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## Review

## Pathogenicity and treatment of Bartonella infections

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### ABSTRACT

*Bartonella* spp. are responsible for emerging and re-emerging diseases around the world. The majority of human infections are caused by *Bartonella henselae*, *Bartonella quintana* and *Bartonella bacilliformis*, although other *Bartonella* spp. have also been associated with clinical manifestations in humans. The severity of *Bartonella* infection correlates with the patient's immune status. Clinical manifestations can range from benign and self-limited to severe and life-threatening disease. Clinical conditions associated with *Bartonella* spp. include local lymphadenopathy, bacteraemia, endocarditis, and tissue colonisation resulting in bacillary angiomatosis and peliosis hepatis. Without treatment, *Bartonella* infection can cause high mortality. To date, no single treatment is effective for all *Bartonella*-associated diseases. In the absence of systematic reviews, treatment decisions for *Bartonella* infections are based on case reports that test a limited number of patients. Antibiotics do not significantly affect the cure rate in patients with *Bartonella* lymphadenopathy. Patients with *Bartonella* spp. bacteraemia should be treated with gentamicin and doxycycline, but chloramphenicol has been proposed for the treatment of *B. bacilliformis* bacteraemia. Gentamicin in combination with doxycycline is considered the best treatment regimen for endocarditis, and erythromycin is the first-line antibiotic therapy for the treatment of angioproliferative lesions. Rifampicin or streptomycin can be used to treat verruga peruana. In this review, we present recent data and recommendations related to the treatment of *Bartonella* infections based on the pathogenicity of *Bartonella* spp.

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## Introduction

*Bartonella* spp. are intracellular bacteria that cause prolonged intraerythrocytic bacteraemia in their hosts and are typically transmitted by haematophagous insects such as phlebotomine sandflies, human body lice and cat fleas, or via animal scratches and bites [1]. To date, more than 30 *Bartonella* spp. and several *Candidatus* spp. have been isolated from humans as well as from wild and domestic animals around the world (Table 1) [2,3]. The suspected role of ticks in the transmission of *Bartonella* spp. is supported by direct and indirect evidence [4–6]. A wide range of mammals serve as reservoirs for *Bartonella* spp., but humans are the only known reservoir for *Bartonella bacilliformis* and *Bartonella quintana* [1].

*Bartonella henselae*, *B. quintana* and *B. bacilliformis* are responsible for the majority of infections in humans [1,7]. The ability to cause acute or chronic infections and vascular proliferative or suppurative manifestations is a remarkable feature of *Bartonella* spp. The severity of clinical manifestations correlates with the patient's

immune status. As a result, *Bartonella* spp. can persist in the blood of their hosts resulting in intraerythrocytic parasitism [1], and they are responsible for a number of diseases including Carrion's disease, cat-scratch disease (CSD), chronic lymphadenopathy, trench fever, chronic bacteraemia, endocarditis, bacillary angiomatosis, peliosis hepatis and neurological disorders [1]. Without treatment, *Bartonella* infections are associated with high mortality and the potential for relapse due to the existence of an intraerythrocytic phase that may provide a protective niche for the bacteria [7].

Owing to the variety of known clinical manifestations and localisations of *Bartonella* spp., no single treatment exists for all *Bartonella*-associated diseases. As a result, treatment approaches must be adapted to each species and clinical situation (Fig. 1) [1,7]. Moreover, clinical studies that include a standard case definition, culture confirmation, rigidly defined disease outcomes, and patients with similar host defences are limited. Clinical data related to the treatment of *Bartonella* infections are primarily based on case reports that test a limited number of patients. As a consequence, current recommendations for the treatment of *Bartonella* infections are based primarily on the clinical course and the immunological status of the patient and rely less on the infective species. The objective of this review is to present recent

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**Table 1**  
*Bartonella* spp. reported to date: epidemiological and clinical data.

<i>Bartonella</i> spp.	First cultivation		Year of description	Reservoir host/vector	Human disease(s)
	Mammal	Area, country			
<i>B. alsatica</i>	Wild rabbit ( <i>Oryctolagus cuniculus</i> )	Alsace, France	1999	Rabbit	Endocarditis, lymphadenopathy
<i>B. australis</i>	Gray kangaroo ( <i>Macropus giganteus</i> )	Queensland, Australia	2007		Not described
<i>B. bacilliformis</i>	Human		1909	Human/sandfly	Oroya fever and verruga peruana
<i>B. birtlesii</i>	Mouse ( <i>Apodemus</i> spp.)	Bodensee, Germany	2000	Rat	Not described
<i>B. bovis</i> ( <i>B. weissii</i> )	Cow	Bissy, France	2002	Cow	Not described
<i>B. capreoli</i>	Roe deer ( <i>Capreolus capreolus</i> )	Chizé, France	2002	Ruminant	Not described
<i>B. chomelii</i>	Domestic cattle ( <i>Bos taurus</i> )	Loire-Atlantique and Nord, France	2004		Not described
<i>B. clarridgeiae</i>	Cat		1996	Cat/cat flea	Lymphadenitis
<i>B. coopersplainsensis</i>	Mottle-tailed rat ( <i>Rattus leucopus</i> )	Queensland, Australia	2008		Not described
<i>B. doshiae</i>	Woodland mammal ( <i>Microtus agrestis</i> )	UK	1995	Rat	Not described
<i>B. elizabethae</i>	Endocarditis patient	USA	1993	Rat	Endocarditis, neuroretinitis
<i>B. grahamii</i>	Woodland mammal ( <i>Clethrionomys glareolus</i> )	UK	1995	Rat, insectivore	Neuroretinitis
<i>B. henselae</i>	Cat		1990	Cat/cat flea	Lymphadenitis, endocarditis, bacillary angiomatosis, bacillary peliosis, Parinaud's oculoglandular, neuroretinitis, osteomyelitis, arthropathy, bacteraemia with fever
<i>B. koehlerae</i>	Domestic cat	California, USA	1999	Cat	Endocarditis, lymphadenitis
<i>B. peromysci</i>	Mouse ( <i>Peromyscus</i> spp.)		1995	Mice	Not described
<i>B. phoceensis</i>	Wild rat ( <i>Rattus norvegicus</i> )	Marseille, France	2004		Not described
<i>B. queenslandensis</i>	Grassland melomys ( <i>Melomys</i> spp.)	Queensland, Australia	2008		Not described
<i>B. quintana</i>	Human		1920	Human/body louse	Trench fever, endocarditis, bacillary angiomatosis, lymphadenitis
<i>B. rattimassiliensis</i>	Rat ( <i>R. norvegicus</i> )	Marseille, France	2004		Not described
<i>B. rattiaustraliensis</i>	Tunney's rat ( <i>Rattus tunneyi</i> )	Queensland, Australia	2008		Not described
<i>B. rochalimae</i>	Human	USA	2007		Bacteraemia, fever, splenomegaly
<i>B. schoenbuchensis</i>	Wild roe deer ( <i>C. capreolus</i> )	Germany	2001	Ruminant/deer ked	Not described
<i>B. talpae</i>	Mole		1995	Mole	Not described
<i>B. tamiae</i>	Human	Khon Kaen, Thailand	2008		Febrile illness
<i>B. taylorii</i>	Woodland mouse ( <i>Apodemus</i> spp.)	UK	1995	Rat	Not described
<i>B. tribocorum</i>	Wild rat ( <i>R. norvegicus</i> )	France	1998		Not described
<i>B. vinsonii</i> subsp. <i>arupensis</i>	Cattle rancher	USA	1999	Dog, rodent/ticks	Bacteraemia, fever, endocarditis
<i>B. vinsonii</i> subsp. <i>berkhoffii</i>	Dog	USA	1995	Dog/ticks	Endocarditis
<i>B. vinsonii</i> subsp. <i>vinsonii</i>	Vole ( <i>Microtus pennsylvanicus</i> )		1946	Vole/vole ear mite	Endocarditis, bacteraemia
<i>B. washoensis</i>	Human	USA	2000	Fleas	Myocarditis, meningitis
<i>B. weissii</i>	Domestic cat	USA	2000	Deer, elk, beef, cattle	Not described
<i>Candidatus</i> spp.					
<i>B. mayotimonensis</i>	Human	France	2010		Endocarditis
<i>B. melophagi</i>	Sheep	USA	2004	Sheep ked ( <i>Melophagus ovinus</i> )	Bacteraemia
<i>B. ancashi</i>	Human	Peru	2013		Verruga peruana
<i>B. merieuxii</i>	Canids	Iraq	2012		Not described
<i>B. antechini</i>	Fleas and ticks	Australia	2011		Not described
<i>B. thailandensis</i>	Rodents	Thailand	2009		Not described

data and recommendations related to the treatment of *Bartonella* infections based on the pathogenicity of *Bartonella* spp.

### Pathogenicity of *Bartonella* spp.

In humans, the infection cycle of *Bartonella* spp. is initiated by colonisation of the primary niche [8,9]. In this stage, the infection is

usually controlled by the immune system and the clinical manifestations are characterised by local lymphadenopathy (i.e. associated with *B. henselae*, *B. quintana* and *Bartonella alsatica*) [1,10,11]. However, under certain poorly defined circumstances, the commensal relationship between reservoir-adapted *Bartonella* spp. and the host is imperfect, resulting in a stealth pathogen strategy [12]. As a result, *Bartonella* spp. are rapidly cleared from the blood after

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