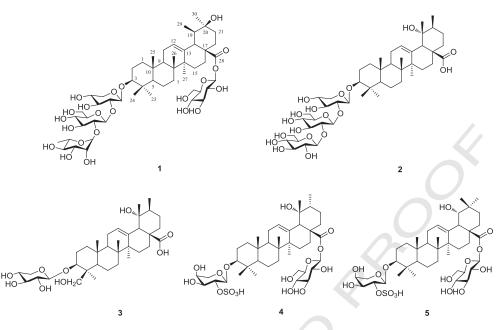


Fig. 1. Structures of compounds 1-5

scattering detector (ELSD). GC analysis was carried out on an 7071 Agilent 6890N gas chromatograph, with an HP-5 capillary 72column (28 m  $\times$  0.32 mm) and an FID detector operated at 73 260 °C (column temp. 180 °C), 1.0 mL/min N<sub>2</sub> as carrier gas. Macroporous resin (HPD100) was purchased from Hebei 74 Bao-En Biotech Ltd.. Thin-layer chromatography and column Q5 76 chromatography were performed using silica gel (Qingdao Haiyang Chemical Co. Ltd., GF<sub>254</sub> or 200-300 mesh) and 77 Sephadex LH-20 (Pharmacia Biotech Ltd.) and ODS (Merck & 78 Co., Inc. USA). All the solvents were of analytical grade and 79 80 were purchased from Beijing Chemical Company Ltd..

#### 81 2.2. Plant material

The roots of *I. pubescens* were purchased from Guilin San-Jin Pharmaceutical Co. Ltd, and were originally collected from Guangxi province, China. The plants were identified by Prof. Peng-Fei Tu (one of the authors in this paper). A voucher specimen (No. 091005) is deposited in the Herbarium of Modern Research Center for Traditional Chinese Medicine (TCM), Peking University, Beijing.

#### 89 2.3. Extraction and isolation

Dry crude materials (18 kg) were grinded and extracted 90 91 with 70% EtOH at the temperature of 70 °C. After the retrieval of the ethanol, the residue suspended in water (50 L) was 92subjected to column chromatography (CC) on macroporous Q6 resin with an EtOH- $H_2O$  gradient (30:70, 70:30, 90:10) 94 to yield three fractions (Frs. 1-3). Fr. 2 (160 g) was 95 chromatographed on silica gel (2.5 kg, 100  $\times$  10 cm) with a 96 gradient of CHCl<sub>3</sub>-MeOH (40:1-1:1) elution to yield thirteen 97 fractions (Frs. A–M). 98

Fr. A (11 g) was recrystallized with mixed solution of MeOH–CHCl<sub>3</sub> (1:1) yields compound **6** (8 g). Fr. C (1.5 g) was subjected to CC on silica gel (50 g,  $15 \times 2$  cm) eluted with 101 CHCl<sub>3</sub>-MeOH (20:1) yields compound 10 (500 mg). Fr. G 102 (25 g) was subjected to CC on silica gel (500 g,  $50 \times 4$  cm) 103 eluted with CHCl<sub>3</sub>–MeOH (10:1, 5:1, 2:1) yields seven fractions 104 (Fr. G-1-Fr. G-7). Fr. G-4 (1.2 g) was chromatographed on 105 Sephadex LH-20 ( $50 \times 3$  cm) in MeOH afforded three 106 fractions (Fr. G-4-1-Fr. G-4-3). Fr. G-4-3 (660 mg) was 107 separated by semi-preparative HPLC (MeOH/H<sub>2</sub>O 80:20, 108 2.0 mL/min) yields compounds 7 (5 mg), 8 (180 mg) and 11 109 (15 mg). Fr. I (3.6 g) was subjected to CC on silica gel (300 g, 110  $50 \times 3$  cm) eluted with CHCl<sub>3</sub>-MeOH (5:1, 2:1) yields two 111 fractions (Fr. I-1–Fr. I-2), Fr. I-1 (2.5 g) was chromatographed 112 on Sephadex LH-20 ( $50 \times 3$  cm) in MeOH, then subjected to 113 RP-C<sub>18</sub> CC (40  $\times$  3 cm) eluted with MeOH-H<sub>2</sub>O (70:30) and 114 further purified by semi-preparative HPLC (MeOH/H<sub>2</sub>O 56:44, 115 2.0 mL/min) yields compound 2 (10 mg). Fr. I-2 (400 mg) was 116 chromatographed on Sephadex LH-20 ( $100 \times 2$  cm) in MeOH 117 and then separated by semi-preparative HPLC (MeOH/H2O 118 65:35, 2.0 mL/min) yields compound 3 (15 mg). Fr. J (18 g) 119 was subjected to CC on silica gel (400 g,  $50 \times 4$  cm) eluted 120 with CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O (60:30:10, low layer) yields three 121 fractions (Fr. J-1-Fr. J-3). Fr. J-3 (2.8 g) was chromatographed 122 on Sephadex LH-20 ( $50 \times 3$  cm) in MeOH afforded three 123 fractions (Fr. J-3-1-Fr. J-3-3). Fr. J-3-3 (1.7 g) was subjected to 124 RP-C<sub>18</sub> CC (40  $\times$  3 cm) eluted with MeOH-H<sub>2</sub>O (70:30) and 125 further purified by semi-preparative HPLC (MeOH/H<sub>2</sub>O 41:59, 126 2.0 mL/min) yields compounds 4 (4 mg), 5 (5 mg) and 9 127 (60 mg). Fr. L (9 g) was chromatographed on RP-C<sub>18</sub> CC 128  $(50 \times 4 \text{ cm})$  using a gradient of MeOH-H<sub>2</sub>O (20:80, 40:60, 129) 60:40, 80:20) afforded five fractions (Fr. L-1-Fr. L-4). Compound 130 1 (10 mg) was obtained from Fr. L-3 (650 mg) by semi- 131 preparative HPLC (MeOH/H<sub>2</sub>O 53:47). Compound 12 (17 mg) 132 was obtained from Fr. L-4 (1.6 g) by Sephadex LH-20 133  $(50 \times 3 \text{ cm})$  and then semi-preparative HPLC (MeOH/H<sub>2</sub>O 134 65:35). 135

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