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REVIEW 2

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THE ROLE OF THE IMMUNE SYSTEM IN CENTRAL NERVOUS 3 SYSTEM PLASTICITY AFTER ACUTE INJURY 4

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- 22 Abstract—Acute brain injuries cause rapid cell death that activates bidirectional crosstalks between the injured brain and the immune system. In the acute phase, the damaged CNS activates resident and circulating immune cells via the local and systemic release of soluble mediators. This early immune activation is necessary to confine the injured tissue and foster the clearance of cellular debris, which would ultimately bring the inflammatory reaction to a close. In the chronic phase, a sustained immune activation is described in many CNS disorders, and the degree of this prolonged response has variable effects on the spontaneous brain regenerative processes. The challenge for treating acute CNS damages is to understand how to optimally engage and modify these immune responses, thus providing new

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Abbreviations: BBB, blood-brain barrier; BDNF, brain-derived neurotrophic factor; CSPG, chondroitin sulfate proteoglycans; DC, dendritic cells; ECM, extracellular matrix; GAP, growth-associated protein; IFN, type 1 interferon; IGF, insulin-like growth factor; IL, interleukin; MC, mast cells; MMP, matrix metalloproteinases; MPO, myeloperoxidase; NGF, nerve growth factor; NSC, neural stem cells; NT, neurotrophins; NVU, neurovascular unit; PID, peri-infarcts depolarizations; PRR, pattern recognition receptors; ROS, reactive oxygen species; SCI, spinal cord injury; SDF-1 α , stromal cell-derived factor-1a; TGF, transforming growth factor; SVZ, subventricular zone; Q2 TH, T helper; VEGF, vascular endothelial growth factor.

strategies that will compensate for tissue lost to injury. Here we have reviewed the available information about the role and function of the innate and adaptive immune responses in influencing CNS plasticity during the acute and chronic phases of recovery after injury. We have examined how CNS damage evolves along the activation of main cellular and molecular pathways that ultimately are associated to intrinsic repair, neuronal functional plasticity and facilitation of tissue reorganization.

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Key words: CNS plasticity, immune system, stroke, spinal cord injury.

Contents		24
Introduction	00	25
BBB damage and reactive gliosis	00	26
Neuronal functional plasticity	00	27
Reparative regeneration	00	28
Conclusions	00	29
Acknowledgments	00	30
References	00	31
		32

23

33

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INTRODUCTION

Although considered for many years an immune privileged 35 tissue, it is today well accepted that the CNS is engaged in 36 an intense bidirectional communication with the immune 37 system. The CNS physiologically controls peripheral 38 immunity through complex humoral signaling and via the 39 direct activation of neuronal pathways that include the 40 hypothalamic-pituitary-adrenal axis and the autonomic 41 nervous system (An et al., 2014). The hypothalamus 42 normally suppresses the release of pro-inflammatory 43 cytokines from T cells, monocytes and macrophages, 44 while promoting the systemic release of anti-inflammatory 45 cvtokines, such as interleukin (IL)-10 (Chamorro et al., 46 2012). Similarly, the release of noradrenaline from the 47 autonomic centers and peripheral organs (including the 48 adrenal medulla, liver and spleen) induces a constitutive 49 anti-inflammatory phenotype in circulating immune cells 50 (Meisel et al., 2005). The immune system is in turn respon-51 sible for CNS development, surveillance and response to 52 damage. In the developing brain, a large percentage of 53

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123

2

the processes underlying neurogenesis and dynamic 54 pruning (i.e. the selective degeneration of whole or parts 55 of dendrites and axon collaterals) is mediated by resident 56 immune cells (Besedovsky and Rey, 2007; Boulanger, 57 2009). Later in adulthood, both resident and circulating 58 immune cells function as primary guardians of the CNS 59 and their sentinel duties contribute to the maintenance of 60 61 normal homeostasis (Chamorro et al., 2012; Ousman and Kubes, 2012). Immune mechanisms are indeed 62 responsible for the constant remodeling of neural circuits, 63 memory consolidation, hippocampal long-term potentia-64 tion and neurogenesis in response to everyday environ-65 mental stimuli (Meisel et al., 2005; Yirmiya and Goshen, 66 67 2011).

After focal damage, the lesioned area undergoes acute 68 loss of function and neurodegeneration, which are later 69 followed by a regenerative response finalized at restoring 70 both structures and functions. As the first line of defense 71 in the CNS, the immune system provides the earliest 72 responses against acute brain injury, consisting of both 73 physical and chemical barriers created by innate immune 74 cells (microglia/macrophages, neutrophils, and natural 75 killer cells) and the complement system (Gelderblom 76 77 et al., 2009). In this acute phase immune cells actively 78 participate in the disruption of the blood-brain barrier 79 (BBB), remodeling of the extracellular matrix (ECM), and 80 activation of glial cells (reactive gliosis), while protecting 81 neurons from increasing excitotoxicity, calcium release and free radicals (Dirnagl et al., 1999). This first intense 82 systemic immune activation orchestrates the clearance of 83 necrotic debris and the containment of the initial damage 84 (Kamel and ladecola, 2012). 85

The role of the immune system in the following 86 regenerative phase within the CNS is however far from 87 being fully elucidated. The CNS copes with injury and 88 loss of function by enacting a variety of functional and 89 90 structural changes in neural pathways and synapses, 91 which are commonly referred to as CNS plasticity. In particular, a first phase of functional plasticity 92 characterized by dendritic reorganization and axonal 93 sprouting is followed by a second phase of structural 94 neuroanatomical plasticity (generation of new neurons 95 and vessels) ultimately leading to the formation of novel 96 97 connections within the damaged brain (Wieloch and 98 Nikolich, 2006). The components of both the innate and adaptive (T and B lymphocytes) immune responses pro-99 foundly shape functional and structural plasticity of the 100 injured CNS by priming (or hindering) brain recovery via 101 modulation of intrinsic growth properties and extrinsic 102 growth-regulatory cues (Martino et al., 2011). 103

104 It is increasingly clear that many of the events that characterize the first acute neurodegeneration are linked 105 (directly or indirectly) with the following regenerative 106 phase, and that the immune activation within the CNS 107 must be interpreted in a continuum between degenerative 108 and reparative processes (Hermann and Chopp, 2012). 109 In this review we focus on the role exerted by the innate 110 and the adaptive immune response in regulating CNS plas-111 ticity through the different phases of acute injury and sub-112 sequent recovery. In particular, we explore the ability of 113 the immune system to modulate the initial BBB damage 114

and glial activation, the following functional plasticity of 115 neurons, and the final reparative regeneration of the injured 116 CNS (Fig. 1). Since most of currently available evidences 117 related to the innate and adaptive immune responses after 118 damage derive from CNS focal sterile injuries, we mainly 119 focus on describing the pathophysiology and the evolution 120 of acute (focal) damage after experimental ischemic stroke 121 and spinal cord injury (SCI). 122

BBB DAMAGE AND REACTIVE GLIOSIS

The BBB is made by endothelial cells, pericytes, 124 astrocytes and ECM that, together with neurons, are 125 organized in a complex cellular system called the 126 neurovascular unit (NVU) (Abbott et al., 2006). Upon 127 ischemic brain injury, the NVU undergoes intense early 128 changes that comprise failure of ion pumps, overaccumu-129 lation of intracellular sodium and calcium, loss of mem-130 brane integrity and necrotic cell death. Release of 131 damage-associated molecular patterns (DAMPs) from 132 necrotic cells activates pattern recognition receptors 133 (PRRs) of the resident immune cells (microglia) that 134 include Toll-like receptors (TLRs), RIG-1-like receptors 135 (RLRs), NOD-like receptors (NLRs), AIM2-like receptors 136 (ALRs) and C-type lectin receptors (Hanke and Kielian, 137 2011; Chamorro et al., 2012). Activation of PRRs on 138 microglial cells triggers downstream signaling pathways, 139 such as the nuclear factor kappa-light-chain-enhancer of 140 activated B cells (NF-kB), the mitogen-activated protein 141 kinase (MAPK) and type 1 interferon (IFN) pathway, 142 which in turn upregulate proinflammatory cytokines, che-143 mokines, costimulatory signals and reactive oxygen spe-144 cies (ROS) (Takeuchi and Akira, 2010). Excessive 145 oxidative damage leads to dysfunction of endothelial cells. 146 degradation of tight junctions and modification of integrins 147 on the abluminal endothelial membrane (Hermann and 148 Elali, 2012). Cell adhesion molecules (CAMs), such as 149 the intercellular cell adhesion molecule (ICAM-1) or the 150 vascular cell adhesion molecule (VCAM-1), and P-selec-151 tins are then upregulated on the endothelium and ulti-152 mately favor the recruitment of blood-borne leukocytes 153 to the ischemic damage. 154

Infiltrating neutrophil granulocytes are the first 155 circulating immune cells to appear within the ischemic 156 lesion and they virtually overwhelm the ischemic 157 hemisphere by 3 days post-reperfusion (Gelderblom 158 et al., 2009). Upon infiltration, neutrophils start producing 159 inducible nitric oxide synthase (iNOS), an enzyme that gen-160 erates toxic amounts of nitric oxide (NO), and release both 161 matrix metalloproteinases (MMPs) and myeloperoxidase 162 (MPO) (Justicia et al., 2003). Release of MMP-9, as well 163 as the upregulation of MPO within the ischemic tissue, con-164 tribute to the further down-regulation of junctional proteins 165 and are the main contributors to the first derangement of 166 the BBB (Bao Dang et al., 2013; Peruzzotti-Jametti et al., 167 2013). Initial BBB disruption is soon enhanced by ECM 168 degradation, which participates in the secondary ischemic 169 brain damage by permitting serum elements to enter the 170 perivascular space (Asahi et al., 2001; Elali et al., 2011). 171 Resident macrophages and mast cells (MCs) become fur-172 ther activated, leading to the release of vasoactive media-173

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