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Platinum Priority – Brief Correspondence Editorial by Adam Pickard and Ian G. Mills on pp. 555–556 of this issue

Telomerase Activity and Telomere Length in Human Benign Prostatic Hyperplasia Stem-like Cells and Their Progeny Implies the Existence of Distinct Basal and Luminal Cell Lineages

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Article info

Article history: Accepted September 22, 2015

Associate Editor: Christian Gratzke

Keywords:

Benign prostatic hyperplasia Telomerase Stem cells

Abstract

Benign prostatic hyperplasia (BPH) treatments have changed little over many years and do not directly address the underlying cause. Because BPH is characterised by uncontrolled cell growth, the chromosomal telomeres should be eroded in the reported absence or low levels of telomerase activity, but this is not observed. We investigated the telomere biology of cell subpopulations from BPH patients undergoing transurethral resection of prostate (TURP). Measurement of TERC, TERT, and telomerase activity revealed that only the epithelial stem-like and progenitor fractions expressed high levels of telomerase activity ($p < 0.01$) and individual enzyme components ($p < 0.01$). Telomerase activity and TERT expression were not detected in stromal cells. Telomere length measurements reflected this activity, although the average telomere length of (telomerase-negative) luminal cells was equivalent to that of telomerase-expressing stem/progenitor cells. Immunohistochemical analysis of patient-derived BPH arrays identified distinct areas of luminal hyperproliferation, basal hyperproliferation, and basal-luminal hyperproliferation, suggesting that basal and luminal cells can proliferate independently of each other. We propose a separate lineage for the luminal and basal cell components in BPH.
Patient summary: We unexpectedly found an enzyme called <i>telomerase</i> in the cells that maintain benign prostatic hyperplasia (BPH), suggesting that telomerase inhibitors could be used to alleviate BPH symptoms.

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Morbidity and loss of quality of life due to benign prostatic hyperplasia (BPH) are relatively high. There are >210million cases of BPH worldwide [1], and the histologic prevalence of BPH is approximately 80% at the age of 80 yr. BPH is characterised by uncontrolled but noninvasive growth of a variety of cell types in the prostate, ranging in frequency from adenomyofibromatous, fibromuscular, fibroadenomatous, stromal, to rare muscular compositions.



Medical treatments for BPH include targeting of the smooth muscle cells by α_{1-} adrenergic receptor inhibitor and use of steroid 5α -reductase inhibitors to target the androgensignalling axis. These treatments take several months to show urinary flow rate benefits but perhaps are ineffective in targeting androgen-independent basal cell proliferation. Surgical intervention by transurethral resection relieves urinary flow obstruction.

In normal somatic cells, repeated cell divisions result in an erosion of the repetitive terminal DNA sequences, the telomeres. The ultimate result of extreme telomere shortening is stress signal-mediated apoptosis or senescence. An understanding of telomerase biology is crucial to comprehend the proliferative dynamics of a tissue. Increased telomerase can be detected in approximately 70-90% of human cancers, including prostate cancer [2]. Although BPH tissue also exhibits a proliferative index two to three times higher than normal prostate [3], telomerase remains undetectable or is only sporadically expressed [2]. The inability to detect telomerase in non-stem-like cells, including the differentiated basal and luminal cells (>95% of the lesion), could have been interpreted as absent or sporadic telomerase expression or activity patterns in previous whole-biopsy analyses [2].

We assessed expression of the RNA (TERC) and protein (TERT) components of telomerase enzyme, telomerase activity, and telomere length in uncultured human BPHderived stem-like cells (SCs: Lin⁻/CD31⁻/EpCAM⁺/CD133⁺/ $CD44^{+}/\alpha_{2}\beta_{1}^{hi}$), transit amplifying cells (TAs: Lin⁻/CD31⁻/ EpCAM⁺/CD133⁻/CD44⁺/ $\alpha_2\beta_1^{hi}$), committed basal cells (CBs: $Lin^{-}/CD31^{-}/EpCAM^{+}/CD133^{-}/CD44^{+}/\alpha_{2}\beta_{1}^{lo}$), luminal cells (LCs: Lin⁻/CD31⁻/EpCAM⁺/CD44⁻/CD24⁺), and stromal cells (Lin⁻/CD31⁻/EpCAM⁻) (Supplementary Fig. 1, Supplement 1) [4,5] freshly purified from BPH tissues (Supplementary Table 1). The expression of NANOG was also higher in SCs (Supplementary Fig. 2). Previously, we reported that TERT expression was undetectable in normal prostate epithelial SCs and TAs [6]. In human BPH, however, the SCs and TAs exhibited significant overexpression of both TERC and TERT compared with the more differentiated CBs and LCs and stromal cells (Fig. 1a and 1b). Telomerase activity was also exclusively restricted to the SC/TA compartment, apart from one CB sample (Fig. 1c). All biopsies were specifically taken from the transitional zone with no detectable prostate cancer lesions on histology from patients with serum prostatespecific antigen levels <4 ng/ml to reduce the possibility of contamination with (telomerase-positive) cancer cells (Supplementary Table 1). TERT expression and telomerase activity were undetectable in stromal cells, indicating that telomerase is not essential for stromal hyperproliferation in BPH—another intriguing disparity in earlier data.

Epithelial cell telomere length measurements revealed that telomerase-expressing SCs and TAs had longer telomeres than those of their immediate differentiated progeny, the telomerase-negative CBs (p < 0.01) (Fig. 1d). The relative telomere length of LCs, however, was almost equivalent to that of SCs and TAs and was always longer than telomere lengths of patient-matched CBs (p < 0.01). Because LCs do not exhibit any detectable telomerase

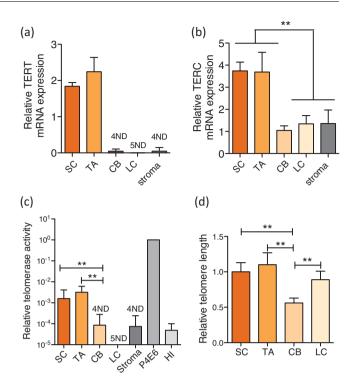


Fig. 1 – Telomerase biology in epithelial subpopulations derived from human benign prostatic hyperplasia. (a) Quantitative reverse transcription polymerase chain reaction (qRT-PCR) analysis for TERC expression (n = 5). Data normalised to RPLP0 expression. (b) qRT-PCR analysis for TERT expression (n = 5). Data normalised to RPLP0 expression. (c) qRT-PCR analysis to measure telomerase activity using TRAPeze RT telomerase detection kit (Millipore, Bedford, MA, USA) (n = 5). Data normalised to telomerase activity in the P4E6 cell line. (d) Relative telomere length determination using modified qRT-PCR method (n = 5). The telomere length for each patient sample was normalised to the individual patients' lymphocyte telomere lengths. Error bars represent mean \pm SD.

^{***} p < 0.001.

BPH = benign prostatic hyperplasia; CB = committed basal cells; HI = heat inactivated control; LC = luminal cells; mRNA = messenger RNA; ND = not detected; SC = stem-like cells; TA = transit-amplifying cells.

activity (Fig. 1c) and are believed to be derived from basal cells, the paradoxical observation of longer LC telomeres compared with CBs suggests that basal and luminal cells may be derived from distinct SC progenitors.

We stained BPH sections to determine basal cell content (using TP63) and luminal cell content (using NKX3.1) (Fig. 2a) and showed that basal and luminal hyperproliferation can occur together and even independently of each other because the basal:luminal ratio was highly variable, ranging from 0.22 to 2.08 (Fig. 2b, Supplementary Fig. 3), providing further evidence that basal and luminal cells may indeed be derived from different progenitors.

In human prostate, we have previously shown that SCs differentiate into CB cells via TA cells [4], unlike mouse cells, in which the situation is less clear [7]. The lineage of LCs, however, has not been determined in humans, although in vitro we can readily derive luminal cells from basal cells under specific culture conditions [5]. We now propose that basal and luminal cell lineages may develop separately in

p < 0.05. p < 0.01.

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