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2 Brief communication

High performance and reversible ionic polypeptide hydrogel based on charge-driven assembly for biomedical applications

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ABSTRACT

In the pursuit of new strategies for the design and synthesis of high performance, physically associated hydrogels, dynamic materials formed through electrostatic interactions can serve as a powerful model. Here, we introduce a convenient strategy to obtain biodegradable hydrogels from ABA triblock ionic polypeptides formed by mixing poly(L-glutamic acid)–block-poly(ethylene glycol)–block-poly(L-glutamic acid) (PGA–PEG–PGA) with poly(L-lysine)–block-poly(ethylene glycol)–block-poly(L-glutamic acid) (PGA–PEG–PGA) with poly(L-lysine)–block-poly(ethylene glycol)–block-poly(L-lysine) (PLL–PEG–PLL). The hydrogels showed tunable physical properties, high strength and reversible response. The reactive function groups in the ionic blocks can conjugate with oppositely charged drugs or proteins and allow for further modification. These ionic ABA triblock polyelectrolytes can also encapsulate intact cells without significantly compromising cell viability, suggesting that the hydrogels have excellent cytocompatibility. In vivo evaluation performed in rats with subcutaneous injection indicated that the gels were formed and degraded, and hematoxylin and eosin staining suggested good biocompatibility in vivo. In addition, these advantages, combined with the synthetic accessibility of the copolymer, make this cross-linking system a flexible and powerful new tool for the development of injectable hydrogels for biomedical applications.

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Hydrogels are three-dimensional (3-D) networks with high 47 48 water content which maintain their structural integrity. Due to 49 their good biocompatibility, flexibility, variable composition and 50 desirable physical characteristics which mimic physical properties 51 of tissues, potential responsiveness to certain stimulus and effec-52 tive loads for chemicals and organisms, the hydrogels are perfect candidate soft materials for many biomedical applications, includ-53 ing as cell scaffolds for tissue regeneration, as carriers for cell 54 encapsulation or drug/gene delivery and so on [1-5]. 55

In particular, in situ formed hydrogels have recently been 56 widely studied for their superiority in moldability and operation 57 [1,6]. They can be easily applied through mild gelation conditions 58 and in a minimally invasive manner. Following injection in vivo, 59 the hydrogels form constructs in situ, providing local biological 60 61 and mechanical cues that may enhance tissue regeneration or 62 responding to local surroundings that may control drug release [7,8]. In situ gelation can be obtained via chemical cross-linking 63 or via self-assembly by reversible interaction. Temperature, pH, 64

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hydrogen bonding, π - π stacking and electrostatic or hydrophobic interaction are examples of the most widely studied external stimuli for self-assembly [1,9–11]. Typically, drawbacks of such selfassembled hydrogels are poor mechanical properties and stability due to their generally weak intermolecular interactions.

Unlike the direct self-assembly of ligand molecules such as metal ions and small molecules as cross-linkers [12] or recombinant peptides and proteins as cross-linkers [13], the hydrogels formed by mixing the synthetic block copolyelectrolytes with oppositely charged polymer blocks showed tunable physical properties, reversible response, high stability, strength and resilience [14–16]. However, in the previous studies, these synthetic copoly-electrolyte hydrogels lacked biodegradability, limiting their biomedical applications.

Herein, we present a convenient strategy for the formation of biodegradable hydrogels from water-soluble ABA triblock ionic polypeptides, which formed by mixing poly(L-glutamic acid)-block-poly(ethylene glycol)-block-poly(L-glutamic acid) (PGA-PEG-PGA) with poly(L-lysine)-block-poly(ethylene glycol)block-poly(L-lysine) (PLL-PEG-PLL) (Scheme 1). The key to this strategy was the phase separation of ion-rich regions from the aqueous environment, which occurred on mixing two ABA triblock copolymers with oppositely charged anionic and cationic blocks

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30 September 2014

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H. Cui et al./Acta Biomaterialia xxx (2014) xxx-xxx

[15,16–18]. The hydrogel assembled and disassembled reversibly
through a pH-trigger operation at both high and low pH controlling
the level of electrostatic interactions. In contrast to mixing directly
the pure poly(glutamic acid) and polylysine, only the precipitate or
coacervate was obtained [19,20].

93 Synthetic ionic polypeptide hydrogels based on electrostatic 94 interaction have several advantages: (1) good water solubility and a low concentration of the unmixed copolyelectrolyte, which 95 facilitate the encapsulation and injection; (2) rapid gelation and 96 97 high modulus; (3) well-controlled ring-opening polymerization of 98 the amino acid NCA, producing symmetric triblock copolymers with a well-defined mass and composition, providing the resulting 99 materials with a high and tunable density of ionic groups; (4) the 100 ability of the charged ionic block to conjugate with oppositely 101 102 charged drugs or proteins to achieve high-efficiency encapsulation 103 and controllable release: (5) the presence of the reactive function 104 **Q2** units for the post-polymerization modification through EDC condensation chemistry. Not limited to the pairs of poly(glutamic acid) 105 and polylysine blocks, this strategy is rather flexible and can lead 106 to a library of well-defined, ABA triblock ionic copolyelectrolytes 107 108 of diverse block lengths and nature of ionic groups, such as pol-109 varginine, poly(aspartic acid) and their copolymers. Therefore, through the use of charge-driven assembly, a new design strategy 110 for robust hydrogels is obtained that could be widely used in a 111 112 variety of materials and potentially useful in biomedical applica-113 tions, such as cell scaffolds and drug carriers.

114 **2. Experimental section**

115 2.1. Materials

116PEG (Mn = 1500 and 4000 Da) was purchased from117Sigma–Aldrich without further purification. γ-Benzyl-L-glutamate

(BLG) and ε -benzyloxycarbonyl-L-lysine (ZLL) were purchased 118 from GL Biochem Co. Ltd. Amino-terminated poly(ethylene glycol) 119 $(NH_2-PEG-NH_2)$, γ -benzyl-L-glutamate-N-carboxyanhydride (BLG-120 NCA) and ε-benzyloxycarbonyl-L-lysine-N-carboxyanhydride (ZLL-121 NCA) were synthesized as described in previous work with slight 122 modification [21-23]. N,N-dimetylformamide (DMF) was dried 123 over calcium hydride (CaH₂) before vacuum distillation. All the 124 other reagents and solvents were purchased from Sinopharm 125 Chemical Reagent Co. Ltd and used as received. All chemicals were 126 of analytical grade or higher. 127

2.2. Synthesis of PGA–PEG–PGA and PLL–PEG–PLL copolymers

As shown in Scheme S.1, PGA–PEG–PGA (G) and PLL–PEG–PLL 129 (L) were synthesized through ring-opening polymerization (ROP) 130 of BLG-NCA and ZLL-NCA in DMF using NH₂–PEG–NH₂ as initiator 131 and followed by deprotection reaction, respectively. 132

A typical procedure for the preparation of PGA₄₄–PEG₉₁–PGA₄₄ 133 was as follows: NH₂-PEG₉₁-NH₂ (4.0 g, 1.0 mmol) was dissolved 134 in toluene (50 ml) and the residual water was removed by azeotro-135 pic distillation. Then a certain amount of BLG-NCA (21.0 g, 136 40.0 mmol) and anhydrous DMF (100 ml) were added to the dried 137 NH₂-PEG-NH₂. The reaction mixture was stirred at 25 °C for 3 days 138 under a dry nitrogen atmosphere. The different reactant feeding 139 molar ratios are listed in Table 1. The solution was precipitated 140 into an excess amount of diethyl ether to give the PBLG44-141 PEG₉₁–PBLG₄₄ triblock copolymers. Subsequently, the copolymer 142 was dissolved in dichloroacetic acid (mass/vol. 1/10) and HBr/ace-143 tic acid (33 wt.%, mass/vol. 1/3) was added. The deprotection reac-144 tion was conducted at 30 °C for 2 h and then the mixture was 145 precipitated into excessive diethyl ether. After dried under vac-146 uum, the precipitate was dissolved in 0.1 M NaOH, dialyzed with 147 distilled water and freeze-dried to give a PGA-PEG-PGA product 148



Scheme 1. Schematic representation of the general macromolecular design of oppositely charged ionic ABA triblock copolyelectrolytes and the formation of reversible hydrogels based on charge-driven assembly.

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