## Ecological replacement of *Enterococcus* faecalis by multiresistant clonal complex 17 *Enterococcus faecium*

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## **ABSTRACT**

The proportion of enterococcal infections caused by ampicillin-resistant Enterococcus (AREfm) in a European hospital increased from 2% in 1994 to 32% in 2005, with prevalence rates of AREfm endemicity of up to 35% in at least six hospital wards. Diabetes mellitus, three or more admissions in the preceding year, and use of β-lactams and fluoroquinolones, were all associated with AREfm colonisation. Of 217 AREfm isolates that were genotyped, 97% belonged to clonal complex 17 (CC17). This ecological change mimics events preceding the emergence of vancomycin-resistant E. faecium (VREF) in the USA and may presage the emergence of CC17 VREF in European hospitals.

**Keywords** Ampicillin resistance, colonisation, ecological replacement, *Enterococcus faecium*, risk-factors

Original Submission: 28 June 2006; Revised Submission: 15 August 2006; Accepted: 21 August 2006

*Clin Microbiol Infect* 2007; **13**: 316–319 10.1111/j.1469-0691.2006.01631.x

Molecular epidemiological studies of *Enterococcus* faecium have revealed the existence of host-specific genogroups, including an ampicillin-resistant genetic lineage, labelled clonal complex 17 (CC17), which are associated with nosocomial

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outbreaks and infections in five continents [1]. In European hospitals, rates of infection with vancomycin-resistant *E. faecium* have been rising since the year 2000 (EARSS Annual Report 2004; http://www.rivm.nl/earss), suggesting that the emergence of vancomycin-resistant *E. faecium* in Europe may be following the pattern observed in the USA, but with a 10-year delay. The emergence of vancomycin-resistant *E. faecium* in the USA was preceded by the emergence of ampicillin resistance in *E. faecium* [2,3].

Stimulated by an increase in ampicillin-resistant E. faecium (AREfm) bloodstream infections during 2003, the present study analysed trends in enterococcal infection and colonisation at the University Medical Centre, Utrecht, The Netherlands (1042 beds). The prevalence of invasive enterococcal infections was assessed retrospectively using microbiological data for 1994–2005. Invasive infections were defined as infectious episodes with enterococci isolated from normally sterile specimens, e.g., blood, abdominal fluid, intravascular catheter tips, cerebrospinal fluid, pus and wound specimens. Enterococci isolated from urine were not considered to represent invasive infections. Yearly proportions of Enterococcus faecalis and E. faecium among enterococcal bloodstream infections were determined on the basis of the first 20 enterococcal blood culture isolates (one per patient) per year. The intestinal AREfm reservoir was measured by monthly point-prevalence studies between August 2005 and January 2006 in seven hospital wards (haematology, 21 beds; gastroenterology/nephrology, 25 beds; adult intensive care unit (ICU), 24 beds; paediatric ICU, 47 beds; geriatrics, 15 beds; general surgery, 30 beds; and dermatology, 12 beds). The AREfm reservoir in the community was investigated using faecal samples from 650 outpatients with abdominal discomfort who visited general practitioners in the Utrecht region during 2004.

Risk-factors for colonisation with AREfm were determined using clinical and demographical data for patients in the mixed gastroenterology/nephrology ward. Statistical analysis was performed with SPSS v.12.0.1 (SPSS Inc., Chicago, IL, USA). The clinical impact of AREfm was determined by analysis of clinical, demographical and outcome data for all patients with an invasive AREfm infection between May 2001 and November 2005.

Enterococcosel enrichment broth and agar plates (Becton Dickinson, Cockeysville, MD, USA), supplemented with aztreonam 75 mg/L and amoxycillin 16 mg/L, were used to obtain isolates of AREfm. Resistance was confirmed by amoxycillin Etests (AB Biodisk, Solna, Sweden). A species-specific multiplex PCR, based on the ddl gene of E. faecalis and E. faecium, was used for speciation [4]. Susceptibilities to ampicillin and imipenem were determined by inoculation of Mueller-Hinton agar containing ampicillin 8 mg/mL and imipenem 16 mg/mL, according to CLSI (formerly NCCLS) guidelines. Isolates were genotyped using multiple locus variable number tandem repeat analysis (MLVA), which is based on variations in the number of tandem repeats at six different loci [5]. MLVA profiles were analysed using BioNumerics v.4.00 (Applied Maths, St-Martins-Latem, Belgium).

The overall number of patients with invasive enterococcal infections decreased from 393 in 1994 to 243 in 2005, but proportions of ampicillinresistant enterococcal infections increased from 2% in 1994 to 32% in 2005 (p <0.001) (Fig. 1). E. faecium increased from 3% of enterococcal bloodstream infections in 1994-1996 to 30% in 2003–2005 (p <0.001), and 75% of E. faecium blood culture isolates were resistant to both ampicillin and imipenem, compared with 0% for E. faecalis (p < 0.001).

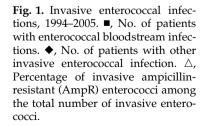
Point-prevalence studies revealed carriage rates ranging from 0% in dermatology to 10.3% in the paediatric ICU, 29% in gastroenterology/nephrology, 29.2% in the adult ICU, 34.6% in haematology and 34.8% in geriatrics. AREfm isolates were obtained from 19 (2.9%) of 650 communityderived faecal samples. No data were available concerning previous hospitalisation or antibiotic use for these patients.

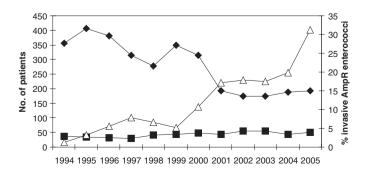
Diabetes mellitus (OR 8.59, 95% CI 2.08–35,44). three or more admissions in the preceding year (OR 14.84, 95% CI 3.44-64.10) and use of β-lactams (OR 2.97, 95% CI 1.09-8.09) or guinolones (OR 5.23, 95% CI 1.22-22.48) were associated AREfm independently with colonisation (Table 1).

Between May 2001 and November 2005, 167 patients had an AREfm infection, with blood (n = 53) and pus (n = 30) as the predominant samples. Overall, 154 (92%) of 167 patients had received antibiotics before developing an infection with AREfm. Nineteen of 20 haematology patients with AREfm infection had bacteraemia. Haematology patients in the studied hospital receive ciprofloxacin prophylaxis during prolonged granulocytopenia, and imipenem is the empirical antibiotic regimen for granulocytopenic fever. During 2002–2005, 7% (n = 12) of all episodes of bacteraemia during granulocytopenia were caused by AREfm. The average period between obtaining blood cultures and commencing appropriate antimicrobial therapy (i.e., vancomycin) was 2 days. Of 167 patients with invasive AREfm infections, 58 (35%) died during hospitalisation, with an average period of 15 days (0-105 days) between identification of the AREfm infection and death.

MLVA typing of 217 AREfm isolates revealed 40 different MLVA types, of which 211 (97%) belonged to CC17 (data not shown). A gradual increase in infection and colonisation episodes with CC17 E. faecium has occurred in this hospital in recent years, with bacteraemia accounting for 32% of all infections, an overall mortality rate of 35%, and CC17 infections being most prevalent among high-risk patients (i.e., transplant or ICU patients).

E. faecium CC17 consists mainly of clinical isolates and isolates associated with hospital





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