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An electrical conductivity based method of determining the particle deposition rate in air-liquid interface devices

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ABSTRACT

A new in-situ method of determining the particle deposition rate onto cell cultures inside air-liquid interface devices is described. It is based on depositing a surrogate aerosol of salt particles onto the water filled wells of a culture plate while measuring the resulting change in electrical conductivity of the solution in situ, in order to derive the accumulated particle mass. For evaluation purposes, the wells of a sixwell cell culture plate were equipped with custom designed electrodes and calibrated with a series of commercially available standard solutions. After the necessary corrections prescribed by theory, the calibration resulted in an accuracy and comparability between cells of ±3% in terms of measured conductivity. The method was then applied to a specific ALI device consisting essentially of the calibrated six-well culture plate inside an electrostatic cross-flow precipitator, and tested with submicron NaCl aerosol of defined size distribution produced by nebulization of a salt solution. 2 h of particle accumulation were sufficient to accumulate between 30 and 10 μ g of salt per well, depending on the location in the precipitator. Resulting deposition rates varied narrowly between the wells by about 2 ng min⁻¹ cm⁻². Factors affecting the overall accuracy and reproducibility are discussed.

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1. Introduction 42

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Air-liquid interfaces (ALI) are the preferred way of exposing cell 43 cultures in vitro to particles under conditions resembling the 44 45 uptake of aerosols through the respiratory system (Aufderheide, 46 2008). On the other hand they are also more complex to operate than submerged systems (Seagrave, 2005). Aside from having to 47 provide a suitable environment for the stress-free survival of the 48 cell cultures (BéruBé et al., 2009), one needs to determine the effi-49 50 ciency and rate of particle deposition on the cells (Paur et al., 2011) - which depends on particle size and operating conditions - in 51 order to calculate an exact dose (Lenz et al., 2009). This makes 52 an accurate characterisation of such exposure devices quite chal-53 54 lenging (Stevens et al., 2008).

Precipitation efficiencies in ALI devices have typically been 55 inferred from the decrease in aerosol concentration during passage 56 57 through the device, measured with standard on-line methods such 58 as SMPS (Bitterle et al., 2006; de Bruijne et al., 2009; Savi et al., 59 2008; Volckens et al., 2009). While this approach is fast and relatively accurate, it is indirect and cannot distinguish between parti-60 cle deposition onto cell culture surfaces and wall losses in general 61 (Savi et al., 2008; Volckens et al., 2009). 62

Direct measurements of particle deposition onto the relevant surfaces have mostly relied on the use of fluorescent particles, which were either non-soluble such as monodisperse polystyrene spheres, or contained a fluorescent dye soluble in water or organic solvents (de Bruijne et al., 2009; Savi et al., 2008; Stevens et al., 2008; Volckens et al., 2009; Teeguarden et al., 2007). In both cases the actual measurement took place outside the ALI chamber, either by counting particles on the substrate with a fluorescent optical microscope (Savi et al., 2008), or by dissolving them and subsequently measuring the fluorescence level in the solvent with a spectrophotometer (de Bruijne et al., 2009). The former technique is currently the only practiced method to provide a deposition rate in terms of particle number (de Bruijne et al., 2009), while the latter expresses it in terms of mass (Savi et al., 2008) assuming that particle mass and fluorescent signal are proportional (Burry, 2010; Hoffmann, 2012). The accuracy and reproducibility achieved by fluorescence based studies appear to be limited, although it is not always clear to what extent a specific experimental protocol or the ALI device itself contributed to the overall uncertainty. Spectrophotometric analysis is normally done on the entire sample surface and thus provides a true average of the deposition rate, while microscopy based particle counting on a large sample surface would not be economical. However, averages based on a limited number of randomly chosen locations depend heavily on the uniformity of the deposit. On the other hand, spectrophotometric

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H. Wiegand et al./Toxicology in Vitro xxx (2015) xxx-xxx

analysis requires solvent extraction that may become a source of
 potential errors, while particle counting under a microscope is
 quite accurate, especially when automated.

91 Another direct approach described in the literature is based on 92 measurements of the deposited particle mass vs. time via the 93 change in resonant frequency of a vibrating piezoelectric crystal 94 (Mülhopt et al., 2009; Lenz et al., 2009). This quartz crystal 95 microbalance (QCM) technique functions in situ, although with a relatively small "field of view" in the order of 40 mm², with a sen-96 sitivity range of $3-160 \ \mu g \ cm^{-2}$ and a reported repeatability (i.e. 97 98 standard deviation of the deposition efficiency divided by the mean deposition efficiency) of 12% (Lenz et al., 2009). The QCM 99 therefore works best with ALI devices that deposit particles in a 100 small spot. 101

102 In the following we describe a new, accurate method of charac-103 terising the particle deposition rate of ALI type devices in situ, via 104 measurements of the electrical conductivity directly inside the 105 wells of a culture plate. Basically, an inorganic salt aerosol - in 106 our case NaCl - of known particle size distribution and concentration is deposited onto the water filled wells, wherein the particles 107 108 dissolve rapidly. The resulting increase in electrical conductivity 109 can be used to infer the accumulated particle mass and thus an average mass based deposition rate for each well. Once an ALI 110 111 exposure chamber is characterised in this way for a range of parti-112 cle sizes, the deposition rate for arbitrary aerosols of known size 113 distribution and concentration can be calculated. Below we first 114 describe the measurement technique including relevant fundamentals of ion conductivity measurements. As an illustration we 115 then operate it in a model electrostatic precipitator equipped with 116 117 a six-well culture plate, in order to demonstrate the properties of 118 this technique.

119 2. Materials and methods

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2.1. The relation between electrical conductivity and electrolyteconcentration

122 The electrical conductivity κ of an ionic solution is basically 123 determined by the concentration and mobility of charge carriers 124 in the liquid. It depends on the viscosity and the dielectric proper-125 ties of the solvent, the type, charge and activity concentration of 126 the dissolved ions, as well as the geometry of the electrodes 127 between which charge carriers are transported (Czeslik et al., 128 2009; Atkins and de Paula, 2009).

The conductivity κ of an ionic solution is the product of the molar conductivity Λ_m and the electrolyte concentration c

$$\kappa = \Lambda_m \cdot c. \tag{1}$$

It is common knowledge that the molar conductivity A_m is not 134 independent of the concentration *c*, i.e. the ration of κ/c is not a 135 136 constant, even for strong electrolytes at low concentrations which can be considered fully ionised in solution. This non-ideal beha-137 138 viour is explained by the relaxation and the electrophoretic phenomena affecting the size and the shape of the hydrate clusters 139 140 formed around the central ions, as well as by the reciprocal influ-141 ence of ions moving in opposite directions. For these reasons, Λ_m 142 also depends on the frequency of the electrical field (Wright, 143 2007). In general, the ions move slower than theoretically 144 expected, meaning a lower conductivity must be taken into consid-145 eration for an accurate interpretation of data (Czeslik et al., 2009). 146 The molar conductivity Δm and the electrolyte concentration *c* are 147 148 connected via the "Kohlrausch Square Root Law",

$$\Lambda_m = \Lambda_m^0 - (a + b \cdot \Lambda_m^0) \cdot \sqrt{c}.$$
⁽²⁾

151 The constants *a* and *b* account for the electrophoresis effect and 152 the relaxation effects described in the Debye–Hückel–Onsager theory. They depend on the temperature, the dielectric properties 153 and the viscosity of the solvent as well as on the number of electric 154 charges of the ions. Values for Λ_m^0 , *a* and *b* can be found in the lit-155 erature (Czeslik et al., 2009; Wright, 2007). Even Eq. (2) is only an 156 approximation that works well for certain substances in specific 157 concentration ranges, but requires further corrections in case of 158 weak electrolytes and/or materials with higher or asymmetrical 159 charge numbers (Wright, 2007). 160

The conductivity κ is typically measured in a conductivity cell as shown in Fig. 1. An external AC voltage *U* causes an electrical current flow *I* through the solution due to ion oscillation between a pair of immersed electrodes. The electrical resistance *R* of the solution, expressed via Ohm's law, in combination with the electrode distance *l* and the cell cross-sectional area *A* determines κ :

$$\kappa = \frac{l}{U} \cdot \frac{l}{A} \cdot Z_{field} \cdot k(T).$$
(3) 169

Suitable values for the so called cell constant *l*/A depend on the 170 range of conductivities to be measured. For example, low conduc-171 tivities between about 1 and 100 μ S cm⁻¹ require cell constants of 172 0.1–0.01 cm⁻¹ (Heule, 2014). The non-dimensional constant Z_{field} is 173 a correction factor to adjust the geometrical cell cross-section for 174 distortions of the electrical field around its edges where the field 175 lines are not ideally parallel. Z_{field} is specific to a certain cell geom-176 etry and must be obtained from calibration experiments with solu-177 tions of known conductivity (Roloff, 2014). The factor k(T) accounts 178 for the temperature dependent viscosity (and thus ion mobility) of 179 the solution, by converting the electrical resistance measured at an 180 arbitrary temperature T to a reference temperature, typically 20 °C 181 or 25 °C. For NaCl a linear temperature compensation is adequate, 182 with suitable factors found in the literature (Flad, 2013). 183

Finally, the combination of Eqs. (1)-(3) gives an expression in which the electrolyte concentration *c* remains the only unknown quantity:

$$\left[\Lambda_m^0 - (a + b \cdot \Lambda_m^0) \cdot \sqrt{c}\right] \cdot c = \frac{I}{U} \cdot \frac{l}{A} \cdot Z_{field} \cdot k.$$
(4) 189

The exact use of this equation for our purposes will be discussed 190 later on, in connection with the cell calibration and the actual 191 concentration measurements. 192

2.2. Experimental setup

The sodium chloride aerosol was generated by dispersing an 194 aqueous sodium chloride solution $(3.75 \text{ g} \text{ l}^{-1})$ in air using an 195





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