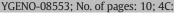
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1 Methods

Community-Analyzer: A platform for visualizing and comparing microbial community structure across microbiomes

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ABSTRACT

A key goal in comparative metagenomics is to identify microbial group(s) which are responsible for conferring 20 specific characteristics to a given environment. These characteristics are the result of the inter-microbial interac- 21 tions between the resident microbial groups. We present a new GUI-based comparative metagenomic analysis 22 application called Community-Analyzer which implements a correlation-based graph layout algorithm that 23 not only facilitates a quick visualization of the differences in the analyzed microbial interactions occurring 24 their taxonomic composition), but also provides insights into the inherent inter-microbial interactions occurring 25 therein. Notably, this layout algorithm also enables grouping of the metagenomes based on the probable inter- 26 microbial interaction patterns rather than simply comparing abundance values of various taxonomic groups. In 27 addition, the tool implements several interactive GUI-based functionalities that enable users to perform standard 28 comparative analyses across microbiomes. For academic and non-profit users, the Community-Analyzer is cur- 29 rently available for download from: http://metagenomics.atc.tcs.com/Community_Analyzer/. 30

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36 1. Introduction

The advent of high throughput sequencing technologies and the 37 emerging field of metagenomics have facilitated rapid extraction and 38 sequencing of the microbial genomic content present in various envi-39 ronments. Several available computational methods/software [1–8] 40 facilitate analysis of the large volume of sequence data obtained from 41 such metagenomic studies. These developments have enabled re-42searchers in profiling the microbial groups they possess in their envi-43 44 ronment(s) of interest. Comparing metagenomic datasets is expected to help in identifying probable key agents responsible for conferring a 45specific phenotypic trait to a given environment. Examples of this in-46 clude traits which are specific to certain disease and physiological disor-4748 ders [9-12].

Currently, a number of methodologies are being used by researchers 49 for performing comparative metagenomic analyses. These include use of 5051Principal Component Analysis (PCA), trees generated using various distance measures and different visualization schemes (force-directed/ 52spring-embedded layout, stacked bar/trend plots, heat maps, etc.). 5354Such analyses have helped researchers in making meaningful inferences. For example, PCA based approach, utilized for comparing 5556gut microbiomes of individuals from diverse nationalities, has helped in identifying three core groups of gut microbiota, referred to as 57

'Enterotypes' [13]. Similarly, use of tree based methods has enabled 58 identification of the variations in microbial communities across various 59 seasons in the Western English Channel [14,15]. 60

A few standalone tools/pipelines are available for comparing micro- 61 bial groups across various environments [3,6,16-20]. In addition, soft- 62 ware package libraries like QIIME have been developed that facilitate 63 comparison of microbial communities using the differential abundance 64 patterns in the SSU-rRNA libraries obtained from the corresponding en- 65 vironments [21]. While the existing tools can distinguish metagenomes 66 based on the composition of the inhabiting organisms in these environ- 67 ments, they do not provide insights into the reason behind the over- or 68 under-abundance of specific groups of organisms. To address this limi- 69 tation, we have developed a standalone user-interactive comparative 70 metagenomics analysis and visualization platform, called 'Community-71 Analyzer'. The platform utilizes a suite of methodologies and graphical 72 layouts in order to facilitate visualization and comparison of microbial 73 community structures within as well as across microbiomes. Most im-74 portantly, in contrast to the above described available analysis servers/75 pipelines, the present tool also facilitates visualization of probable 76 inter-microbial interaction patterns based on the co-occurrence pat-77 terns of microbial groups across environments. 78

Recent studies have indicated that understanding together microbial community structures and the inter-microbial interactions can provide a more comprehensive insight into the subtle differences across 81 microbiomes [22]. The phenotypic trait(s) of any given environment 82 can be associated with the complex inter-microbial interactions within 83 the consortium of microbial groups residing therein. It is therefore 84 important to identify not only the key taxonomic group(s) (specific 85

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group of organisms) that may be responsible for conferring a specific 86 87 trait to a given environment, but also microbial groups that regulate the action of these taxonomic groups. Insights gained from such analy-88 89 ses will be especially useful in metagenomic studies investigating microbial communities associated with diseases and disorders. In such 90 studies, the identified key microbial groups and the organisms regulat-9192 ing them may help in devising appropriate diagnostic as well as thera-93 peutic strategies. In order to get such insights, one needs to obtain 94 answers to the following two questions. What are the inherent similar-95 ities and differences, in terms of microbial composition, amongst the samples under study? Can we infer the probable inter-microbial 96 interaction patterns present in different environments? We address 97 these two key questions by introducing a new graph based layout in 98 Community-Analyzer. The layout has been implemented along with 99 the standard visualization methods like PCA, force-directed/spring-em-100 bedded layouts, heat maps, bar/trend plots and distance-based trees. 101 The implementation of a combination of visualization layout along 102 with several standard features for comparative analyses provides an in-103 novative way of identifying similarities/differences across various mi-104 crobial communities. 105

We exemplify Community-Analyzer's visualization and analysis capabilities using two real metagenomic datasets (as case studies). We also
provide an extensive walk-through of the functionalities of the tool.
End users, without any programming knowledge, can easily perform
analysis using the simple interactive options provided in CommunityAnalyzer.

112 **2. Materials and methods**

Community-Analyzer is available as a standalone tool for Windows and Linux platforms. Fig. 1 summarizes a typical metagenomic analysis work-flow and the application of Community-Analyzer in this workflow. In a typical metagenomic sequencing project, genomic DNA (corresponding to the resident microbial groups) from an environment are extracted. Subsequently either the entire set of genomic fragments 118 (obtained using the shotgun sequencing techniques) or fragments of 119 specific marker genes (e.g., 16S rDNA) are sequenced. The taxonomic affiliations of each of the sequenced fragments (referred to as reads) are 121 then obtained using suitable binning algorithm(s) [1,2,4,7,8,23]. Based 122 on the assignments, these algorithms also provide the abundance profile of the various taxonomic groups in the given environment. These abun-124 dance profiles (referred to as taxonomic classification summaries) from multiple environments can then be provided as inputs to Community-Analyzer for visualization and comparative analysis. 127

2.1. Obtaining 'Community-Analyzer layout' for a set of metagenomic 128 samples 129

A majority of organisms present in any environment are hitherto un- 130 known. These organisms may belong to new species/genus/family/order/ 131 class/phylum. Thus, sequences originating from these organisms are 132 expected to be assigned to taxa at various taxonomic levels (e.g., genus, 133 family, order, class, phylum, etc.). Consequently, most of the binning 134 algorithms generate taxonomic summaries (consisting of abundance 135 profiles of taxonomic groups) at different taxonomic levels. In order to ef- 136 fectively compare two or more metagenomic datasets, it is thus impor- 137 tant to evaluate the differences amongst the metagenomes at various 138 taxonomic levels. Based on the input data (from the taxonomic summary 139 files), Community-Analyzer generates a graphical layout (displaying 140 microbial community structures) at user-specified taxonomic 141 level. The graphical layout is generated to capture and display not 142 only the inter-microbial interaction patterns across the microbial 143 communities under study, but also the similarities/differences 144 across the communities. The generated graphical layout, hereafter 145 referred to as the 'Community-Analyzer layout' (Fig. 2), displays 146 two distinct aspects of the analyzed microbial communities. First, 147 the resident microbial (taxonomic) groups are arranged based on 148 the correlations of the abundance patterns of various taxa across 149

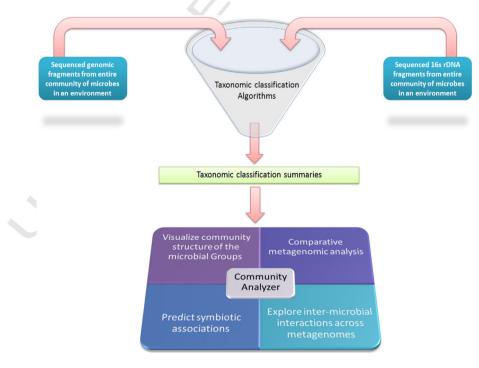


Fig. 1. Taxonomic classification summaries (obtained using representative binning algorithms) are provided as input to Community-Analyzer. In a typical metagenomic sequencing project, genomic DNA (corresponding to resident microbial groups) from an environment are extracted. This is followed by sequencing of either the entire set of genomic fragments (obtained using shotgun sequencing techniques) or the genomic fragments of specific marker genes (e.g. 16S rDNA). The taxonomic affiliations of each of these sequenced genomic fragments are then obtained using any of the available taxonomic classification methods. The obtained taxonomic classification summaries are then provided as input to the Community-Analyzer.

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