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# Human cytochrome P450 epoxygenases: Variability in expression and role in inflammation-related disorders

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# ABSTRACT

Beyond their contribution to the metabolism of xenobiotics, cytochrome P450 (CYP) epoxygenases are actively involved in the metabolism of endogenous substances, like arachidonic acid (AA). The main human CYP 16 epoxygenases, i.e. CYP2C8, CYP2C9, CYP2C19 and CYP2J2, convert AA to four regioisomer epoxyeicosatrienoic 17 acids (EETs). EETs possess a wide range of established protective effects on the human cardiovascular system 18 of which anti-inflammatory actions have gained great recent interest. The expression of CYP epoxygenases is reg-19 ulated through an extremely complex network of nuclear receptors, microRNAs and genetic/epigenetic factors. 20 Accordingly, a large number of biological variables as well as xenobiotics and environmental factors can influence 21 the expression of CYP epoxygenases, resulting in a significant intra- and inter-individual variability in the expres- 22 sion and activity of these enzymes and subsequently in EET biosynthesis. Moreover, human CYP epoxygenases 23 are mainly expressed in the liver; however, these enzymes are also expressed, at various extents, in most extra- 24 hepatic tissues, resulting in a marked inter-tissue variability in the expression of CYP epoxygenases. The inter- 25 tissue, inter- and intra-individual variability in the expression of epoxygenases may lead to differences in the 26 relative abundance of EETs among tissues, among individuals of a population and/or different ethnicities and 27 in a given individual under various conditions. The variation in the abundance of EETs may explain, at least in 28 part, the inter-tissue and inter-individual differences observed in the prevalence of inflammation-related disor- 29 ders including cardiovascular disease, and why in a given individual, various conditions can contribute to the de- 30 velopment of diseases with an important inflammatory component. 31

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*Abbreviations*: AA, arachidonic acid; AhR, aryl hydrocarbon receptor; AP-1, activator protein-1; bZIP, basic leucine zipper; CAR, constitutive androstane receptor; CCRP, cytoplasmic CAR retention protein; CNV, copy number variation; COUPTF, chicken ovalbumin upstream promoter transcription factor; DHA, docosahexaenoic acids; DMet, DNA methylation; EC, endothelial cell; EDHF, endothelium-derived hyperpolarizing factors; EDP, epoxydocosapentaenoic acid; EEQ, epoxyeicosatetraenoic acid; EET, epoxyeicosatrienoic acid; EPA, eicosapentaenoic; FXR, farnesoid X receptor; GR, glucocorticoid receptor; GTF, general transcription factor; HDAC, histone deacetylase; HETE, hydroxyeicosatetraenoic acid; HNF4α, hepatocyte nuclear factor 4α; HSP90, heat shock protein 90; LETF, liver-enriched transcription factor; LPS, lipopolysaccharide; LXR, liver X receptor; miRNA, meroRNA; mRNA, messenger RNA; NADPH, nicotinamide adenine dinucleotide phosphate; NF+κB, nuclear transcription factor; POR, CYP oxidoreductaes; PP-2A, phosphatase 2A; PUFA, polyunsaturated fatty acids; PXR, pregnane X receptor; RC, receptor; RC, receptor; SH, soluble epoxyhydrolase; SHP, small heterodimer partner; SRC-1, steroid receptor; TIF-2, transcriptional intermediary factor 2; TF, transcription factor; TGF, transforming growth factor; VDR, vitamin D receptor.

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## 1. Introduction 49

Inflammation is a complex and highly orchestrated process that 50 51has a domino-like effect on the initiation and development of a wide range of systemic illnesses including cardiovascular disease 52(CVD) (Gabay & Kushner, 1999). Our knowledge about the contri-53bution of inflammation to the development of CVDs has progressed 5455to the point that, currently, coronary artery disease (CAD) and arte-56rial hypertension (HTN) are believed to be diseases of inflammation 57characterized by interaction among platelets, leukocytes and endo-Q3 Q2 thelial cells (ECs) (Harrison et al., 2011; Libby, 2002; May et al., 2007). 59

A growing body of evidence suggests that, beyond their well-60 61established contribution to the metabolism of xenobiotics, cytochrome P450 (CYP) epoxygenase enzymes play an indispensable role in the 62 63 regulation of inflammation through the biosynthesis of endogenous bioactive lipid mediators that modulate inflammation, such as the ara-64 65 chidonic acid (AA) derivatives, epoxyeicosatrienoic acids (EETs) (Deng et al., 2010; Fleming, 2011). 66

Even though human CYP epoxygenases are predominantly 67 expressed in the liver, they are also expressed, on a smaller scale and 68 in a heterogeneous pattern, in extrahepatic tissues (Zanger & Schwab, 04 70 2010), resulting in inter-tissue variability in CYP epoxygenase expression in a given individual. In addition, a large number of factors have 71been demonstrated to affect the expression pattern of CYP genes, 72leading to significant inter- and intra-individual differences in the ex-73 pression levels of CYP epoxygenases. These heterogeneities could po-74 75tentially give rise to a marked inter-tissue, inter- and intra-individual 76variability in the production and relative abundance of EETs and, subse-77 quently, in the EET-mediated inflammation-reduction. Given the im-78portance of inflammation in the pathophysiology of a wide range of CVDs, variability in the anti-inflammatory effects of EETs may have clin-7980 ical consequences.

In this review, we first provide an overview of the structure, activ-81 ity and regulation of expression of human CYPs as well as the mech-82 anism of synthesis and action of CYP-dependent AA metabolites, 83 i.e. EETs. The review then presents in some detail, the factors affecting 84 the expression of CYP epoxygenase genes and resulting in an inter-85 and intra-individual heterogeneity in the epoxygenase-mediated 86 EET formation. Next, we discuss the potential influence of variation 87 in the relative expression of CYP epoxygenase genes in various 88 89 human organs on the local/systemic abundance and biological functions of EETs in a given individual. Lastly, the possible clinical implica-90 91 tion of the inter-tissue, intra-individual and inter-individual variability 92in the epoxygenase expression and the EET production will be discussed. 93

### 2. Cytochrome P450, human epoxygenases 94

## 2.1. Cytochrome P450 enzyme; general considerations 05

CYP enzymes belong to the ubiquitous superfamily of thousands of 96 closely related hemoproteins found throughout the phylogenetic spec-97 trum, from animals, plants and fungi to bacteria. Of more than 11,000 98genes and 1000 CYP families which have been found in nature in total, 99 approximately 115 CYP genes, including 57 putatively active genes 100 and 58 pseudogenes have been identified in human, and are grouped 101 into 18 and 44 families and subfamilies, respectively (http://drnelson. 102uthsc.edu/human.P450.table.html). 103

CYP enzymes share a common structure but also share a confor-104 mational flexibility. In addition to the structural similarities, CYP 105enzymes essentially function in the same way. The structure and 106 catalytic activities of CYP enzymes have been described in detailed, 107 previously (Coleman, 2010; Denisov et al., 2005; Hart & Zhong, 108 109 2008).

## 2.2. Human cytochrome P450 epoxygenases; epoxyeicosatrienoic acid formation

CYP enzymes, particularly those in the CYP1, CYP2 and CYP3 genes 112 families, catalyze the biotransformation of 75% of xenobiotics and phar- 113 maceuticals in human (Sim & Ingelman-Sundberg, 2010; Zanger et al., Q6 2010). In addition, human CYPs play central roles in the overall metab- 115 olism and disposition of a wide range of endogenous chemicals includ- 116 ing steroid hormones (estrogen and testosterone), vitamin D, bilirubin, 117 cholesterol and fatty acids (Fleming, 2011). 118

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AA is a 20-carbon, omega-6 polyunsaturated fatty acid (PUFA) that 119 resides in the cell membrane phospholipids in the stereospecific num- 120 bering (sn)-2 position and is released upon stimulation by the cytosolic 121 enzyme phospholipase A<sub>2</sub> (Sudhahar et al., 2010). The free intracellular 122 AA is converted to a series of biological active metabolites referred to as 123 "eicosanoids" (Imig, 2012). Cyclooxygenases catalyze the biosynthesis 124 of prostaglandins/thromboxanes and lipoxygenases mediate the pro- 125 duction of leukotrienes (Kuehl & Egan, 1980; Zordoky & El-Kadi, 126 2010). Referred to as the "third eicosanoid enzymatic pathway", AA is 127 also metabolized to EETs and hydroxyeicosatetraenoic acids (HETEs) 128 by CYP epoxygenases and CYP hydroxylases, respectively (Oliw, 1994; 129 Zeldin, 2001). 130

CYP epoxygenase enzymes belong to a complex superfamily of genes 131 with a common evolutionary origin, a conserved peptide that provides 132 them with a cysteine heme ligand, and the capacity to take an active 133 form of atomic oxygen to ground state carbons (Imig, 2012). Indeed, 134 CYP epoxygenases catalyze the epoxidation of four olefin bonds of the 135 intracellular AA that gives rise to four corresponding regioisomers: 136 5,6-EET, 8,9-EET, 11,12-EET and 14,15-EET (Spector & Norris, 2007) 137 (Fig. 1). Several CYP subfamilies including CYP1A, CYP2B, CYP2C, 138 CYP2E and CYP2J have been classified as CYP epoxygenases (Imig, 139 2000; Zeldin, 2001), and potentially contribute to the conversion of AA 140 to EETs; however, human epoxygenases are predominantly members 141 of CYP2C and CYP2J classes (Imig, 2000; Zeldin, 2001; Capdevila & 142 Falck, 2002Imig, 2000; Elbekai & El-Kadi, 2006; Zordoky, et al., 2010). Q7 In particular, CYP2C8, CYP2C9, CYP2C19 and CYP2J2 isoforms have 144 been identified as the main CYPs involved in the biotransformation of 145 AA to EETs in human (Enavetallah et al., 2004; Campbell & Fleming, 146 2010; Pfister et al., 2010). Each epoxygenase is, supposedly, capable to 147 convert AA to all four EETs; however, 11,12- and 14,15-EETs are the 148 main products in many cases (Capdevila et al., 2000; Zeldin, 2001). Fur- 149 thermore, there is mounting evidence that human CYP epoxygenases 150 are relatively regioselective for EET formation. For instance CYP2C8 cat- 151 alyzes the biosynthesis of 14,15-EET and 11,12-EET in a ratio of 1.3:1.0 152 whereas its capability for the formation of 8,9-EET and 5,6-EET is negli- 153 gible. Similarly, CYP2C9 also generates 14,15-EET and 11,12-EET in a 154 ratio of 2.3:1.0; this epoxygenase, however, does not form significant 155 amounts of 8,9-EET (Daikh et al., 1994; Zeldin et al., 1995). 08

2.3. Physiological roles of epoxyeicosatrienoic acids in vascular biology 157

There is a large volume of evidence that EETs possess a wide range of 158 potent cardiovascular protective effects including regulation of vascular 159 tone (Harder et al., 1995; Campbell et al., 1996; Fisslthaler et al., 1999; 160 Campbell et al., 2010), and homeostasis (Enayetallah et al., 2004; Q9 Spector et al., 2007; Campbell et al., 2010; Sudhahar et al., 2010; Imig, Q11 2012). Fig. 2 is a schematic illustration of the protective effects and 163 the postulated mechanism of action of EETs on the myocardium and 164 vasculature. It long has been known that EETs act as endothelium- 165 derived hyperpolarizing factors (EDHFs) and exhibit vasodilatory ef- 166 fects in various arterial territories including coronary, renal, mesenteric 167 and cerebral vasculatures (Campbell et al., 2010; Pfister et al., 2010; Q12 Sudhahar et al., 2010; Imig, 2012). In addition, EETs stimulate EC prolif- 169 eration and angiogenesis, protect ECs from apoptosis (Medhora et al., 170 2003; Sudhahar et al., 2010), have anti-migratory effects on smooth 171 muscle cells (SMCs) (Spector et al., 2007; Sun et al., 2002; Sudhahar Q13

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