



# The insertion domain 1 of class IIA dimeric glycyl-tRNA synthetase is a rubredoxin-like zinc ribbon

Gurmeet Kaur, Srikrishna Subramanian \*

CSIR-Institute of Microbial Technology, Sector 39A, Chandigarh, India



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## ABSTRACT

The insertion domain 1 (ID1) of class IIA dimeric glycyl-tRNA synthetase ( $\alpha_2$ GRS) is an appended domain in the core catalytic region of the enzyme. ID1 has been shown to play a role in tRNA aminoacylation, mediating interaction with the acceptor arm of tRNA and diadenosine tetraphosphate (Ap4A) synthesis. Mutations in  $\alpha_2$ GRS, including those in the ID1 region, have been implicated in distal hereditary motor neuropathy-V (dHMN-V) and Charcot-Marie-Tooth (CMT) disease. Through sequence and structure based evolutionary analysis, we show that ID1 of  $\alpha_2$ GRS is a rubredoxin-like zinc ribbon domain. The zinc-chelating cysteines of ID1 are well conserved in all archaeal versions of the enzyme and also in several eukaryotes, which most likely have acquired them via horizontal gene transfer from bacteria; but in all other eukaryotes, the zinc-chelating residues are not preserved. ID1 from bacteria display a selective preservation of zinc-binding residues, ranging from complete conservation to complete loss. The ID1 from different organisms harbor variable-sized non-conserved insertions between the two zinc-binding half-sites of the zinc ribbon. Three of the previously identified CMT-associated mutations in  $\alpha_2$ GRS, viz., human D146N, mouse C157R and human S211F, are located in the zinc ribbon region of ID1. Interestingly, human Asp146 which is implicated in the synthesis of Ap4A, a molecule known to act during neuronal transmission, has also been reported to be mutated in dHMN-V, suggesting a possible link between hereditary motor neuropathy and Ap4A synthesis.

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## 1. Introduction

Aminoacyl-tRNA synthetases (aaRSs) are enzymes that help in the translation of genetic information (Söll and Schimmel, 1974). Glycyl-tRNA synthetase (GRS) is an interesting enzyme in that two phylogenetically and structurally different forms are known (O'Donoghue and Luthey-Schulten, 2003; Shiba, 2005; Woese et al., 2000). A tetrameric form ( $(\alpha\beta)_2$ ) of GRS is exclusively found in bacteria and a dimeric ( $\alpha_2$ ) form is present in eukaryotes, archaea and some bacteria (Fig. 1A) (O'Donoghue and Luthey-Schulten, 2003; Woese et al., 2000). The  $(\alpha\beta)_2$ GRS is related to the class IIC aaRSs (Phe and  $(\alpha\beta)_2$ GRS) and the  $\alpha_2$ GRS belongs to class IIA (Ser, Pro, Thr, His and  $\alpha_2$ GRS) (O'Donoghue and Luthey-Schulten, 2003). In the  $(\alpha\beta)_2$ GRS, the  $\alpha$ -subunit harbors the catalytic domain and the  $\beta$ -subunit is the anticodon-binding domain, whereas  $\alpha_2$ GRS displays a modular arrangement with both the catalytic and anticodon-binding domains present in a single polypeptide chain (Cader et al., 2007; Logan et al., 1995;

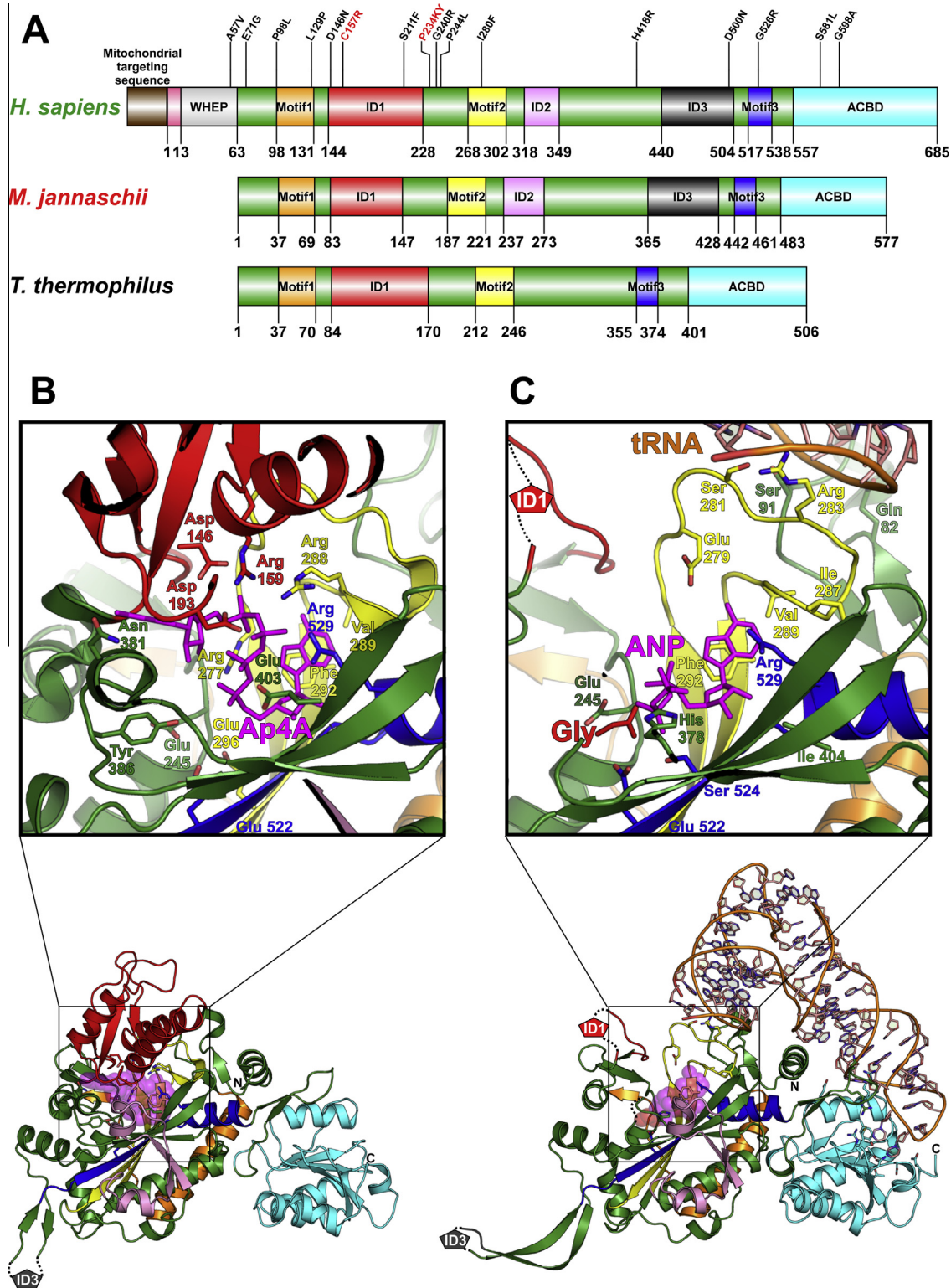
Mazauric et al., 1998; O'Donoghue and Luthey-Schulten, 2003) (Fig. 1A).

The catalytic domain of  $\alpha_2$ GRS has a typical class II aaRS fold (SCOP identifier 55680) made up of a seven-stranded antiparallel  $\beta$ -sheet (Logan et al., 1995) (Fig. 1B). Three consensus sequence motifs define class II aaRSs; wherein motif 1 (G $\phi$ xx $\phi$ xxP $\phi$ ) mediates dimerization while motif 2 (FRxE-loop-(H/R)xxxFxxx(D/E)) and motif 3 (G $\phi$ G $\phi$ G $\phi$ (D/E) $\phi$  $\phi$  $\phi$  $\phi$ ) (where x is any aminoacid,  $\phi$  is a hydrophobic aminoacid, underlined residues are absolutely conserved) contribute crucial active site residues (Cusack, 1995; Eriani et al., 1990) (Fig. 1A). Motif 2 and motif 3 are preserved in  $\alpha_2$ GRS, but motif 1 is atypical, in that the invariant proline residue can be substituted by other aminoacids (Logan et al., 1995; Mazauric et al., 1998). The structure of the anticodon binding domain consists of a mixed  $\beta$ -sheet flanked by  $\alpha$ -helices (SCOP fold identifier 52953) (Cader et al., 2007; Logan et al., 1995). In some  $\alpha_2$ GRS, N- and C-terminal extensions and additional appended domains are also observed (Logan et al., 1995; Mazauric et al., 1998; Shiba, 2005).

Insertion domain 1 (ID1) is an ~85 residue domain in the structurally characterized  $\alpha_2$ GRS (residues 144–228 in human, PDB

\* Corresponding author at: CSIR-Institute of Microbial Technology (IMTECH), Sector 39-A, Chandigarh 160036, India. Fax: +91 1722695215.

E-mail address: [krishna@imtech.res.in](mailto:krishna@imtech.res.in) (S. Subramanian).



**Fig. 1.** Domain organization of  $\alpha_2$ GRS. (A) The domain architectures of representative members of the class IIA GRS are shown. Catalytic domain is colored green and various inserted domains and motifs are indicated. Sixteen known mutations in GRS which lead to CMT and related neuropathies are marked on the human  $\alpha_2$ GRS. The mutations that have been identified in humans are labeled black and those identified in mouse are labeled red. (B) Ribbon diagram of human  $\alpha_2$ GRS with Ap4A bound at the active site (PDB identifier 2ZT5). (C) Ribbon diagram of human  $\alpha_2$ GRS with tRNA, ANP and glycine bound at the active site (PDB identifier 4KR3). Insets reveal a detailed view of the active site pockets and interactions of various residues with the bound moieties. Ap4A and ANP are colored in magenta and glycine at the active site is colored red and shown as spheres and sticks in the full structure and in the inset, respectively. The side chains of various interacting residues are shown as sticks. ANP, Ap4A, tRNA and Gly at the active site and various key residues are labeled. The coloring scheme of the various regions of GRS structures in (B, C) follows that used in (A). Abbreviations: ID (insertion domain), ACBD (anticodon binding domain), ANP (phosphoaminophosphonic acid-adenylate ester), *H. sapiens* (*Homo sapiens*), *M. jannaschii* (*Methanococcus jannaschii*), *T. thermophilus* (*Thermus thermophilus*).

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