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## Choosing supplementary markers in forensic casework

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#### ABSTRACT

The vast majority of human familial identifications based on DNA end up with a well founded conclusion, normally using a standard set of genetic short tandem repeat (STR) loci. There are, however, a proportion of cases that show ambiguous results. For such occasions a number of different supplementary markers could be typed in order to gain further information. There are numerous markers available for such supplementary DNA typing, including STRs, deletion and insertion polymorphisms (DIPs), and single nucleotide polymorphisms (SNPs). The purpose of this work was to describe a precise method for decision making, aiming to aid the comparison of different sets of markers for different case scenarios in order to find the most efficient set for routine casework. Comparisons are based on a particular function relating the expected additional value of information from new data to the amount of information already obtained from initial data. The function can be computed approximately by approximating likelihood-based error rates using simulation. In this paper we focused on paternity investigations, more specifically the use of supplementary markers in cases where a smaller number of genetic inconsistencies make the matter inconclusive. We applied the method to a comparison of three different kits: Investigator HDplex (STRs), Investigator DIPplex (DIPs), and the SNPforID-plex (SNPs) to study their efficiencies in gaining information in different case scenarios involving various alternative relationships between the tested man and the tested child. We show that the Investigator HDplex was the most efficient set of supplementary markers for the standard paternity case. However, for paternity cases with a close relative being the alternative father, the Investigator HDplex and the SNPforID-plex showed similar patterns in their ability to deliver a well-founded conclusion. The Investigator DIPplex was the least efficient set.

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#### 1. Introduction

For a long period of time the CODIS short tandem repeat (STR) loci [1] have provided the basis for DNA analysis for human identification. Until a few years ago, there were only a smaller number of kits commercially available that included other DNA markers. During the past couple of years there has been an expansion on the forensic STR kit market, which now offers more diverse kits with a larger number of new genetic loci. One reason for this increase is the expansion of the core European standard set (ESS) [2]. Apart from the standard STR markers, there are also kits available for forensic use that include biallelic markers such as DIPs

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http://dx.doi.org/10.1016/j.fsigen.2014.06.019 1872-4973/© 2014 Elsevier Ireland Ltd. All rights reserved. (Deletions and Insertion Polymorphisms) and SNPs (Single Nucleotide Polymorphisms) [3,4].

Although generally a good thing that a larger number of sets of markers are available to choose from, there is a challenge in choosing the best kit for a forensic laboratory's routine casework. Apart from technical issues with each new kit, the usefulness (in terms of efficiency to solve specific cases) of the additional markers depends on the type of marker, population uniqueness, number of alleles, mutation rates, and which types of genetic relationships are to be tested, among other things. Even though there are various general measurements of the genetic diversity of a given set of markers (e.g. overall match probability, power of discrimination, power of exclusion), it can, for numerous applications, be difficult to compare different sets of markers only on the basis of such general efficiency parameters.

The purpose of this paper is to describe a method for decision making when having multiple sets of markers to choose from. Furthermore, we have applied our method in a comparison of three







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different sets of markers; one set that includes 11 STRs (Investigator HDplex, Qiagen), a second that includes 30 biallelic markers of the type Insertion/Deletions (InDels) (Investigator DIPplex, Qiagen), and a third set that includes 52 biallelic markers of the type Single Nucleotide Polymorphisms (SNPs) (SNPforID). The goal was to find the most efficient set to be used as supplementary markers in paternity investigations.

The chosen sets of markers represent two different types. namely biallelic markers (SNPs [3] or InDels [4]) and additional STRs. These types have different characteristics. The main advantage of using SNPs/Indels in paternity testing is their low mutation rate, thus confirming a genetic inconsistency as a true genetic exclusion. The disadvantage is that generally only two alleles exist per locus making it harder to find true genetic exclusions. For example, in paternity duo cases, the alleged father (AF) and the child need to be opposite homozygous in order to yield a genetic exclusion. STR loci on the other hand have larger number of alleles but also a higher mutation rate, increasing the uncertainty of whether a genetic inconsistency really is a true exclusion or in fact a mutation. There are studies promoting the use of biallelic markers as supplementary markers in relationships testing [5,6] while other, more recent studies, have concluded that smaller sets of biallelic markers should be used with caution, especially when relatives might be involved [7,8].

When it comes to paternity investigations, the vast majority of such cases end up with a well-founded conclusion: If  $H_1$  and  $H_2$  are the two competing hypotheses, with  $H_1$  representing paternity, the likelihood ratio  $L(D) = Pr(D|H_1)/Pr(D|H_2)$  in favor of  $H_1$  is computed, with D representing the test data. If  $L(D) > L_H$  for some cutoff value, for example  $L_H = 10,000$ , paternity is declared, whereas if there are numerous genetic inconsistencies, non-paternity is declared [9]. Different laboratories may use different rules to declare non-paternity. In this paper, we will assume that computational models that allow mutations and silent alleles are used, and that the rule for declaring non-paternity is in fact formulated as  $L(D) < L_L$  for some  $L_L$ . For inconclusive cases where  $L_L < L(D) < L_H$ , a number of different supplementary markers can be typed in order to reach a definite conclusion.

To investigate the usefulness of acquiring data D, or additional data  $D_1$  or  $D_2$  if the initial test is inconclusive, a central issue is the error rates for the data. Assuming that one of the considered hypotheses is true, what is the probability that the likelihood ratio will lead to an erroneous conclusion, when compared to the cutoff values. Specifically, we need to consider the functions

 $E_1(\ell) = \Pr(L(D) < \ell | H_1)$ 

and

$$E_2(\ell) = \Pr(L(D) > \ell | H_2)$$

for various types of data. We will see how these functions can be approximated using simulation for several specific data types and hypotheses  $H_1$  and  $H_2$ .

Decision theory is a general way to aid decisions in situations of uncertainty, by specifying costs of various possible outcomes from decisions, and then selecting the decision with the lowest expected cost [10]. Application of decision theory is often hampered by the difficulty in assigning costs to outcomes. In our case, it is not obvious how to assign a monetary cost to falsely concluding with paternity when there is no paternity, or to the opposite type of error. However, when laboratories decide on cutoff values  $L_H$  and  $L_D$ , they are implicitly making decisions about the relative costs of various errors. We show how we can use these cutoff values computed from initial data analyses to obtain indirect cost estimates whose numerical values can be used as part of guides for decisions.

The data used in the case examples were based on simulations. Simulation of families and their DNA profiles gives the opportunity to rather simply investigate different issues and also test the impact of model change. Both mutations and silent alleles were modeled and accounted for in our simulations.

#### 2. Material and methods

#### 2.1. Decision theory

Assume two competing hypotheses  $H_1$  and  $H_2$  have probabilities  $Pr(H_1)$  and  $Pr(H_2) = 1 - Pr(H_1)$ , and that a choice should be made between  $H_1$ ,  $H_2$ , or possibly making no decision. To facilitate a choice one may assign costs to various outcomes. Without loss of generality, we assume that making no decision has unit cost, so that all other costs are measured relative to this. If  $H_1$  is true and we decide on  $H_2$  we assume a cost  $1 + c_1$  is incurred  $(c_1 > -1)$ , while we assume a cost of  $1 + c_2$  in the opposite case ( $c_2 > -1$ ). The expected costs of deciding on  $H_1$ ,  $H_2$ , and making no choice are  $Pr(H_2)(1 + c_2)$ ,  $Pr(H_1)(1 + c_1)$ , and 1, respectively. If  $c_1c_2 \le 1$ , minimizing expected costs leads to choosing  $H_1$  if  $Pr(H_1) > (1 +$  $(c_2)/(2 + c_1 + c_2)$  and otherwise  $H_2$ . A more interesting case for us is when  $c_1c_2 > 1$ , which also implies that  $c_1 > 0$  and  $c_2 > 0$ . In this case one should choose  $H_1$  if  $Pr(H_2)(1 + c_2) < 1$ ,  $H_2$  if  $Pr(H_1)(1 + c_1) < 1$ , and otherwise one should make no decision. In terms of the odds ratio  $o = Pr(H_1)/(1 - Pr(H_1))$ , where  $Pr(H_1) = o/(o + 1)$  and  $Pr(H_2) = 1/(o+1)$ , the decision rules are as follows: If  $c_1c_2 \le 1$ , decide on  $H_1$  if  $o > (1 + c_2)/(1 + c_1)$ , otherwise on  $H_2$ . If  $c_1c_2 > 1$ , decide on  $H_1$  if  $o < c_2$ ,  $H_2$  if  $o < 1/c_1$ , and otherwise make no decision. Generally o will be the posterior odds based on data D. According to Bayes formula on odds form we have  $o = L(D)o_0$ , where L(D) is the likelihood ratio and  $o_0$  the prior odds.

In DNA testing laboratories, decisions are generally not made by first estimating costs  $c_1$  and  $c_2$ . Instead, one often uses fixed lower and upper bounds to which L(D) is compared: If L(D) is very high,  $H_1$  is declared true, if L(D) is very low,  $H_2$  is declared true, and in between one delays a decision. This paper focuses on helping laboratories choose additional data sets when L(D) gives an inconclusive result. That inconclusive results are possible means as we saw above that  $c_1c_2 > 1$  and that both  $c_1$  and  $c_2$  are positive.

When making a decision based on *D*, the theoretically most sound approach is to compare  $o = L(D)o_0$  to cutoff values  $L_H$  and  $L_L$ , and declare  $H_1$  true if  $o > L_H$ , declare  $H_2$  true if  $o < L_L$ , and otherwise declare the result inconclusive. In practice, however, laboratories for convenience often ignore the prior odds  $o_0$ , comparing L(D)directly with  $L_H$  and  $L_L$  to make the decision. Let us define  $L_H^* = L_H$ for the first type of decision making, and  $L_H^* = o_0 L_H$  for the second type, and similar for  $L_L^*$ . Then in all cases,  $H_1$  is chosen if  $o > L_H^*$ ,  $H_2$  is chosen if  $o < L_{L_1}^*$ , and in between no decision is made. Comparing with the above, we see that  $c_1 = 1/L_L^*$  and  $c_2 = L_H^*$ , in other words, the decision routines of the laboratories implicitly correspond to estimating costs  $c_1$  and  $c_2$  at these values.

We now consider the situation where results using data D are inconclusive, i.e.,  $L_L^* < o < L_H^*$ , and we would like to optimally choose between producing additional data sets  $D_1, D_2, \ldots, D_N$ , or possibly produce none of these datasets. We start by defining for  $i = 1, \ldots, N$  and any  $\ell > 0$  functions

$$E_{i1}(\ell) = Pr(L(D_i) < \ell | H_1)$$

$$E_{i2}(\ell) = Pr(L(D_i) > \ell | H_2)$$

As these cumulative probability distribution functions measure the probability to make "errors" in the sense of making a wrong conclusion, we will refer to them as error rates. As we assume the different data sets are independent given the hypotheses, the decision after  $D_i$  has been acquired should be based on comparing Download English Version:

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