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# *Blastocystis* and urticaria: Examination of subtypes and morphotypes in an unusual clinical manifestation

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#### ABSTRACT

Blastocystis is a human common enteric protist that may colonize a large variety of non-human hosts linked to symptoms and diseases such as abdominal pain, constipation, diarrhea, urticaria, flatulence and irritable bowel syndrome (IBS). Blastocystis exhibits remarkable genetic diversity and multiple subtypes (STs) within the genus with no absolute associations with clinical symptomatology. Here we analyzed fecal samples from Argentinean patients (n = 270) belonging to symptomatic (urticaria and non-specific gastrointestinal symptoms, n = 39) and asymptomatic control (n = 28). Those patients infected with Blasto cystis (n = 67) were submitted for morphological analysis, DNA extraction, 18S PCR, sequencing and STs identification according to DNA barcoding. Blastocystis vacuolar forms were the predominant morphotype (75%), ameboid-like forms were evidenced in 1.5% of samples. Blastocystis ST3 was detected in 71.6% (n = 48), of which 71.4%, (n = 35) and 28.6% (n = 14) belonged to symptomatic and asymptomatic respectively. Other subtypes identified were ST1 (14.9%), ST6 (7.5%) and ST2 (5.9%). Blastocystis 18S barcoding evidenced in non-urticaria symptomatic patients and asymptomatic control group the presence of allele 134 (ST3) (p < 0.0001), while allele 34 (ST3) was detected in 85.7% (18/21) of symptomatic uricaria as compared with control group (1/21) (p < 0.0001). The presence of a particular allele (a34) significantly associated with urticaria patients was detected and the clinical implications of these findings are herein discussed.

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#### 21 1. Introduction

22**03** For a long time, the taxonomy of *Blastocystis* was controversial. Despite the application of molecular phylogenetic approaches, it 23 was only recently that *Blastocystis* sp. was unambiguously classi-24 fied within the Stramenopiles. This eukaryotic major lineage, also 25 called Heterokonta, encompasses very diverse organisms (unicellu-26 27 lar or multicellular, heterotrophic or photosynthetic) such as slime nets, diatoms, water molds and brown algae (Silberman et al., 28 1996). With a cosmopolitan distribution, *Blastocystis* is often the 29 30 most common protozoan isolated in parasitological surveys and presents a different number of species living in the gastrointestinal 31 tract of humans, farm animals, birds, rodents, reptiles and others 32 (Yoshikawa et al., 2004). The transmission is thought to occur via 33 fecal-oral route and the cyst is the only form involved; the extent to 34

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http://dx.doi.org/10.1016/j.actatropica.2015.05.004 0001-706X/© 2015 Published by Elsevier B.V. which human-human, human-animal and animal-human trans-35 mission occur remains under debate (Yoshikawa et al., 2004). 36 Blastocystis displays a relevant genetic diversity where several dis-37 tinct ribosomal lineages or subtypes (STs) have been reported. 38 In 2007, a consensus of *Blastocystis* nomenclature was proposed 39 revealing nine subtypes infecting humans, other mammals and 40 birds, based on the polymorphisms of the small subunit (SSU) rRNA 41 gene. So far, no strict associations between the STs and the hosts 42 have been reported, although moderate host specificity is seen 43 (Stensvold et al., 2009; Victory et al., 2009). Today, at least 17 44 genetically distinct SSU rRNA clusters are known (Alfellani et al., 45 2013 and Graham et al., 2013). In humans, nine subtypes have been Q4 46 reported; the subtypes 1, 2, 3 and 4 are common in Europe; while 47 ST1, ST2 and ST3 seem to present equal prevalences in patients 48 with diarrhea and healthy individuals; ST4 is also linked to diar-49 rhea and/or IBS (Ramírez et al., 2014). In America, the information 50 is scarce: in Colombia, ST1 (34%) and ST2 (23%) and lower propor-51 tions of ST 3 (11.4%), ST 4 (0.8%), ST 6 (19.8%) and ST 8 (10.5%) 52 have been described. Domesticated mammals shared the same 53 STs as those usually seen in humans (ST1, ST2, ST3), while birds 54 and marsupials had STs, which are usually rare in humans (ST6, 55

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ST8) (Ramírez et al., 2014). In Brazil, ST1 (41%), ST2 (32%) and ST3 56 (17%) were identified in an ethnic group (Malheiros et al., 2011) 57 in Mexico a local survey identified 51% of ST1, 23% of ST2, 19% of 58 ST3 and 2% of ST7 (Villalobos et al., 2014). Despite of the initial 59 descriptions of this protozoan in the early 1900s, there is scarce 60 information and misleading assumptions regarding the pathogen-61 esis of this Stramenophile (Tan, 2008). Similarly, until a few years 62 ago Blastocystis was considered as a gut commensal protozoan 63 but later, accumulative epidemiological data suggested not only a 64 close relationship between its presence or changing morphotypes 65 with symptoms but often as a true pathogen (Tan, 2008; Roberts 66 et al., 2013). Thus, the pathogenic role of Blastocystis sp. as the 67 primary cause of enteric symptoms is dubious. Clinical features 68 of illness attributed to this pathogen are non-specific (Tamalee 69 et al., 2014) and include nausea, abdominal pain, flatulence, acute 70 or chronic diarrhea, irritable bowel syndrome (IBS), (Yakoob et al., 71 2010), skin disorders such as allergic cutaneous lesions, urticaria 72 (Ur) and angioedema (Logar et al., 1994; Surangsrirat et al., 2010; 73 Vogelberg et al., 2010; Hameed et al., 2011; Verma and Delfanian, 74 2013). Ur is a relatively common skin disorder that has been 75 attributed to immunological, non-immunological and idiopathic 76 77 causes. Only few case reports have been published about the associ-78 ation of *Blastocystis* infection and Ur but it seems to be an indication that pathogenicity might depend on the subtype (Vogelberg et al., 79 2010; Hameed et al., 2011; Verma and Delfanian, 2013; Katsarou-80 Katsari et al., 2008). Regarding the associations of subtypes with 81 morphotypes and clinical findings, PCR subtyping indicated not 82 only that diverse STs correlates with different morphotypes but 83 also their close relationship with defined symptoms (Dogruman-Al 84 et al., 2008; Jones et al., 2009). Although ST3 is the most common 85 subtype found in the majority of human epidemiological studies, 86 there has been a lower association reported of ST3 with symptoms 87 as compared with ST1 (Ozyurt et al., 2008; Yan et al., 2006). In 88 Argentina, Blastocystis sp. represents one of the most common par-89 asites detected in stools. In a previous report, of 924 asymptomatic 90 and symptomatic patients (diarrhea, abdominal pain, constipa-91 tion, flatulence, bloating, and urticaria), we detected a Blastocystis 97 sp. prevalence of 24% (221/924), where urticaria and non-specific 93 gastrointestinal symptoms were significantly associated with this 94 protist as compared with asymptomatic carriers (p < 0.03). Accord-95 ing to our previous results, the aim of this study was to determine the morphotypes and subtypes of *Blastocystis* present in stools from a Cordoba city population in Argentina. A further aim was to determine if non-specific gastrointestinal symptoms (NSGI) or urticaria (Ur), were related with any Blastocystis subtype or 100 morphotype. 101

#### 102 **2. Materials and methods**

#### 103 2.1. Patients

A total of 270 patients (141 female, 129 male, ranging from 104 1 to 57 years old) from the metropolitan area of Cordoba city, 105 in Argentina attended to the University Hospital and the Benito 106 Soria Dispensary during a 6-month period and were subsequently 107 enrolled in the study. After the regular health checks, clinical his-108 tories with data of recent travels, clinical examinations (within 109 the last 6 months) were recorded and patients were submitted 110 to the laboratory for a fecal microscopy examination. Patients 111 were grouped in asymptomatic control group and symptomatic 112 group, where individuals with chronic Ur or NSGI (intestinal 113 symptoms like diarrhea, constipation, abdominal pain and bloat-114 ing) were included. The local study ethics committee approved 115 this study and written informed consent was obtained from all 116 participants. 117

#### 2.2. Stool specimens and microscopy

Fresh and SAF (saline, sodium acetate, formalin) preserved stool specimens (four different specimens per person, along four days collection) were examined immediately by performing formalin-ethyl acetate and Hoffman sedimentation concentrations techniques and iodine wet mounts, Gram-chromotrope, trichromestained smears, modified acid-fast and modified-trichrome stains were performed in parallel (Garcia, 2007). Positive stools for Blastocystis and Blastocystis co-parasitized with other intestinal parasites were selected for culturing. Duplicate fresh samples containing Blastocystis from symptomatic and asymptomatic patients were immediately cultivated in Jones's medium at 37 °C (Suresh and Smith, 2004) and checked by light microscopy to record parasite growth and morphologic features on days 1, 2 and 7 as described previously (Tan, 2008). Blastocystis morphotype prevalence was estimated by counting 100 parasites in 50 fields (light microscopy, magnification  $400 \times$ ). Blastocystis forms were classified as vacuolar (V): round or spherical trophozoites with the presence of an evident central body surrounded by a cytoplasm with peripheral nucleus; granular (Gr): trophozoites with central body and cytoplasm covered with granules; ameboid-like forms (Am): irregular ameboid-shaped trophozoites with absent central body in the cytoplasm but with peripheral nucleus as described before (Tan and Suresh, 2006; Zhang et al., 2012). The percentage and type of parasite (morphotype) in co-parasitized samples in symptomatic and asymptomatic control group was also estimated.

#### 2.3. DNA extraction, PCR and sequence analysis

Fresh portion of stool (500 mg) was stored in 70% ethylic alcohol after collection and frozen at -20 °C for DNA extraction. From each sample, 250 mg were submitted to DNA extraction using the QIAmp DNA Stool Mini Kit (Qiagen, Hilden, Germany) according to manufacturer's instructions. Genomic DNA was preserved until analysis. The samples were verified for the presence of Blastocystis via PCR amplification of Blastocystis-specific SSU rDNA using the primers RD5 (5'-ATC TGG TTG ATC CTG CCAG T-3') and BhRDr (5'-GAG CTT TTT AAC TGC AAC AAC G-3') as recently recommended (Scicluna et al., 2006; Stensvold, 2013). PCR products were purified and sequenced by both strands using de dideoxy-terminal method (Macrogen, Korea). Sequences were edited in MEGA 4.0 (Tamura et al., 2007) and compared with reference sequences representing each ST in GenBank by BLAST gueries. Additionally, sequences were submitted to sequence queries at the Blastocystis 18S database available at http://pubmlst.org/blastocystis/ for Blastocystis 18S allele calling and confirmation of ST.

#### 2.4. Statistical analysis

*Blastocystis* morphotypes and STs in asymptomatic (control group) and symptomatic patients (Ur and NSGI) were analyzed statistically using the chi-square test from the program MedCal Version 10.0.2. p < 0.05 was considered statistically significant.

#### 3. Results

The partial results of this research were presented in the International Conference of Parasitology held in Mexico city in 2014 (Casero DR, Mongi F, Ramírez JD. 2014). *Blastocystis* subtype 3 (Allele 34) is associated with urticaria in Argentinean patients. ICOPA XIII. P#2212. 124 125

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