



Molecular evolution of $\alpha 4$ integrin binding site to lentiviral envelope proteins in new world primates

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ARTICLE INFO

Article history:

Received 13 March 2012

Received in revised form 2 May 2012

Accepted 25 May 2012

Available online 9 June 2012

Keywords:

Integrins

$\alpha 4\beta 7$

ITGA4

New world primates

Polymorphisms

ABSTRACT

Integrin epitopes encoded by *ITGA4* exons 5 and 6 encompass the $\alpha 4\beta 7$ binding site to natural ligands and HIV-1 gp120. Functional assays of $\alpha 4$ variants of new world primates (NWP) showed reduced binding of several ligands, including the HIV-1 envelope, probably accounting for restriction phenotypes conferring resistance to lentiviral infection (Darc et al., 2011). In this paper, we have analyzed, by cloning and sequencing, the $\alpha 4$ domain polymorphisms present in 10 NWP species and four old world primates (including human). Analyses of differential selection at codon sites and along evolutionary lineages were carried out. We identified codons under positive selection, including polymorphic variations at codon 201, presumably convergent during NWP radiation and significant positive selection leading to a single allele (SagVar2).

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1. Introduction

Early stages of infection by primate lentiviruses are characterized by substantial depletion of activated CD4⁺ T cells in hosts that succumb to immunodeficiency, as is the case of rhesus monkeys, although the mechanisms underlying CD4⁺ T cell depletion in the gut of these primates remain controversial (Ho et al., 1995; Mehanderu et al., 2004; Veazey et al., 2000, 2001). Conversely, natural reservoirs of lentiviruses, like sooty mangabeys and African green monkeys, exhibit limited disease and pathogenesis upon infection (Hazenberget al., 2003; Hirsch et al., 1995; LaBonte et al., 2002; Lederer et al., 2009; Silvestri, 2009), while New World primates (NWP) are not apparently infected *in natura* or in captivity by lentiviruses (McClure et al., 1989; Spertzel, 1989), probably protected by factors that prevent infection (LaBonte et al., 2002; Ribeiro et al., 2005; Sawyer et al., 2004; Sayah et al., 2004; Sebastian and Luban, 2005; Wong et al., 2009).

Integrins are a family of heterodimeric glycoproteins composed of α and β subunits playing a relevant role in cell-to-cell and cell-to-extracellular matrix interactions and defining specific patterns of cell migration to different tissues in vertebrates (Hemler,

1990; Hynes, 1992). Integrin $\alpha 4\beta 7$, encoded by *ITGA4* and *ITGB7*, is essential for cellular trafficking to gut-associated lymphoid tissue (GALT), the principal reservoir of CD4⁺ T cells, and a marker of cellular activation (Agace, 2008; Erle et al., 1994; Hamann et al., 1994; Holzmann et al., 1989; Springer, 1994). In 2008, $\alpha 4\beta 7$ was also recognized as an HIV-1 receptor (Arthos et al., 2008).

Recently, we reported that the $\alpha 4\beta 7$ binding site to natural ligands and HIV-1 gp120 protein was located at epitopes encoded by *ITGA4* exons 5 and 6 (Darc et al., 2011). In addition, analyses of functional binding of NWP $\alpha 4$ variants showed that some species-specific amino acid substitutions reduced binding of some ligands, including the HIV-1 envelope. We postulated that some of these $\alpha 4$ variants might have resulted from positive selection at different residues of the binding site and were relevant in determining restriction phenotypes rendering NWP resistant to lentiviral infection. Most of these changes were lineage-specific, characteristic of different species, genera or families, apparently following the evolutionary NWP radiation, and their emergence could be traced in the phylogeny of this group (Darc et al., 2011).

In this study we analyzed *ITGA4* exons 5 and 6 of 10 NWP species and the presence of polymorphisms with the purpose of identifying sites under positive selection of potential functional significance to lentiviral binding. Positive and negative selection was tested using codon substitution models including data from African and Asian primates. These studies provided evidence of the evolutionary pressures at definite codon sites and branches of the $\alpha 4\beta 7$ lineage in new world primates.

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2. Materials and methods

2.1. Samples and genomic DNA

Genomic DNA of 151 different NWP primate specimens, belonging to all three families and 51 different species, has been isolated and analyzed in a previous report (Darc et al., 2011) (Table 1). We

selected 10 species (*Aotus azarae*, *Callithrix geoffroyi*, *Callithrix jacchus*, *Callithrix kuhlii*, *Callithrix penicillata*, *Callithrix [Cebuella] pygmaea*, *Mico [Callithrix] argentata*, *Mico [Callithrix] emiliae*, *Mico [Callithrix] humeralifer*, *Mico [Callithrix] melanura*) whose $\alpha 4$ domain electropherograms indicated heterozygosity for cloning and sequencing.

Table 1
Specimens analyzed in this study.

Species name	Common name ^a	No. of individuals and cloned samples	Amino acid sequence (AAS)	Nucleotide sequence (NS)
<i>Homo sapiens</i>	Human	1	AAS1	NS1
<i>Pan troglodytes</i>	Common chimpanzee	1	AAS1	NS1
<i>Pongo pygmaeus</i>	Orangutan	1	AAS1	NS1
<i>Macaca mulatta</i>	Rhesus monkey	1	AAS2	NS2
<i>Alouatta belzebul</i>	Red-handed howler	6	AAS3	NS3
<i>Alouatta caraya</i>	Black howler	4	AAS3	NS3
<i>Alouatta guariba</i>	Brown howler	1	AAS3	NS3
<i>Alouatta seniculus</i>	Venezuelan red howler	1	AAS3	NS3
<i>Ateles paniscus paniscus</i>	Red-faced spider monkey	1	AAS4	NS4
<i>Brachyteles arachnoides</i>	Southern muriqui	3	AAS4	NS4
<i>Aotus sp.</i>	Night monkey	4	AAS5	NS5
<i>Aotus azarae</i> (A1)	Azara's night monkey	18 (13)	AAS5	NS5
<i>Aotus azarae</i> (A2)	Azara's night monkey		AAS5	NS6
<i>Aotus a. boliviensis</i>	Azara's night monkey	3	AAS5	NS5
<i>Aotus a. infulatus</i>	Azara's night monkey	1	AAS5	NS5
<i>Aotus infulatus</i>	Azara's night monkey	1	AAS5	NS5
<i>Callimico sp.</i>	Goeldi's marmoset	1	AAS6	NS7
<i>Callimico goeldii</i>	Goeldi's marmoset	1	AAS6	NS7
<i>Callithrix aurita</i>	Buffy-tufted marmoset	2	AAS7	NS8
<i>Callithrix geoffroyi</i>	White-headed marmoset	3 (1)	AAS8	NS9
<i>Callithrix jacchus</i>	Common marmoset	2 (1)	AAS7	NS10
<i>Callithrix kuhlii</i> (A1)	Wied's marmoset	2 (1)	AAS7	NS10
<i>Callithrix kuhlii</i> (A2)	Wied's marmoset		AAS9	NS11
<i>Callithrix penicillata</i> (A1)	Black-tufted marmoset	3 (1)	AAS7	NS10
<i>Callithrix penicillata</i> (A2)	Black-tufted marmoset		AAS8	NS9
<i>Callithrix [Cebuella] pygmaea</i>	Pygmy marmoset	2 (1)	AAS7	NS12
<i>Cebus sp.</i>	Capuchin	6	AAS10	NS13
<i>Cebus albifrons</i>	White-fronted capuchin	5	AAS10	NS13
<i>Cebus apella</i>	Tufted capuchin	11	AAS10	NS13
<i>Cebus apella paraguayanus</i> (A1)	Tufted capuchin from Paraguay	8	AAS10	NS13
<i>Cebus a. paraguayanus</i> (A2)	Tufted capuchin from Paraguay		AAS10	NS14
<i>Cebus capucinus</i>	White-headed capuchin	1	AAS10	NS13
<i>Cebus olivaceus</i>	Weeper capuchin	1	AAS10	NS13
<i>Cebus o. nigrivittatus</i>	Weeper capuchin	5	AAS10	NS13
<i>Cebus xanthosternus</i>	Golded-bellied capuchin	4	AAS10	NS13
<i>Leontopithecus chrysomelas</i>	Golden-headed lion tamarin	4	AAS6	NS15
<i>Leontopithecus chrysopygus</i>	Black lion tamarin	2	AAS6	NS15
<i>Leontopithecus rosalia</i>	Golden lion tamarin	2	AAS6	NS15
<i>Mico [Callithrix] argentata</i>	Silvery marmoset	5 (1)	AAS7	NS10
<i>Mico [Callithrix] emiliae</i>	Emilia's marmoset	6 (1)	AAS7	NS10
<i>Mico [Callithrix] humeralifer</i>	Santarem marmoset	3 (1)	AAS7	NS10
<i>Mico [Callithrix] melanura</i>	Black-tailed marmoset	2 (1)	AAS7	NS10
<i>Saguinus bicolor bicolor</i>	Pied tamarin	1	AAS11	NS16
<i>Saguinus imperator</i>	Emperor tamarin	3	AAS6	NS17
<i>Saguinus martinsi</i>	Martins' tamarin	1	AAS11	NS16
<i>Saguinus midas midas</i>	Red-handed tamarin	3	AAS11	NS16
<i>Saguinus mystax</i>	Moustached tamarin	2	AAS6	NS17
<i>Saguinus niger</i>	Black tamarin	1	AAS11	NS16
<i>Saimiri sp.</i>	Squirrel monkey	4	AAS12	NS18
<i>Cacajao melanocephalus</i>	Black-handed uacari	3	AAS13	NS19
<i>Callicebus sp.</i> (A1)	Titi	2	AAS14	NS20
<i>Callicebus sp.</i> (A2)	Titi		AAS14	NS21
<i>Callicebus coimbrae</i>	Coimbra Filho's titi	1	AAS14	NS20
<i>Callicebus moloch</i>	Red-bellied titi	1	AAS14	NS21
<i>Callicebus nigrifrons</i>	Black-fronted titi	3	AAS14	NS20
<i>Callicebus personatus</i>	Atlantic titi	1	AAS14	NS21
<i>Callicebus p. personatus</i>	Atlantic titi	2	AAS14	NS20
<i>Chiropotes sp.</i>	Saki	3	AAS13	NS19
<i>Chiropotes albinasus</i>	White-nosed saki	1	AAS13	NS19
<i>Chiropotes israelita</i>	Brown-backed bearded saki	1	AAS13	NS19
<i>Pithecia irrorata</i>	Rio Tapajos saki	1	AAS13	NS19

Numbers in parentheses indicate number of cloned samples (one per individual). All other sequences were previously reported by Darc et al. (2011), except for *Homo sapiens* and *Pan troglodytes* which were retrieved from GenBank. A1, A2: alleles.

^a Common names follow Groves (2005).

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