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SEX- AND AFFERENT-SPECIFIC DIFFERENCES IN HISTAMINE RECEPTOR EXPRESSION IN VAGAL AFFERENTS OF RAT: A POTENTIAL MECHANISM FOR SEXUAL DIMORPHISM IN PREVALENCE AND SEVERITY OF ASTHMA

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Abstract—The incidence of asthma is more common in boys than in girls during the puberty, and more common in premenopausal female than age-matched males. Our previous study demonstrated a gender difference in histamine-mediated neuroexcitability in nodose ganglia neurons (NGNs), highlighting a possibility of histamine-mediated gender difference in asthma via visceral afferent function. In the present study, we aimed to explore the gender difference in expression profiles of histamine receptors (HRs) in nodose ganglia (NG) and individual identified NGNs to provide deeper insights into the mechanisms involved in sexual dimorphism of asthma. Western-blot and SYBR green RT-PCR showed that H₂R and H₃R were highly expressed in NG of females compared with males and downregulated in ovariectomized females. H₁R was equally expressed in NG of both sexes and not altered by ovariectomy. Furthermore, this highly expressive H₂R and H₃R were distributed in both myelinated and unmyelinated NGNs isolated from adult female rats by immunofluorescence and single-cell RT-PCR. H₃R widely distributed in all tested neuron subtypes and its expression did not show significant difference among neuron subtypes. H₂R was widely and highly expressed in low-threshold and sex-specific subpopulation of myelinated Ah-types compared with

myelinated A- and unmyelinated C-type NGNs. Unexpectedly, weak expression of H₁R was detected in both myelinated and unmyelinated NGNs by immunofluorescence, which was further confirmed by single-cell RT-PCR. Our results suggest that the sexual dimorphism in the expression of H₂R and H₃R in vagal afferents very likely contributes, at least partially, to the gender difference in prevalence and severity of asthma. © 2015 Published by Elsevier Ltd. on behalf of IBRO.

Key words: histamine receptor, vagal afferents, asthma, gender difference, electrophysiology.

INTRODUCTION

A number of studies have demonstrated that the prevalence and severity of asthma show striking difference and closely relate to gender and age. However, no single explanation has been able to fully explain the disparities in recent perspective reviews (Kynyk et al., 2011; Townsend et al., 2012). Exploring the possible mechanisms in the sexually dimorphic presentation of asthma, therefore, is an important issue to establish effective and individualized asthma management strategies for all patients. Sensory nerves have long been believed to play an important pathophysiological role in asthma (Undem and Carr, 2002). Stimulating vagal sensory fibers produce a variety of effects including bronchoconstriction (Lundberg et al., 1983; Forsberg et al., 1988), mucous secretion (Tokuyama et al., 1990), microvascular leakage (Lundberg et al., 1983) and cough (Forsberg et al., 1988), all of which are characteristic symptoms of asthma.

Histamine, a major inflammatory mediator released from mast cells, can sensitize unmyelinated C-type vagal afferents via histamine H₁ receptor (H₁R) activation (Higashi et al., 1982; Leal-Cardoso et al., 1993; Undem and Weinreich, 1993; Jafri et al., 1997; Yu et al., 2007), and then enhance the reflex response of vagal afferents to other stimuli such as mechanical or chemical stimuli (Lee and Morton, 1993; Mizumura et al., 1994; Michaelis et al., 1998). Histamine can also inhibit cholinergic neurotransmission in guinea-pig airways via histamine H₃ receptor (H₃R) activation in vagal afferents, which further inhibits the airway smooth muscle

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† These authors contributed equally for this work. **Abbreviations:** ANOVA, analysis of variance; AP, action potential; APD₅₀, AP duration measured at 50% amplitude of AP; APFT, AP firing threshold; UV_{MAX}, the maximal upstroke velocity of depolarization; HR, histamine receptor; NG, nodose ganglion; NGNs, nodose ganglion neurons; OVX, ovariectomized; PBS, phosphate-buffered saline; SD, Sprague–Dawley.

52 contraction elicited by vagal nerve stimulation (Ichinose
53 et al., 1989). Other reports demonstrated that the activa-
54 tion of H₃R could inhibit neurogenic microvascular leak-
55 age in guinea-pig airways by prejunctional inhibition of
56 neuropeptide release from airway vagal sensory nerves
57 (Ichinose et al., 1990) and protect the lung against
58 acetylcholine- and capsaicin-induced edema via a pre-
59 junctional modulatory effect on the C-fibers (Delaunois
60 et al., 1995). All of these functions for histamine and
61 histamine receptors (HRs) in vagal afferents are probably
62 involved in the pathophysiology of asthma. Recently, a
63 study demonstrated that lafutidine, a histamine H₂ recep-
64 tor (H₂R) antagonist, could inhibit vagal afferent signaling
65 of a gastric acid insult in adult female Sprague–Dawley
66 (SD) rats (Edelsbrunner et al., 2009), suggesting that
67 H₂R may also play a role in vagal afferents. Although all
68 above evidences indicate indirectly, from a pharmacolog-
69 ical point of view, that histamine H₁R, H₂R and H₃R may
70 express in vagal afferents, there is no direct molecular
71 biological evidence so far to confirm in this regard. To
72 our knowledge, there was only one direct study
73 (Kashiba et al., 1999) using *in situ* hybridization histo-
74 chemistry showing that none of nodose ganglion neurons
75 (NGNs) were labeled with H₁R cDNA probe in male
76 guinea pigs, which contradicts with the electrophysiological
77 studies showing that histamine sensitizes vagal afferents
78 by activating H₁R (Leal-Cardoso et al., 1993; Udem and
79 Weinreich, 1993; Jafri et al., 1997).

80 In our previous study, we surprisingly found that in
81 adult female SD rats, histamine could sensitize all types
82 of NGNs including a low-threshold and sex-specific
83 myelinated Ah-type neurons (Li et al., 2013) that exist
84 predominantly in adult female rats and rarely in adult male
85 rats (Li and Schild, 2007; Li et al., 2008; Qiao et al., 2009;
86 Santa Cruz Chavez et al., 2014). In stark contrast, his-
87 tamine only sensitizes unmyelinated C-type NGNs with-
88 out any effects on myelinated A-type NGNs in adult
89 male SD rats (Li et al., 2013), which is consistent with
90 the results from male species in other reports (Higashi
91 et al., 1982; Udem and Weinreich, 1993). These findings
92 in our laboratory highlight a potential mechanism involved
93 in the basic pathophysiology in the sexually dimorphic
94 presentation of asthma; meanwhile, these data also
95 suggest that the expression profiles of HRs in vagal
96 afferents are probably different between adult male and
97 age-matched female rats. Moreover, because of the
98 important functions of H₁R, H₂R and H₃R in vagal affer-
99 ents as described above, to study the gender different
100 expression of these receptors would have important impli-
101 cations for better understanding the gender difference of
102 histamine-mediated effects in vagal afferents, such as
103 histamine-mediated vagal afferent sensitization or cholin-
104 ergic neurotransmission inhibition or neurogenic
105 microvascular leakage inhibition in airways, which proba-
106 bly contribute, at least partially, to the sexually dimorphic
107 presentation of asthma.

108 Based on these considerations, the aim of the present
109 study is to detect the expression of H₁R, H₂R and H₃R
110 and reveal the sexually dimorphic expression of these
111 receptors in nodose ganglia by using western-blot and
112 SYBR Green RT-PCR. Furthermore, afferent-specific

expression of HRs in individual identified neurons, with
known afferent fiber types, isolated from adult female
rats is also detected using single-cell RT-PCR. Our
results will give insight into the mechanisms involved in
the sexual dimorphic presentation of asthma.

EXPERIMENTAL PROCEDURES

Animals

Adult male and age-matched female SD rats (180–250 g)
were provided by Weitong-Lihua Experimental Animal
Tech Corp (Grade: SPF, certificate #: SCXK, 2012-
0001, Beijing, China). Ovariectomy was performed as
previously described (McCallister et al., 2013; Qiao
et al., 2013). Briefly, the dorsal surgical area of anes-
thetized intact females was shaved and swabbed with
surgical scrub. Bilateral flank incisions were made to pro-
vide access to the peritoneal cavity. Both ovaries and the
majority of oviducts were exteriorized through the muscle
wall incision. The surgical wounds were sutured closed
and a topical antibiotic was applied. The animals were
allowed to recover and kept under normal condition for
at least 1 month before study performed. The reason we
chose 1 month as the recovery time of ovariectomized
(OVX) rats was based on a report, in which blood
17β-estradiol (E₂) concentration of female SD rats
(180–200 g) undergone OVX surgery was markedly
decreased from two weeks after the OVX operation com-
pared with non-OVX female rats and remained at a stably
low level throughout the experimental period of 12 weeks
(Wang and Abdel-Rahman, 2002). Experimental proto-
cols were pre-approved by the Institutional Animal Care
and Use Committee of School of Medical Science at
Harbin Medical University.

Drug and chemical agents

Primers and SuperScript™ III CellsDirect cDNA Synthesis
System were obtained from Invitrogen (Carlsbad, CA,
USA). Antibodies were obtained from Alomone Lab.
(Jerusalem, Israel). All other experimental materials
were obtained from Fisher Scientific or other regular
commercial sources.

Real Time SYBR-green quantitative PCR

Total RNAs were extracted using the TRIzol® Reagent
(Invitrogen) according to the manufacturer's instructions.
The cDNAs were synthesized using the Reverse
Transcription Kit (Applied Biosystems). The reverse
transcription conditions were as follows: 25 °C for
10 min, 37 °C for 120 min, 85 °C for 5 min. RT-PCR was
performed using the SYBR-Green Master Mix (Applied
Biosystems) kit with a 20-μl reaction volume containing
1 μl of reverse transcription product, 10 μl of
SYBR-Green, 1 μl of PCR forward primer, 1 μl of PCR
reverse primer and 7 μl of double-distilled water
(ddH₂O). Quantitative PCR reactions were run on an
ABI 7500 fast Real-Time PCR System (Applied
Biosystems) using 96-well reaction plates and began
with an initial cycles of 95 °C for 10 min followed by

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