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- Inulin supplementation during gestation mitigates acrylamide-induced
- maternal and fetal brain oxidative dysfunctions and neurotoxicity
- in rats[☆]
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ABSTRACT

Accumulating evidence suggests that the developing brain is more susceptible to a variety of chemicals. Recent 21 studies have shown a link between the enteric microbiota and brain function. While supplementation of non- 22 digestible oligosaccharides during pregnancy has been demonstrated to positively influence human health 23 mediated through stimulation of beneficial microbiota, our understanding on their neuromodulatory propensity 24 is limited. In the present study, our primary focus was to examine whether supplementation of inulin (a well 25 known fructan) during gestation can abrogate acrylamide (ACR)-induced oxidative impairments and neurotox- 26 icity in maternal and fetal brain of rats. Initially, in a dose-determinative study, we recapitulated the impact of 27 ACR exposure during gestation days (GD 6-19) on gestational parameters, extent of oxidative impairments in 28 brain (maternal/fetal), cholinergic function and neurotoxicity. Subsequently, pregnant rats orally (gavage) 29 administered with inulin (IN, 2 g/kg/day in two equal installments) supplements during gestation days (GD Q6 0-19) were exposed to ACR (200 ppm) in drinking water. IN supplements significantly attenuated ACR-induced 31 changes in exploratory activity (reduced open field exploration) measured on GD 14. Further, IN restored the 32 placental weights among ACR exposed dams. Analysis of biochemical markers revealed that IN supplements 33 effectively offset ACR associated oxidative stress not only in the maternal brain, but in the fetal brain as well. 34 Elevated levels of protein carbonyls in maternal brain regions were completely normalized with IN supplements. 35 More importantly, IN supplements significantly augmented the number of Bifidobacteria in the cecum of ACR rats 36 which correlated well with the neurorestorative effect as evidenced by restored dopamine levels in the maternal 37 cortex and fetal brain acetylcholinesterase activity among ACR-exposed dams. Further, IN supplements also 38 conferred significant protection against mitochondrial dysfunction induced by ACR in both milieus. Although 39 the precise mechanism/s by which IN supplements during pregnancy attenuate ACR induced neurotoxic impact 40 merits further investigations, we hypothesize that it may mediate through enhanced enteric microbiota and 41 abrogation of oxidative stress. Further, our study provides an experimental approach to explore the neuro- 42 protective role of prebiotic oligosaccharides during pregnancy in reducing the adverse impact of developmental 43

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1. Introduction

Recent evidence has revealed that gut microbiota possesses the propensity to influence brain development and behavior. Epidemiological and animal studies have shown the beneficial effects of the proliferation of bacterial strains on brain chemistry (Messaoudi et al., 2011; Collins and Bercik, 2013). Administration of live bacterial

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supplements (probiotics) to rodents is shown to alter trophic cytokine 56 (brain-derived neurotrophic factor, BDNF) gene expression (O'Sullivan 57 et al., 2011). Interestingly, probiotic cocktail reduced the process 58 aligned with anxiety in rats and serum cortisol levels associated with 59 psychological distress in humans (Messaoudi et al., 2011). Although 60 the precise mechanisms mediating these effects remain unclear, the 61 involvement of dampening down of the elevated levels of oxidative 62 free radicals and pro-inflammatory cytokines seems likely (Cryan and 63 O'Mahony, 2011). Hence, dietary interventions using prebiotics 64 to stimulate the growth of intrinsic beneficial intestinal microbiota 65 is being explored as a tool to achieve various beneficial effects to the 66

Fructans, such as inulin are plant derived nondigestible carbohy- 68 drates (NDO) are mainly digested by cecal microbiota (Yasuda et al., 69

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2007) and increase the indigenous bacterial community (Kelly, 2008; van Vlies et al., 2012). Previously, supplementation of non-digestible oligosaccharides during pregnancy, has been reported to ameliorate various metabolic disorders (Qiu et al., 2008). Further, studies have demonstrated a significant diminution in the levels of procarcinogenic biomarkers in rats following prophylactic treatment with inulin (Verma and Shukla, 2013). A recent study reported the potential of inulin in alleviating the LPS-induced oxidative stress in human colon epithelial cell lines (Pasqualetti et al., 2014).

Acrylamide (ACR) is a vinyl, water-soluble alkene used in the production of glue and plastics that have various commercial applications (LoPachin, 2004; Erkekoglu and Baydar, 2014). ACR is demonstrated to be neurotoxic in humans and affects both central and peripheral nervous system. This neurotoxin has gained tremendous importance owing to its formation during frying/baking of commonly consumed foods such as fried potatoes, biscuits and coffee (JECFA, 2011). Studies in rodents have demonstrated that ACR crosses the placenta (Annola et al., 2008; von Stedingk et al., 2011) and causes reproductive/developmental toxicity coupled with adverse effects on the fetus such as lower birth weight, skeletal abnormalities (Manson et al., 2005; El-Sayyad et al., 2011) and neurodevelopmental toxicity (Takahashi et al., 2009; Ferguson et al., 2010; Garey and Paule, 2010; El-Sayyad et al., 2011). The involvement of oxidative stress and inflammatory responses in ACR-induced neurotoxicity are widely accepted (Lopachin and Gavin, 2008; Prasad and Muralidhara, 2012, 2013). Further, ACR is shown to form conjugates with reduced glutathione (GSH), and the resulting complex is metabolized by cytochrome P450 (subtype CYP 2E1) to form glycidamide. Recent evidence suggests that ACR forms adducts within presynaptic proteins resulting in altered neurotransmission and inactivation of enzymes involved in neuronal energy production (LoPachin and Gavin, 2012; Martyniuk et al., 2013).

Compelling evidence exists that chemical agents widely disseminated in the environment are important triggers of neurological anomalies (Grandjean and Landrigan, 2006; Drechsel and Patel, 2008). Developing brain is of particular interest since it is highly susceptible to neurotoxin insult during the critical window of vulnerability. Given the wide industrial applications and the possible neurotoxic effects associated with the chronic human exposure to ACR starting in utero, it is relevant to develop specific therapeutic strategies to abrogate ACR-induced fetal neurotoxic effects. Hence, in the present study our primary objective was to understand whether supplementation of inulin, (a widely consumed NDO) to dams during pregnancy possess the potential to attenuate ACR-induced oxidative stress and neurotoxicity in a rat model. As a prelude, we characterized some of the biochemical perturbations (oxidative impairments and neurotoxicity) in rats associated with gestational exposure to varying doses of ACR. Subsequently we investigated the ameliorative effect of inulin supplements against ACR intoxication in maternal as well as fetal brain.

2. Materials and methods

2.1. Chemicals

Acrylamide (electrophoresis grade; purity > 99%, Product #A8887), inulin from chicory (Product #I2255), 2-thiobarbituric acid, 1,1,3,3-tetramethoxypropane, 2',7'-dichlorofluorescein (DCF), 2',7'-dichlorofluorescein diacetate, N,N,N',N'-tetramethylethylenediamine, L-glutathione reduced, acetylthiocholine iodide, S-butyrylthiocholine iodide, nicotinamide adenine dinucleotide, thiazolyl blue tetrazolium bromide, dopamine were procured from Sigma Aldrich, St Louis, MO, USA. 2,4-Dinitrophenylhydrazine, 1-chloro-2,4,-dinitro benzene, nicotinamide adenine dinucleotide phosphate, cytochrome c were procured from Sisco Research Laboratories Pvt. Ltd., India. All other reagents used were of analytical or HPLC grade.

2.2. Experimental animals

Male and female adult Wistar rats were randomly drawn from the 133 stock colony of the institute's animal house facility (200 \pm 10 g), and 134 acclimatized for 1 week. Rats were housed in vented polypropylene 135 cages $(40 \times 28 \times 16 \text{ cm}^3)$ with dust-free shaved wood bedding. Animals 136 housed in a light- and temperature-controlled room (14 h/10 h light- 137 dark cycle, 21 °C, 50% humidity) were provided with commercial rodent 138 pellet diet (Sai Durga Feeds & Foods, Bengaluru, India) and tap water ad 139 libitum. Following acclimatization, virgin female rats were allowed to 140 mate overnight with males (2:1 ratio). Vaginal smear using 0.9% sodium 141 chloride was performed daily to ascertain pregnancy. The swab was 142 examined on a slide using a bright-field microscope for the presence 143 of spermatozoa. The presence of sperm was designated as gestation 144 day (GD) 0, and the females were separated, weighed and individually 145 housed. All protocols were approved by the institutional animal ethics 146 committee in conformance with the guidelines established by the 147 Committee for the Purpose of Control and Supervision of Experiments 148 on Animals (CPCSEA), Government of India, India. Handling, as well as 149 care of animals during sacrifice, was strictly according to the standard 150 guidelines of the Institutional Ethics Committee (Registration #49/ 151 1999/CPCSEA).

2.3. Experimental procedure

2.3.1. Study 1: exposure of pregnant rats to acrylamide (ACR): Effect on gestational parameters and oxidative markers in fetal and maternal brain 155

Sperm-positive female rats were placed in plastic cages and ran- 156 domly divided into 4 groups. While dams (n = 6) of treatment groups 157 (Groups II, III and IV) received 50, 100 and 200 ppm ACR in drinking 158 water during gestation days (GD) 6 to 19, those of control group 159 (group-I) received deionized water. Water consumption and feed intake were recorded daily while body weight gain was monitored on al- 161 ternate days. On GD 19, all dams were sacrificed, maternal brains and 162 uterine horns were rapidly exteriorized, immersed in ice-cold phos- 163 phate buffered saline (PBS), blotted dry and cryostat cooled as previous- 164 ly described (Shivananjappa and Muralidhara, 2012). Fetuses and 165 placenta were separated from uterine horns after fixing it to a wax 166 base, immersed in ice-cold phosphate buffer saline (PBS). Further, 167 fetal brain were excised from fetuses, rinsed in ice-cold PBS, the meninges were removed and stored at -80 °C until further processing. Similarly, brains were excised from the dams and brain regions viz., cortex 170 and cerebellum were separated on ice and stored until used. Markers 171 of oxidative stress and antioxidant/detoxifying enzymes were deter- 172 mined in the maternal brain regions (cortex and cerebellum) and 173 whole fetal brain.

2.3.2. Study 2: protective effects of inulin supplements in the ACR model Pregnant rats were assigned four groups and treated follows: 176 Group I — Untreated controls; Group II — Inulin (IN); Group III and 177 Group IV – ACR (200 ppm, GD 6–19) in drinking water. Rats of 178 Groups II and IV were orally administered (gavage) with inulin (IN, 179 2 g/kg/day) supplements during gestation days (GD 0-19). The 180 total daily dose of IN was given as split dose (twice a day at the 181 rate of 1 g/kg at 09:00 and 15:00 h each day). The IN dose was 182 selected based on our preliminary study, and the doses were 183 adjusted daily to account for the changes in body weight of the 184 dams. Dams of all treatment groups were subjected to open field 185 test on GD 14 and were sacrificed on GD 19. Further procedures 186 were identical as described in Study 1. Biochemical parameters were 187 determined in the maternal brain (cortex and cerebellum) and fetal 188 brain. The cecum from dams of all treatment groups was collected, 189 and subjected to quantification of Bifidobacteria and Lactobacteria 190 adopting standard micobiological procedures. 191

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