



Antigenic requirement for Gag in a vaccine that protects against high-dose mucosal challenge with simian immunodeficiency virus

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ABSTRACT

We reported previously on a vaccine approach that conferred apparent sterilizing immunity to SIVsmE660. The vaccine regimen employed a prime–boost using vectors based on recombinant vesicular stomatitis virus (VSV) and an alphavirus replicon expressing either SIV Gag or SIV Env. In the current study, we tested the ability of vectors expressing only the SIVsmE660 Env protein to protect macaques against the same high-dose mucosal challenge. Animals developed neutralizing antibody levels comparable to or greater than seen in the previous vaccine study. When the vaccinated animals were challenged with the same high-dose of SIVsmE660, all became infected. While average peak viral loads in animals were slightly lower than those of previous controls, the viral set points were not significantly different. These data indicate that Gag, or the combination of Gag and Env are required for the generation of apparent sterilizing immunity to the SIVsmE660 challenge.

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Introduction

Recent reports have demonstrated that “functional” cures for HIV infection might sometimes be possible (Saez-Cirion et al., 2013; Hutter et al., 2009). Cessation of antiretroviral therapy, with or without allogeneic bone marrow transplantation, however, often leads only to a delay in the rebound of viral loads (Henrich et al., 2013; Persaud et al., 2013). Even if some individuals can maintain control of their viral loads throughout their lifetime, the limitations of using these strategies globally are apparent. Development of a prophylactic vaccine is still an essential component of any strategy to eradicate HIV infection worldwide.

Recombinant rhesus cytomegalovirus (RhCMV) vaccine vectors expressing Gag, Pol, Env, and a Rev–Tat–Nef fusion protein have been shown to induce immunity that can control and eventually clear a pathogenic SIVmac239 challenge virus in roughly half of the vaccinated rhesus macaques (Hansen et al., 2011; Hansen et al., 2013). A variety of other vaccine vectors have also achieved varying levels of apparent sterilizing protection in multiple low dose challenges (Barouch et al., 2012; Patel et al., 2013; Flatz et al., 2012; Letvin et al., 2011). Multiple low-dose challenge mo-

dels closely mimic a heterosexual exposure to HIV because of the transmission of a small number of founder viruses from the challenge stock (Keele et al., 2009; Li et al., 2010). In men who have sex with men (MSM), however, multiple founder viruses are often transmitted during the time of initial exposure (Li et al., 2010; Tully et al., 2012). Since a greater number of founder viruses are transmitted during a high-dose challenge, this model may more accurately predict the efficacy of vaccines to prevent the transmission of HIV in MSM.

We previously described a vaccine regimen that resulted in apparent sterilizing immunity to a high-dose mucosal challenge with the pathogenic SIVsmE660 quasiespecies. This vaccine used a heterologous prime–boost vaccine strategy with vectors based on recombinant vesicular stomatitis virus (VSV) and virus-like vesicles (VLVs) derived from a Semliki Forest Virus (SFV) replicon packaged with VSV G (SFV-G). These vectors encoded Gag and Env proteins derived from the SIVsmE660 viral swarm (Schell et al., 2011). Four out of six animals challenged never showed detectable viral loads, and the two that were infected rapidly controlled their viral loads to below the limit of detection. Control of viral loads in the infected animals was mediated by CD8⁺ T cells, as evidenced by the rebound in viral loads when CD8⁺ T cells were depleted from these animals (Schell et al., 2011). The 4 protected animals showed no rebound in viral loads upon CD8⁺ T cell depletion implying that the vaccine generated sterilizing immunity. These animals had high neutralizing antibody (nAb) to E660 Envs prior to challenge, but

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When the Env-only animals were compared to the previous Gag+Env animals, similar neutralization titers against E660.11 were seen over the vaccination course. In Fig. 4D, the average neutralization of E660.11 envelope is shown for the Env-only (green line) and Gag+Env vaccinated animals. The Gag+Env group was divided into protected (orange line) and infected (red line) animals. By the time of the VSV boost (day 119), the Env-only animals had developed significantly higher titers of nAbs against E660.11 envelope than either the protected or infected animals of the Gag+Env group as determined by a two-tailed T test ($p < 0.0006$ and $p < 0.02$, respectively). This significant difference in nAb titers between the two groups was maintained through the day of challenge (day 147). Only those animals that were unprotected (green and red lines) showed an anamnestic nAb response after the challenge.

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