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# Extracting a few functionally reproducible biomarkers to build robust subnetwork-based classifiers for the diagnosis of cancer

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## ABSTRACT

In microarray-based case-control studies of a disease, people often attempt to identify a few diagnostic or prog- 22 nostic markers amongst the most significant differentially expressed (DE) genes. However, the reproducibility 23 of DE genes identified in different studies for a disease is typically very low. To tackle the problem, we could 24 evaluate the reproducibility of DE genes across studies and define robust markers for disease diagnosis using 25 disease-associated protein-protein interaction (PPI) subnetwork. Using datasets for four cancer types, we 26 found that the most significant DE genes in cancer exhibit consistent up- or down-regulation in different 27 datasets. For each cancer type, the 5 (or 10) most significant DE genes separately extracted from different 28 datasets tend to be significantly coexpressed and closely connected in the PPI subnetwork, thereby indicating 29 that they are highly reproducible at the PPI level. Consequently, we were able to build robust subnetwork-30 based classifiers for cancer diagnosis. 31

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#### 37 1. Introduction

Numerous microarray studies have been performed to identify 38 genes that are differentially expressed (DE) between cancer samples 39 and normal controls with the objective of discovering diagnostic or 40 prognostic biomarkers (Berchuck et al., 2009; Finak et al., 2008). 41 However, DE genes extracted from different datasets for a particular 42cancer are often very inconsistent (Ein-Dor et al., 2005) mainly due 43 to insufficient statistical power of detecting DE genes in small 44 datasets (Zhang et al., 2008). It is known that thousands of samples 4546 are required in microarray studies to find reproducible biomarkers for cancer (Ein-Dor et al., 2006). Thus, new approaches to evaluating 47 the reproducibility of biomarkers extracted from high-throughput 48 49 biological data are needed (Qiu et al., 2006; Ransohoff, 2005).

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0378-1119/\$ - see front matter © 2013 Elsevier B.V. All rights reserved. http://dx.doi.org/10.1016/j.gene.2013.05.011 Considering that diverse molecular changes in cancers are functionally 50 correlated (Klebanov et al., 2006; Subramanian et al., 2005), we have 51 proposed the use of functional relationships between disease bio-52 markers for evaluating reproducibility (Gong et al., 2010; Gong et al., 53 2011; Yao et al., 2010; Zhang et al., 2009). For example, non-54 overlapping DE genes identified in different datasets for a specific type 55 of cancer tend to be highly consistent when considering their 56 coexpression relationship (Zhang et al., 2009). 57

Using scores based on certain reasonable biological assumptions 58 (or molecular models), we can specify the reproducibility of DE gene 59 discovery at different functional levels. Importantly, the biological 60 assumption underlying a functional consistency score is statistically 61 testable: if the score is significantly higher than expected by chance, 62 then the assumption can explain a large fraction of diverse disease bio- 63 markers. Based on this general framework, we determined the specific 64 functional relationships between disease biomarkers on the protein- 65 protein interaction (PPI) network level. Yao et al. (Yao et al., 2010) 66 showed that non-overlapping PPI network signatures for breast metas- 67 tasis identified from different studies may actually regulate the same 68 sets of interacting protein neighbours. Moreover, Gong et al. found 69 that cancer genes extracted from different databases tend to share 70 significantly more PPI links (Gong et al., 2010). Given that genes 71 encoding interacting proteins tend to share similar functions (Sharan 72 et al., 2007) and DE genes for a disease are often connected in an active 73 PPI subnetwork in response to a disease condition (Guo et al., 2007; 74

Abbreviations: DE, differentially expressed; PPI, protein–protein interaction; SAM, significance analysis of microarray; FDR, false discovery rate; POD, percentage of overlapping deregulations; PO, percentage of overlap; PON, percentage of overlap in the PPI network; SVM, support vector machine; RFE, recursive feature elimination.

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Ideker et al., 2002), it is reasonable to combine gene expression data 75 76 with PPI data to evaluate the 'activated' functional relevance of DE gene lists extracted from different studies (Dittrich et al., 2008; 77 78 Ulitsky and Shamir, 2009).

It is common practice to select a few of the most significant DE 79 genes from thousands of genes as diagnostic or prognostic markers. 80 However, most markers that are selected from a cohort of samples 81 using this simple method, as well as using other complicated feature 82 83 selection algorithms, often fail to work in other independent studies. In an attempt to address this problem, researchers have proposed 84 building classifiers at a "meta-gene" level (Huang et al., 2003; 85 Tamayo et al., 2007) using module-based (Mi et al., 2010), pathway-86 based (Lee et al., 2008) or PPI subnetwork-based approaches 87 88 (Auffray, 2007; Chuang et al., 2007; Dao et al., 2011) rather than at the level of the individual gene. More specifically, several studies 89 have suggested using a combination of gene expression and PPI data 90 to identify "active PPI subnetworks" as relatively reproducible diagnos-91 92tic or prognostic biomarkers for a disease (Chuang et al., 2007; Dao et al., 2011; Su et al., 2010). However, the performance of a classifier 93 based on "meta-genes", such as subnetworks extracted from a dataset 94 for a particular disease, still tends to decline in other independent 95 datasets for the same disease (Chuang et al., 2007; Su et al., 2010). 96 97 This trend could be because certain subnetworks extracted from one 98 cohort of samples using a heuristic optimisation method may consist of genes with less prominent changes in gene expression in other 99 cancer samples. 100

In this paper, we evaluated the reproducibility of the 5 (or 10) 101 102 most significant DE genes extracted from one study in other independent studies for a particular cancer type. First, for each of four cancer 103 types, we evaluated the consistency of the deregulation directions of 104 DE genes (i.e., up- or down-regulated in cancer samples relative to 105normal controls) extracted from different datasets. Then, we pro-106 posed a scoring system to evaluate the reproducibility of two lists 107of the n most significant DE genes in terms of their significant 108 coexpression and close connection in the human PPI network. Our re-109sults supported the assumption that the n most significant DE genes 110 that are separately identified in different datasets for a particular 111 112 type of cancer tend to be significantly coexpressed and closely connected in an active PPI subnetwork associated with the cancer. Fi-113 nally, for each of the four cancer types, we built a classifier using the 114 active PPI subnetworks based on the n most significant DE genes 115extracted from one dataset and evaluated its robustness in another 116 independent dataset. 117

#### 2. Results 118

#### 119 2.1. Reliability of DE gene detection

First, we evaluated the consistency of the deregulation directions, 120 representing increased or decreased average expressions of cancer 121 samples compared to normal samples, between the DE genes that 122123 were separately identified in two datasets for each cancer type (see 124 Table 1). For each dataset, we selected DE genes using SAM with a 1% FDR level. For colon cancer, 1149 DE genes were found in the 125Colon23 dataset, and 1127 (98%) of these genes were included in the 126set of 5478 DE genes identified in the Colon64 dataset, which were sig-127nificantly more than expected by chance  $(P = 2.46 \times 10^{-12})$ 128hypergeometric test). All of the DE genes shared between these two 129 datasets were found to be deregulated in the same directions in the 130 two datasets, which was unlikely to occur by chance  $(P < 1 \times 10^{-12})$ 131 binomial test). Similarly, for each of the other three cancer types, 132almost all DE genes shared between the two datasets were found to 133 be deregulated in the same directions in the two datasets, as indicated 134 by the  $POD_1$  score shown in Table 2. 135

Many of the genes that were selected as DE genes in a dataset but 136 137 not in another dataset may actually be differentially expressed in the

| Cancer  | Datasets <sup>a</sup> | T <sup>b</sup> | N <sup>c</sup> | GEO ACC No. | Platforms      | t1.3 |
|---------|-----------------------|----------------|----------------|-------------|----------------|------|
| Colon   | Colon23               | 15             | 8              | GSE4183     | HG-U133_Plus_2 | t1.4 |
|         | Colon64               | 32             | 32             | GSE8671     | HG-U133_Plus_2 | t1.5 |
| Gastric | Gastric24             | 12             | 12             | GSE19826    | HG-U133_Plus_2 | t1.6 |
|         | Gastric62             | 31             | 31             | GSE13911    | HG-U133_Plus_2 | t1.7 |
| Breast  | Breast58              | 31             | 27             | GSE10810    | HG-U133_Plus_2 | t1.8 |
|         | Breast185             | 42             | 143            | GSE10780    | HG-U133_Plus_2 | t1.9 |
| Lung    | Lung52                | 26             | 26             | GSE7670     | HG-U133A       | t1.1 |
|         | Lung107               | 58             | 49             | GSE10072    | HG-U133A       | t1.1 |
|         | Lung88 <sup>d</sup>   | 44             | 44             | GSE18842    | HG-U133_Plus_2 | t1.1 |
|         | Lung120 <sup>d</sup>  | 60             | 60             | GSE19804    | HG-U133_Plus_2 | t1.1 |
|         | Lung156 <sup>d</sup>  | 91             | 65             | GSE19188    | HG-U133_Plus_2 | t1.1 |

the total number of samples. T denotes the number of tumour samples

N denotes the number of normal samples.

t1.17 <sup>d</sup> These three datasets were used to further evaluate the stability of the classifiers trained for this cancer. t1.18

latter cases. For example, 97% of the 4351 DE genes that were solely 138 identified in the Colon64 dataset showed consistent deregulation di- 139 rections in the Colon23 dataset, which was unlikely to occur by chance 140  $(P < 1 \times 10^{-12})$ , binomial test), indicating that the differential expres- 141 sion signals of most of these DE genes were actually represented in 142 the Colon23 dataset. Similarly, for each of the other three cancer 143 types, we also observed that nearly 90% of the DE genes solely identi- 144 fied in the dataset with greater statistical power showed the same 145 deregulation directions in the dataset with the smaller power, as indi- 146 cated by the POD<sub>2</sub> score shown in Table 2. 147

Taken together, the above results suggested that effective differen- 148 tial expression signals also widely exist in the smaller dataset for each 149 of these cancer types. The high consistency of deregulation directions 150 between the lists of DE genes determined from independent datasets 151 for a particular cancer also validated the reliability of the majority of 152 the DE genes that were identified in different studies for each type of 153 cancer. 154

2.2. Reproducibility of top-ranked most significant DE genes at the PPI 155 level 156

For each cancer type, most of the top n1 (n1 = 5, 10) DE genes 157 extracted from one study were not among the top n2 (n2 = 5, 10) 158 DE genes extracted from another study, as indicated by the low 159  $PO_{n1-n2}$  scores shown in Table 3. However, all of the top 10 DE 160 genes identified in one dataset showed the same deregulation direc- 161 tions in another dataset, thereby indicating that these most signifi- 162 cant DE genes are likely to show differential expressions in other 163 independent cohorts of samples for the same cancer type. 164

We assumed that two DE genes were functionally related if they 165 were significantly coexpressed and connected within two steps of 166 PPI links in the PPI network (see details in Materials and methods, 167

| Table 2POD scores for two lists of DE genes for each cancer type. |                  |                               |           |           |      |  |  |  |
|---|------------------|-------------------------------|-----------|-----------|------|--|--|--|
| Dataset   | DEG <sub>1</sub> | DEG <sub>2</sub> <sup>b</sup> | $POD_1^c$ | $POD_2^d$ | t2.3 |  |  |  |
| Colon23 vs. Colon64   | 1149             | 5478                          | 100%      | 97%       | t2.4 |  |  |  |
| Gastric24 vs. Gastric62   | 100              | 5335                          | 100%      | 95%       | t2.5 |  |  |  |
| Breast58 vs. Breast185  | 4919             | 6742                          | 99%       | 89%       | t2.6 |  |  |  |
| Lung52 vs. Lung107  | 2967             | 4928                          | 99%       | 94%       | t2.7 |  |  |  |

<sup>a</sup>DEG<sub>1</sub> (or <sup>b</sup>DEG<sub>2</sub>) denotes DE genes selected from the former (or the latter) dataset.  $t_{2.8}$ <sup>c</sup>POD1 (or <sup>d</sup>POD2) denotes the proportions of genes that showed the same deregulation t2.9 directions in both datasets among the DE genes shared between the two lists (or among t2.10 the DE genes that solely appeared in DEG<sub>2</sub>). All P-values for the POD<sub>1</sub> and POD<sub>2</sub> scores in t2.11 Table 2 are less than  $1 \times 10^{-12}$ . t2.12

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