

Identification of *Leishmania* at the species level with matrix-assisted laser desorption ionization time-of-flight mass spectrometry

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Abstract

Matrix-assisted laser desorption ionization time-of-flight MALDI-TOF mass spectrometry (MS) is now widely recognized as a powerful tool with which to identify bacteria and fungi at the species level, and sometimes in a rapid and accurate manner. We report herein an approach to identify, at the species level, *Leishmania* promastigotes from *in vitro* culture. We first constructed a reference database of spectra including the main *Leishmania* species known to cause human leishmaniasis. Then, the performance of the reference database in identifying *Leishmania* promastigotes was tested on a panel of 69 isolates obtained from patients. Our approach correctly identified 66 of the 69 isolates tested at the species level with log (score) values superior to 2. Two *Leishmania* isolates yielded non-interpretable MALDI-TOF MS patterns, owing to low log (score) values. Only one *Leishmania* isolate of *Leishmania peruviana* was misidentified as the closely related species *Leishmania braziliensis*, with a log (score) of 2.399. MALDI-TOF MS is a promising approach, providing rapid and accurate identification of *Leishmania* from *in vitro* culture at the species level.

Keywords: Identification, *Leishmania*, MALDI-TOF MS, mass spectra, parasite

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Introduction

Human leishmaniasis, caused by 21 species of the obligate intracellular protozoan *Leishmania*, is one of the most neglected tropical diseases according to the WHO, despite the 2 million new cases recorded yearly [1]. This vector-borne disease encompasses several clinical syndromes, ranging from self-healing cutaneous infection to life-threatening visceral infections, partly depending on the *Leishmania* species involved [2]. Knowledge of the species involved is critical for predicting the evolution and prognosis, and for determining the appro-

priate treatment. However, conventional diagnostic techniques, i.e. microscopic examination of Giemsa-stained smears of infected tissues, *in vitro* cultivation of clinical samples on diphasic NNN or axenic media, and *Leishmania* serology assays, do not allow diagnosis at the species level [3]. Identification at the species level requires molecular biology techniques such as PCR or sequencing. These molecular biology techniques remain at an experimental stage, and are confined to research care centres. The reference standard method for identifying *Leishmania* species, as recommended by the WHO, remains isoenzyme analysis, which is labour-intensive, costly, and confined to reference centres [1]. Thus, no rapid and simple method for the species identification of *Leishmania* currently exists.

For approximately 10 years, matrix-assisted laser desorption ionization time-of-flight (MALDI-TOF) mass spectrometry (MS) has been proposed as an alternative to conventional and routine identification methods in clinical microbiological

laboratories. This identification technique was originally adapted for the identification of prokaryotic organisms [4]. New developments have extended its use to a few eukaryotic organisms, such as yeasts and moulds, making this technique a straightforward, fast and reliable identification method for bacteria, yeasts and moulds in a cost-effective way [5–7]. However, MALDI-TOF MS has not yet been adapted for the identification of *Leishmania* or any other protozoan parasites. In this article, we show how we first constructed a reference mass spectra library (MSL) by using cultures of 56 well-characterized *Leishmania* isolates, and then tested it by identifying a panel of 69 isolates with a MALDI-TOF MS-based approach.

TABLE I. Main characteristics of the *Leishmania* isolates furnished by the Centre National de référence des Leishmanioses included in the reference database

Taxon	Isolate	Provenance	Host	CF	Zymodeme
<i>Leishmania infantum</i>	MHOM/EG/87/RTC2	Egypt	<i>Homo sapiens</i>	VL	MON-98
<i>L. infantum</i>	MHOM/FR/87/RMI	France	<i>Canis familiaris</i>	VL	MON-108
<i>L. infantum</i>	MHOM/TN/80/IPT1	Tunisia	<i>H. sapiens</i>	VL	MON-I
<i>L. infantum</i>	MHOM/DZ/82/LIPA59	Algeria	<i>H. sapiens</i>	CL	MON-24
<i>L. infantum</i>	MHOM/FR/78/LEM75	France	<i>H. sapiens</i>	VL	MON-I
<i>Leishmania donovani</i>	MHOM/IN/00/DEVI	India	<i>H. sapiens</i>	VL	MON-2
<i>L. donovani</i>	MHOM/IN/96/THAK57	India	<i>H. sapiens</i>	VL	MON-2
<i>L. donovani</i>	MHOM/SD/98/LEM3580	Sudan	<i>H. sapiens</i>	VL	MON-18
<i>L. donovani</i>	MHOM/ET/67/HU3	Ethiopia	<i>H. sapiens</i>	VL	MON-18
<i>L. donovani</i>	MHOM/IN/80/DD8	India	<i>H. sapiens</i>	VL	MON-2
<i>Leishmania archibaldi</i>	MHOM/ET/72/GEBREI	Ethiopia	<i>H. sapiens</i>	VL	MON-82
<i>L. archibaldi</i>	MCAN/SD/98/LEM3556	Sudan	<i>C. familiaris</i>	VL	MON-82
<i>Leishmania major</i>	MHOM/SU/73/5-ASKH	Turkmenistan	<i>H. sapiens</i>	CL	MON-4
<i>L. major</i>	MHOM/IL/81/FRIEDLINa	Israel	<i>H. sapiens</i>	CL	MON-103
<i>L. major</i>	MHOM/TN/2001/LEM4286	Tunisia	<i>H. sapiens</i>	CL	MON-25
<i>L. major</i>	MHOM/MA/2003/LEM4685	Morocco	<i>H. sapiens</i>	CL	MON-25
<i>L. major</i>	MHOM/YE/76/LEM62	Yemen	<i>H. sapiens</i>	CL	MON-26
<i>Leishmania aethiopica</i>	MHOM/ET/72/L100	Ethiopia	<i>H. sapiens</i>	CL	MON-14
<i>L. aethiopica</i>	MHOM/ET/91/KASSAYE	Ethiopia	<i>H. sapiens</i>	DCL	MON-248
<i>Leishmania killicki</i>	MHOM/TN/80/LEM163	Tunisia	<i>H. sapiens</i>	CL	MON-8
<i>L. killicki</i>	MHOM/TN/80/LEM180	Tunisia	<i>H. sapiens</i>	CL	MON-8
<i>Leishmania tropica</i>	MHOM/MA/95/LEM3015	Morocco	<i>H. sapiens</i>	CL	MON-264
<i>L. tropica</i>	MHOM/SU/74/K27	Azerbaijan	<i>H. sapiens</i>	CL	MON-60
<i>L. tropica</i>	MHOM/GR/80/GR-L35	Greece	<i>H. sapiens</i>	CL	MON-56
<i>L. tropica</i>	0000/00/84/LEM643	NA	NA	VL	MON-104
<i>L. tropica</i>	MRAT/IQ/72/ADHANISI	Iraq	<i>Rattus rattus</i>	VL	MON-5
<i>Leishmania braziliensis</i>	MHOM/BR/75/M2904	Brazil	<i>H. sapiens</i>	CL	MON-165
<i>L. braziliensis</i>	MHOM/BR/84/LTB300	Brazil	<i>H. sapiens</i>	MCL	MON-166
<i>L. braziliensis</i>	MHOM/BR/82/LTB12JULY82	Brazil	<i>H. sapiens</i>	MCL	MON-208
<i>L. braziliensis</i>	MHOM/BR/75/M2903b	Brazil	<i>H. sapiens</i>	CL	MON-43
<i>L. braziliensis</i>	MHOM/GF/2003/LAV008	FG	<i>H. sapiens</i>	CL	MON-43
<i>Leishmania guyanensis</i>	MHOM/GF/98/LEM3657	FG	<i>H. sapiens</i>	CL	MON-45
<i>L. guyanensis</i>	MHOM/GF/2003/LEM4570	FG	<i>H. sapiens</i>	CL	MON-45
<i>L. guyanensis</i>	MHOM/GF/2004/LAV016	FG	<i>H. sapiens</i>	CL	MON-45
<i>L. guyanensis</i>	MHOM/GF/79/LEM85	FG	<i>H. sapiens</i>	CL	MON-45
<i>Leishmania panamensis</i>	MHOM/EC/90/CALDERON	Equador	<i>H. sapiens</i>	CL	MON-161
<i>L. panamensis</i>	MDAS/PA/71/LS94	Panama	<i>H. sapiens</i>	CL	MON-47
<i>Leishmania peruviana</i>	MHOM/PE/84/CE49	Peru	<i>H. sapiens</i>	CL	MON-140
<i>Leishmania lainsoni</i>	IUBI/BR/00/M12025	Brazil	<i>Lutzomyia ubiquitalis</i>	None	MON-150
<i>L. lainsoni</i>	MCUN/BR/85/M19342	Brazil	<i>Cuniculus paca</i>	None	MON-151
<i>Leishmania naiffi</i>	MDAS/BR/79/M5533	Brazil	<i>Dasyurus novemcinctus</i>	None	MON-148
<i>Leishmania amazonensis</i>	MHOM/BR/72/M1845	Brazil	<i>Proechimys sp.</i>	None	MON-41
<i>L. amazonensis</i>	MHOM/BR/73/M2269	Brazil	<i>H. sapiens</i>	CL	MON-132
<i>L. amazonensis garnhami</i>	MHOM/VE/76/JAP78	Venezuela	<i>H. sapiens</i>	CL	MON-41
<i>Leishmania mexicana</i>	MNYC/BZ/62/M379	Belize	<i>Nyctomyia sumichrasti</i>	CL	MON-40
<i>L. mexicana</i>	MHOM/BZ/82/BEL21	Belize	<i>H. sapiens</i>	CL	MON-156
<i>L. mexicana pifanoi</i>	MHOM/VE/57/LL1	Venezuela	<i>H. sapiens</i>	DCL	MON-40
<i>Leishmania pifanoi</i>	MHOM/VE/57/LL1a	Venezuela	<i>H. sapiens</i>	DCL	MON-40
<i>Leishmania deanei</i>	MCOE/BR/78/M5088	Brazil	<i>Coendou sp.</i>	None	MON-52
<i>L. deanei</i>	MCOE/BR/74/M2674	Brazil	<i>Coendou prehensilis</i>	CL	MON-134
<i>Leishmania arabica</i>	MPSA/SA/83/JISH220	Saudi Arabia	<i>Psammomys obesus</i>	VL	MON-99
<i>Leishmania enrietti</i>	MCAV/BR/45/L88	Brazil	<i>Cavia porcellus</i>	CL	MON-97
<i>L. enrietti</i>	MCAV/BR/95/CUR2	Brazil	<i>C. porcellus</i>	CL	MON-97
<i>Leishmania gerbilli</i>	MRHO/CN/60/GERBILLI	China	<i>Rhomboomys opimus</i>	CL	MON-22
<i>Leishmania turanica</i>	MRHO/SU/65/VL	Turkmenistan	<i>R. opimus</i>	VL	MON-21
<i>L. turanica</i>	MMEU/SU/79/MEL	Georgia	<i>Meles meles</i>	None	MON-65

CL, cutaneous leishmaniasis; DCL, diffuse cutaneous leishmaniasis; FG, French Guyana; MCL, mucocutaneous leishmaniasis; NA, not available; VL, visceral leishmaniasis.

Materials and Methods

Leishmania isolates and culture

A panel (panel 1) of 56 isolates was used to construct a mass spectra library, and is summarized in Table I. The isolates were provided and accurately identified with the isoenzyme method by the Centre National de référence des Leishmanioses (CNR-L) (French National Reference Centre for Leishmaniasis) in Montpellier, France. A different panel (panel 2) of 69 isolates was used to assess identification performance (Table 2). These isolates were provided by the CNR-L, the Institut Pasteur of Alger, Algeria, and the Institut

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