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Antioxidant cerium oxide nanoparticle hydrogels for cellular 4 01 encapsulation

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ABSTRACT

Oxidative stress and the resulting radical by-products cause significant toxicity and graft loss in cellular transplantation. Here, the engineering of an auto-catalytic, antioxidant, self-renewing cerium oxide nanoparticle (CONP)-composite hydrogel is reported. This enzyme-mimetic material ubiquitously scavenges ambient free radicals, with the potential to provide indefinite antioxidant protection. Here, we evaluated the potential of this system to enhance the protection of encapsulated beta cells. Co-incubation of CONPs, free in solution with beta cells, demonstrated potent cytoprotection from superoxide exposure; however, phagocytosis of the CONPs by the beta cells resulted in cytotoxicity at concentrations as low as 1 mM. When CONPs were embedded within alginate hydrogels, the composite hydrogel provided cytoprotection to encapsulated beta cells from free radical attack without cytotoxicity, even up to 10 mM concentrations. This nanocomposite hydrogel has wide applicability in cellular transplantation, with the unique advantage of localization of these potent antioxidant CONPs and their capacity for sustained, long-term scavenging.

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46 1. Introduction

47 Oxidative stress is defined as the imbalance between the production of oxidants or reactive oxygen species (ROS), such as 48 49 superoxide (O_2^-) and hydrogen peroxide (H_2O_2) , and their elimina-50 tion via antioxidants, such as superoxide dismutase (SOD) and cat-51 alase. Sustained oxidative stress results in significant destruction of cellular structures and functions and has been implicated in 52 numerous pathological conditions, such as atherosclerosis, cancer, 53 renal disease, and diabetes [1–3]. Given that a toxic oxidative 54 milieu can be generated via hypoxia, cytokines, and inflammation, 55 cellular transplants are particularly susceptible to oxidative dam-56 age, resulting in increased cell death and decreased efficacy of 57 implants [4,5]. Protecting cellular grafts from oxidative damage 58 59 due to this noxious environment is particularly challenging in 60 the context of pancreatic islet transplantation for treatment of

Abbreviations: CONP, cerium oxide nanoparticle; SO, superoxide; SOD, superoxide dismutase; XA, xanthine; XO, xanthine oxidase; ROS, reactive oxygen species. * Corresponding author at: Department of Biochemistry and Molecular Biology,

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Type 1 diabetes mellitus, due to the inherently low gene expression and activity of important antioxidant enzymes in pancreatic islets [6–8]. Oxidative damage of islets following transplantation is one of the contributing factors resulting in graft destabilization and decreased long-term efficacy [9].

The encapsulation of transplanted donor islets within semi-permeable polymers is an appealing method for protecting allogeneic grafts from detrimental host responses [10–12]. The encapsulating polymer permselectivity permits passage of nutrients and release of secreting proteins or waste products, but blocks direct host cell interactions with graft cells. The most commonly used encapsulation material is alginate, due to its high biocompatibility, ease in encapsulation method, and demonstrated efficacy in small animal models [13–15]. While cellular encapsulation aids in reduction of generalized host cell responses via blocking direct cell–cell interactions, this strategy fails to protect donor cells from soluble byproducts of the inflammatory response, in particular ROS [16–18].

Numerous anti-oxidant agents, such as edaravone and78gliclazide, have been incorporated into islet transplants, either via79systemic infusion, pre-culture treatment, or transgenic overexpression, with varying degrees of protective effects [19–23]; however,81

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82 these approaches are limited by the need for systemic delivery, the 83 decreased duration of effect, and the complexities of transfection, 84 respectively. Notable biomaterial strategies have sought to scav-85 enge ambient free radicals via supplementation of encapsulation 86 polymers with antioxidant enzymes such as SOD and catalase 87 [24–28]. Inevitably, the catalytic reactivity of these free radical 88 scavenging agents is exhausted, resulting in transient protection. 89 A more potent approach to combat the continual inflammatory 90 assault to the transplant would be in the development and applica-91 tion of a sustainable anti-oxidant mimetic.

92 The unique redox properties of selected metal oxides, e.g., 93 yttrium and cerium, have been recently explored as scavenging agents for cellular oxidative stress [29]. The oxide form of the rare 94 95 earth element cerium, found in the lanthanide series of elements, 96 has the ability to cycle between its cerium(III) and cerium(IV) oxi-97 dation states, due to a lattice structure with a high tolerance for 98 reversible oxidation/reduction [30]. Cerium oxide nanoparticles 99 (CONPs) exhibit enhanced catalytic activity over bulk forms due 100 to increased surface area, resulting in an amplified number of available oxygen vacancies [30,31]. The oxidative state of CONP 101 102 appears related to its catalytic activity, whereby cerium(IV) corre-103 lates with catalase-like behavior and cerium(III) exhibits SOD mimetic responses [31,32]. The unique ability of CONPs to switch 104 105 their oxidative states between III and IV lends itself to its desirable 106 self-renewing property [33]. Further, CONPs have the potential to 107 provide broad free radical protection, with demonstrated quenching of hydroxyl radicals, superoxide, peroxide, and nitric oxide 108 109 [32.34 - 36].

CONP's potent scavenging capacity, with low loading volume 110 111 and theorized unlimited auto-catalytic potential, inspired explora-112 tion of their pharmaceutical potential, with the aim of reducing 113 oxidative damage in a variety of injury models. Co-culture of free CONPs with cells has resulted in radioprotective [34,37,38], cardio-114 115 protective [39], and neuroprotective effects [29] (for full review see 116 [40]). Selected studies have also explored the potential of CONPs to 117 protect beta cells and islets [41–43]. While highly promising, cyto-118 toxic effects have been observed, particularly for particle sizes 119 exceeding 100 nm or at concentrations higher than 1 mM (particle 120 sizes 3-50 nm), broad assessment of CONP toxicity is complicated 121 by the variable size, surface geometry, and zeta potential of the particles [31,34,44]. CONP cytotoxicity likely results from the cel-122 123 lular internalization and accumulation of the nanoparticles, as

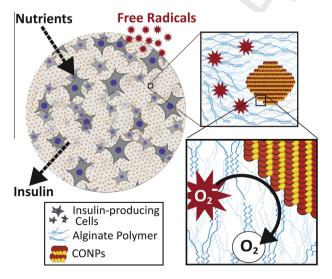


Fig. 1. Illustration of cerium oxide nanoparticle (CONP)-alginate composite hydrogel. Alginate microbead provides matrix for cellular encapsulation and permselectivity to permit nutrient diffusion in and insulin secretion out of the hydrogel. CONP, embedded within the alginate matrix, provides ubiquitous, renewable, antioxidant protection from external free radical damage.

autophagy-induced apoptosis, a common outcome of nanoparticle124phagocytosis, has been observed [40,44–46]. Additionally, many125cellular processes are mediated by intracellular free radical126signaling and internalization of CONPs may have unpredictable127long-term consequences on these processes [47].128

A means to mitigate the cytotoxicity of CONPs may be entrap-129 ment within an encapsulation hydrogel. This delivery strategy pro-130 vides the means for localization of particles to the site of interest, 131 minimizing their phagocytosis, while retaining their catalytic 132 potential. Herein, we sought to engineer a nanocomposite, anti-133 oxidant biomaterial via incorporation of cerium oxide nanoparti-134 cles within an encapsulating alginate hydrogel (see Fig. 1). The 135 potential of CONPs, embedded within a hydrogel, to retain their 136 catalytic and self-renewal activity was examined. The capacity of 137 CONPs and CONP-composite hydrogels to prevent ROS-induced 138 beta cell death, as well as enhance cytocompatibility, was also 139 evaluated. The benefits of this approach to provide the local pre-140 sentation of potent CONPs at the transplant site, thereby reducing 141 potential downstream or systemic effects, are discussed. 142

2. Experimental section

2.1. Materials

All chemicals were obtained from Sigma–Aldrich unless otherwise noted. 145

2.2. CONP synthesis

Dextran coated, cerium oxide nanoparticles (CONP) were syn-148 thesized using a method similar to published reports [33]. Dextran 149 coating was used to enhance stability of the CONPs in solution [48]. 150 A 1 mL solution of 1 M cerium(III) nitrate was mixed evenly with a 151 2 mL solution of 100 mM dextran T-10, added drop-wise to 6 mL of 152 ammonium hydroxide (30%), and stirred overnight. To remove 153 excess dextran and reaction by-products, CONP solutions were dia-154 lyzed against PBS using 30 kDa MWCO centrifuge filters (Millipore) 155 at 4000 rpm in 10 min intervals, until effluent pH was \sim 7.0. The 156 CONP concentration is expressed in mM, per convention, and cal-157 culated as described elsewhere [33]. CONP solutions were further 158 processed for analysis and cell culture by sonication, to prevent 159 CONP aggregation, and sterile filtration (0.2 μ m). Due to small size 160 of the particles, no detectable particle loss was observed during 161 filtration. 162

2.3. CONP solution characterization

All tests were performed at physiological pH. CONP size was 164 characterized by dynamic light scattering (DLS) using a DynaPro 165 Titan and Dynamics v6.0 software (Wyatt Technology), all samples 166 diluted in PBS (Gibco), where percent polydispersity represents the 167 standard deviation of detected peaks normalized to their mean 168 intensity value. A polydispersity >30% indicates low homogeneity. 169 CONP solution composition was characterized by Fourier trans-170 form infrared (FTIR) analysis on a Perkin-Elmer Spectrum 100 FTIR 171 Spectrometer (average of four scans with a resolution of 4 cm^{-1}) 172 using a lyophilized, processed sample. HR-TEM imaging was per-173 formed at 300 kV on a FEI Tecnai F30 TEM by the Advanced Mate-174 rials Processing and Analysis Center (AMPAC) at the University of 175 Central Florida (UCF). XPS analysis was performed on a Physical 176 Electronics 5400 ESCA, also at AMPAC UCF, and peaks identified 177 using PeakFit v4.12 (Systat Software, 8% Savitsky-Golay smoothing, 178 linear two-point baseline subtraction). CONP catalytic activity and 179 renewability were assessed by addition of CONPs (1 mM) to H₂O₂ 180 (1 mM) and spectral analysis performed on a Molecular Devices 181 SpectraMax M5 Microplate reader. CONP solutions were incubated 182

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