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# Both ozone exposure and soil water stress are able to induce stomatal sluggishness

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#### 1. Introduction

Tropospheric ozone  $(O_3)$  is an important secondary pollutant resulting from emission of volatile organic compounds and nitrogen oxides, and is also recognized as a significant greenhouse gas (Bytnerowicz et al., 2007; Serengil et al., 2011). Ozone is seriously phytotoxic and causes negative effect on plants (e.g., NIES, 1980, 1984; Omasa et al., 2002; Paoletti, 2007). Stomatal O<sub>3</sub> uptake is crucial for assessing the adverse effect of O<sub>3</sub> on plants (Omasa et al., 1979; UNECE, 2004; Paoletti and Manning, 2007; Grulke et al., 2007a; Cieslik et al., 2009). However, our understanding about stomatal responses to O<sub>3</sub> is still imperfect (Paoletti and Grulke, 2005). Previous studies reported that O<sub>3</sub> generally induces both stomatal closure (Wittig et al., 2007, 2009) and sluggishness of stomatal response to change of environmental factors (Paoletti and Grulke, 2005, 2010). Because plants live in a fluctuating environment, both steady-state stomatal conductance and stomatal dynamics play an important role in regulating leaf gas exchange. Ozone-induced sluggishness of stomata has been reported in response to change in photosynthetic photon flux density (PPFD) (Reich and Lassoie, 1984; Reiling and Davison, 1995; Paoletti, 2005; Grulke et al., 2007a; Paoletti and Grulke, 2010), vapor pressure deficit (VPD) (Grulke et al., 2007b) and severe water stress imposed by severing a leaf (Paoletti, 2005; Paoletti et al., 2009; Mills et al., 2009). Such aberrations may increase nighttime transpiration, as reported for

#### ABSTRACT

We tested whether short-term exposure to realistic ozone pollution ( $\leq$ 150 ppb, 1 h) and soil water stress (soil water content  $\leq$ 15%) slow stomatal dynamics in an ozone-sensitive cultivar of snapbean. Both ozone exposure and water stress caused stomata to be sluggish in the degree of closure after leaf severing, while ozone also delayed the time the closing signal was perceived. Ozone-induced aberrations lasted up to the night and caused incomplete closure of stomata. No synergic effect was observed in the dynamic measurements. In contrast, at steady-state, water stress protected the plants from the negative ozone effects on stomatal conductance. Ambient ozone peaks may thus cause sluggish stomatal response and increase leaf water loss both under well watered and drought conditions.

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several tree species exposed to daytime  $O_3$  exposure in controlled (Skärby et al., 1987; Matyssek et al., 1995; Wieser and Havranek, 1995; Günthardt-Goerg et al., 1997; Grulke et al., 2007c) and ambient conditions (Grulke et al., 2004). Ozone is known to induce up-regulation of ethylene emission, which has been suggested to be responsible for a reduction in stomatal sensitivity to abscisic acid (ABA) and thus to closure (Wilkinson and Davies, 2010). Several stressors, including water deficit, are known to stimulate ethylene production by plants (Morgan and Drew, 1997).

Climate change is expected to increase  $O_3$  levels and alter precipitation regimes, regionally increasing the probability of drought (Ciais et al., 2005). Drought has the capacity to limit  $O_3$ injury through stomatal closure (Tingey and Hogsett, 1985), while  $O_3$ -induced impairment of stomatal response may increase susceptibility to drought (Nali et al., 2004).

Our main objective was to test whether short-term exposure to realistic O<sub>3</sub> pollution and soil water stress, singly and in combination, may slow stomatal dynamics in an ozone-sensitive cultivar of snapbean (*Phaseolus vulgaris*, S156).

#### 2. Materials and methods

Seeds of the ozone sensitive cultivar S156 of snapbean developed at the Raleigh USDA-ARS (Burkey and Eason, 2002; Flowers et al., 2007; Booker et al., 2009), were planted in 17-cm (1.7–1) pots, filled with sand:peat:soil = 1:1:1 (v:v:v). Seed were planted over several days so that same-age (4-week old) plants were used in the experiment. All plants were grown in a room with controlled environmental conditions (air temperature of 20 °C, PPFD

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**Fig. 1.** Changes in soil water content ( $\pm$ S.D., n = 18) (%)and predawn leaf water potential ( $\pm$ S.D., n = 2-3) after irrigation until field capacity. Different letters indicate significant differences among mean values (Tukey HSD test, P < 0.05).

of  $500 \,\mu\text{mol}\,\text{m}^{-2}\,\text{s}^{-1}$ , and relative humidity of 55%). Well-watered plants were supplied with water every day and in particular immediately before the ozone exposure, while water-stressed plants received no irrigation for 10 days before O<sub>3</sub> fumigation. Soil water content was more than 30% in well-watered plants and 6–15% in water-stressed plants, corresponding to a predawn leaf water potential of -0.32 MPa and -0.53 MPa, respectively (Fig. 1). Water content was measured in 18 pots with an ECH2O EC-5 soil moisture sensor (Decagon Devices, Pullman WA, USA) at root level. Leaf water potential was measured at dawn in a selection of 2–3 plants per day by means of an SKPM 1400 chamber pressure (Skye, Powys, UK).

The fully expanded central leaf of the second trifoliate leaf was selected as a target leaf. After 1-h exposure to  $1000 \,\mu mol \,m^{-2} \,s^{-1}$ light, steady state leaf gas exchange was measured with a portable infra-red gas-analyzer (CIRAS-2 PP Systems, Herts, UK), equipped with a  $2.5 \text{ cm}^2$  leaf cuvette which controlled leaf temperature (20°C), leaf-to-air vapour pressure deficit (0.9 kPa), saturating light  $(1800 \,\mu\text{mol}\,\text{m}^{-2}\,\text{s}^{-1})$  and CO<sub>2</sub> concentration (365 ppm). Ozone exposure to one of four levels  $(\pm SD)$ , i.e. low  $(48 \pm 6.7 \text{ ppb})$ , middle  $(87 \pm 9.4 \text{ ppb})$ , high  $(150 \pm 10.9 \text{ ppb})$ , and control  $(0 \text{ ppb}, \text{ no } O_3)$ , was then carried out for 60 further min by a web fumigation system (Velikova et al., 2005; Pinelli and Tricoli, 2008). Ozone was added by an O<sub>3</sub> generator (Model Heliozon, Milano, Italy) to the fumigating air for the target leaf through a teflon tube. The concentration around the leaf was recorded with an ozone monitor (Mod. 205, 2B Technologies, Boulder CO, USA), and adjusted through mass flow controllers (Mod. GFC171S A alborg). At 30 min after the end of O<sub>3</sub> exposure, steady-state stomatal conductance (g<sub>s</sub>) was measured again. When gs reached equilibrium under constant light at 1800  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, the methodology described by Paoletti (2005) was applied to assess dynamic variations of gs after cutting the leaf petiole (Fig. 2). Data were logged at 1 min intervals in the 45 min after severing. In the experiment, two phases of  $g_s$  response were observed (Fig. 2). At first, gs increased for Iwanoff effect. This transient increase called as the transient 'wrong-way response' (WWR) is due to a difference in turgor pressure between guard cell and epidermal cells (Omasa and Maruyama, 1990; Powles et al., 2006). Subsequently, stomatal conductance decreased with increasing leaf water stress. WWR duration and magnitude of WWR and gs decrease at 45 min ( $\Delta g_s$ ) were recorded.

After  $O_3$  exposure and steady-state measurements, 22 plants were placed in the dark for 10 h. Nocturnal steady-state  $g_s$  was measured with leaf temperature of 20 °C, leaf-to-air vapour pressure deficit of 0.9 kPa, no exposure to light and  $CO_2$  concentration of 365 ppm.

Data were checked for normal distribution and homogeneity of variance (Levene's test). Percents were arcsine square root



**Fig. 2.** Time courses of stomatal conductance  $(g_s)$  after severing a leaf at time 0. Graph A shows an example of calculation of WWR magnitude and duration, and of the degree of  $g_s$  decrease  $(\Delta g_s)$ . At first,  $g_s$  showed a transient increase called 'wrongway response' (WWR) and a subsequent decrease with increasing leaf water stress. Graph B shows the time course of average  $g_s$  ( $\pm$ SE) under well-watered condition (soil water content > 30%) and water-stressed condition (soil water content = 6–15%) in control (0 ppb O<sub>3</sub>) and ozone-exposed (150 ppb O<sub>3</sub>) leaves.

transformed prior to analysis. Effects of soil water status and  $O_3$  exposure were tested using two-way analysis of variance (ANOVA). Results were considered significant at p < 0.05. Differences among means were tested by Tukey's HSD test. Statistical analysis was performed with STATISTICA software (6.0, StatSoft Inc., Tulsa, OK, USA), according to Statsoft (2001).

#### 3. Results

#### 3.1. Steady-state and dynamic stomatal responses

Ozone exposure induced a decline of steady-state  $g_s$  under wellwatered conditions (Fig. 3).  $g_s$  was 73% in the low O<sub>3</sub> treatment and 25% in the high O<sub>3</sub> treatment compared to the control plants. A reduced soil water availability significantly reduced  $g_s$  relative to well-watered plants and resulted in no effect of O<sub>3</sub> on  $g_s$ .

After severing a leaf, two phases of  $g_s$  response were observed (Fig. 2): a transient increase as WWR and then a linear decrease. Ozone exposure increased WWR duration from  $11.9 \pm 2.7$  min in the control plants to  $17.1 \pm 3.2$  min in the high O<sub>3</sub> treatment, while the effect of water stress was not significant (Fig. 4A). Magnitude of WWR was not affected by both O<sub>3</sub> and soil water availability (Fig. 4B). Ozone reduced the degree of stomatal closure over time ( $\Delta g_s$ ) in both soil water conditions (Fig. 4C). Also soil water deficit reduced  $\Delta g_s$  relative to the optimal soil water availability. In well-watered plants, the high O<sub>3</sub> treatment resulted in smaller stomatal closure than in control leaves (48% vs. 75%). In

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