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# Isocoumarins from American cockroach (*Periplaneta americana*) and their cytotoxic activities



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#### ABSTRACT

Four new isocoumarins (1–4), along with three known ones (5–7), were isolated from the 70% ethanol extract of the whole body of the traditional Chinese insect medicine, American cockroach (*Periplaneta americana*). The structures with absolute configurations of new compounds were elucidated by extensive spectroscopic methods in combination with X-ray diffraction experiment and CD analyses. Compounds **3–5** showed significant cytotoxic activities in HepG2 and MCF-7 cells with IC<sub>50</sub> values in the ranges 6.41–23.91 µM and 6.67–39.07 µM, respectively.

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#### 1. Introduction

The American cockroach, *Periplaneta americana*, is the largest species of pest insect in family Blattidae. *P. americana* is a well-known worldwide domestic pest, which is native to Africa and has spread throughout world especially in the tropical and subtropical regions [1]. In China, the ethanol extract of the dried whole body of *P. americana* has been used as traditional Chinese medicine for the treatment of blood-stasis syndrome, acne and abdominal mass for hundred years [2]. Recent pharmacological studies demonstrated that the crude extract of *P. americana* showed significant anticancer, anti-inflammation and promoting tissue regeneration activities

[3–5]. Previous chemical investigations on *P. americana* mainly focused on the bioactive peptides and enzymes [6–11]. Up to now, however, there is scarcely any literature about its small molecule chemical ingredient. In order to search for the significant bioactivity compounds from P. americana, we carried out a systematical isolation on the 70% ethanol extract of the whole body of P. americana. As a result, four new isocoumarins, periplanetins A-D (1-4), along with three known ones, (3R)-ethyl-6,8-dihydroxy-7-methyl-3,4-dihydroisocoumarin (5) [12], (R)-6-hydroxymellein (6) [13] and (3R)-methyl-7hydroxymethyl-8-hydroxy-3,4-dihydroisocoumarin-6-0-β-D-glucopyranoside (7) [14], were isolated (Fig. 1). Their structures with absolute configurations were established by a combination of NMR, HR-ESI-MS, CD spectra and X-ray diffraction methods. Furthermore, the cytotoxic activities of all isolated compounds on HepG2 and MCF-7 cells were evaluated with the MTT assay. Among them, compounds **3–5** showed significant cytotoxic activities on HepG2 and MCF-7 cells. Herein, the isolation and structural elucidation of these new compounds, as well as the cytotoxic activities of all isolated compounds were described.



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Fig. 1. Chemical structures of 1-7.

#### 2. Experimental

#### 2.1. General

Optical rotations were measured in CH<sub>3</sub>OH on a JASCO P-1020 digital polarimeter at room temperature. Melting point was measured on an X-5 melting point apparatus. UV spectra were measured in CH<sub>3</sub>OH on a JASCO V-550 UV/VIS spectrophotometer with a 1 cm length cell. IR spectra were recorded on a JASCO FT/IR-480 plus Fourier transform infrared spectrometer using KBr pellets. HR-ESI-MS data were obtained on an Agilent 6210 ESI/TOF mass spectrometer and a Waters XeVO G2 Q-TOF mass spectrometer. <sup>1</sup>H, <sup>13</sup>C, and 2D NMR spectra were measured on Bruker AV-400 and AV-500 spectrometers. CD spectra were obtained on a JASCO J-810 spectropolarimeter at room temperature. Column chromatographic separations were performed on silica gel (300-400 mesh, Qingdao Marine Chemical Group Corporation, Qingdao, P. R. China), macroporous resin Diaion HP-20 (Mitsubishi Chemical Corporation, Tokyo, Japan), Sephadex LH-20 (Pharmacia Biotech AB, Uppsala, Sweden), reverse-phase C<sub>18</sub> and C<sub>8</sub> gel (Merck, Darmstadt, Germany). TLC analyses were carried out using precoated silica gel GF<sub>254</sub> plates (Yantai Chemical Industry Research Institute, Yantai, P. R. China). Analytic HPLC was performed on an Agilent chromatography equipped with a G1311C pump and a G1325D diode-array detector (DAD) with a Cosmosil  $5C_{18}$ -MS-II column (4.6  $\times$  250 mm, 5  $\mu$ m, Nacalai Tesque, Kyoto, Japan). Preparative HPLC separations were performed on an Agilent instrument equipped with a G1310B pump and a G1365D UV/VIS detector with a Cosmosil 5C<sub>18</sub>-MS-II column  $(10 \times 250 \text{ mm}, 5 \mu\text{m}, \text{Nacalai Tesque, Kyoto, Japan}).$ 

#### 2.2. Insect material

The dried whole bodies of *P. americana* (killing in hightemperature sterilization conditions) were purchased from Weishan American Cockroach Breeding Base in Dali city, Yunnan province of P. R. China. A voucher specimen (No. 2011052501) was deposited in the Institute of Traditional Chinese Medicine & Natural Products, Jinan University, Guangzhou, P. R. China.

#### 2.3. Extraction and isolation

The dried whole bodies of P. americana (2.5 kg) were powdered and extracted with 70% (v/v) EtOH under percolation twice  $(2 \times 25 \text{ L}, 24 \text{ h} \text{ each})$  at room temperature. The solution was concentrated under vacuum to yield a residue (210 g), which was suspended in H<sub>2</sub>O and subsequently partitioned between CH<sub>2</sub>Cl<sub>2</sub> and H<sub>2</sub>O. The CH<sub>2</sub>Cl<sub>2</sub> extract was evaporated to give a residue (53 g), which was then subjected to silica gel column ( $10 \times 80$  cm) eluted with cyclohexane-EtOAc mixtures (100:0  $\rightarrow$  0:100, v/v) to afford six major fractions (Fr. A–Fr. F). Fr. D (12 g) was subjected to a reverse-phase  $C_{18}$  gel column (3 × 20 cm) eluted with gradient mixtures of MeOH-H<sub>2</sub>O (15:85; 30:70; 50:50; 70:30; 85:15, v/v) to afford five subfractions (Fr. D-1-Fr. D-5). Fr. D-2 (232 mg) was then purified by preparative HPLC on a reversed-phase C<sub>18</sub> column  $(10 \times 250 \text{ mm}, 5 \mu\text{m})$  using MeCN-H<sub>2</sub>O (64: 36, 3 mL/min) as eluent to yield **1** (16 mg,  $t_{\rm R} =$  16.0 min), **3** (9 mg,  $t_{\rm R} =$ 19.3 min), and **4** (19 mg,  $t_R = 18.5$  min). Compounds **2** (7 mg,  $t_{\rm R} = 18.0$  min), **5** (16 mg,  $t_{\rm R} = 20.5$  min), and **6** (24 mg,  $t_{\rm R} = 22.0$  min) were obtained from Fr. D-4 (152 mg) by preparative HPLC using MeOH-H<sub>2</sub>O (73:27, 3 mL/min) as mobile phase. The H<sub>2</sub>O soluble fraction (145 g) was subjected to macroporous resin HP-20 column ( $15 \times 60$  cm) eluted with EtOH-H<sub>2</sub>O (0:100; 35:65; 70:30; 90:10, v/v) to yield four fractions (Fr. a-Fr. d). Fr. b (43 g) was subjected to reversephase C<sub>8</sub> gel column ( $10 \times 80$  cm) eluted with gradient mixtures of MeOH-H<sub>2</sub>O (15:85; 30:70; 50:50, v/v) to afford four subfractions (Fr. b-1-Fr. b-5). Fr. b-3 (2 g) was separated by a Sephadex LH-20 column ( $2 \times 80$  cm, MeOH) to afford **7** (9 mg).

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