

Contents lists available at ScienceDirect

Chemical Engineering Journal

journal homepage: www.elsevier.com/locate/cej



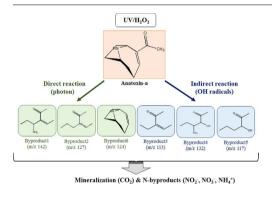
CrossMark

Degradation mechanism of anatoxin-a in UV-C/H₂O₂ reaction

So-Yeon Tak, Moon-Kyung Kim, Jung-Eun Lee, Young-Min Lee, Kyung-Duk Zoh*

Department of Environmental Health Sciences, School of Public Health, Seoul National University, 1 Gwanak-ro, Gwanak-gu, Seoul, Republic of Korea

G R A P H I C A L A B S T R A C T



A R T I C L E I N F O

Keywords: UV-C/H₂O₂ UV photolysis By-products Pathway Acetate Mineralization

ABSTRACT

In this study, the kinetics and removal mechanism of anatoxin-a ($C_{10}H_{15}NO$) during a UV-C/H₂O₂ reaction were investigated. The removal of anatoxin-a was more effective during a UV-C/H₂O₂ reaction than with either UV photolysis or H₂O₂ alone, due to the effective production of hydroxyl (OH) radicals. The UV-C/H₂O₂ reaction of anatoxin-a resulted in an approximately 60% decrease in total organic carbon (TOC) within 420 min, while 45% of the carbon in anatoxin-a was converted into acetate, and most of the nitrogen in anatoxin-a was converted into NH₄⁺, NO₂⁻, and NO₃⁻ ions. More than 50% of the nitrogen in anatoxin-a was mineralized, mainly into NO₃⁻ ions, and complete nitrogen recovery was achieved after 120 min of the UV-C/H₂O₂ reaction. Using liquid chromatography-electrospray ionization-tandem mass spectrometry (LC-ESI-MS/MS), we identified six degradation by-products in the UV-C/H₂O₂ reaction ($[M+H]^+ = 142, 127, 113, 132, 117, and 124, respectively$), which were further degraded as the reaction continued. Using these by-products, we proposed a degradation pathway for anatoxin-a during the UV-C/H₂O₂ reaction. Our results indicate that anatoxin-a can be effectively removed by a UV-C/H₂O₂ reaction during water treatment processes.

1. Introduction

Cyanobacteria are prokaryotic photoautotrophs that secrete various secondary metabolites. Certain cyanobacteria produce cyanotoxins including hepatotoxins (microcystins, nodularins, and cylindrospermopsin), neurotoxins (anatoxin-a and saxitoxin), and dermatotoxins [1,2]. There is much concern regarding the secondary

metabolites of cyanobacteria, because it has been reported that they are harmful to humans, fish, and animals, despite the low range of concentrations at which they are typically present in the environment [3,4]. Through the ingestion of recreational water, untreated drinking water, and contaminated fish, humans can be exposed to cyanotoxins [5,6].

Anatoxin-a (C10H15NO) is a neurotoxin secreted mainly by

* Corresponding author.

E-mail address: zohkd@snu.ac.kr (K.-D. Zoh).

http://dx.doi.org/10.1016/j.cej.2017.10.081

Received 23 April 2017; Received in revised form 7 October 2017; Accepted 16 October 2017 Available online 17 October 2017 1385-8947/ © 2017 Elsevier B.V. All rights reserved.

Anabaena flos-aqueae, Aphanizomenon, and Microcystis [7]. It is an alkaloid neurotoxin, and is not only a tumor promoter but also a tumor initiator. Anatoxin-a can mimic acetylcholine by its amine and carbonyl functionality binding to the acetylcholine receptors [2,8,9]. In the environment, anatoxin-a has been detected at levels of 120 µg/L in lake water in Poland [10], 5.2–35.7 µg/L in fresh water, and 0.71–10.18 µg in plants in the United States [11]. Concentrations of anatoxin-a in water and algae materials in Korea reportedly range from 0.01–0.08 µg/ L to 0.61–8.68 µg/g (dry wt), respectively [12].

During drinking water treatment processes, intracellular cyanotoxins such as anatoxin-a can also be released through the degradation of cyanobacteria [13]. Therefore, these residual cyanotoxins in water must be removed during water treatment processes. Cyanobacteria and intracellular cyanotoxins can be removed by conventional water treatment processes such as coagulation, flocculation, sedimentation, and sand filtration. However, extracellular cyanotoxins in the dissolved phase cannot be degraded effectively by such processes [14].

As an alternative, advanced oxidation processes (AOPs) can be applied for the removal of anatoxin-a in water. The degradation of anatoxin-a in water has been studied using ozone (O₃), hydrogen peroxide (H₂O₂), chlorine (Cl), chloroamine (NH₂Cl), and potassium permanganate (KMnO₄); while anatoxin-a is not effectively removed by chlorine and chloramine [15], ozone and KMnO₄ have proven effective in degrading the anatoxin-a molecule [9,15,16]. Al Momani [17] observed that the combinations of H₂O₂ or Fe(II) with ozone (O₃/H₂O₂ and O₃/ Fe(II)) were more efficient than O₃ or H₂O₂ oxidation alone. However, ozonation related process can produce toxic ozonation byproducts.

The UV related AOP processes were effective in degrading anatoxina. UV was able to decompose anatoxin-a by direct photolysis or indirect photolysis through an AOP process. UV/H₂O₂ reaction involves the production of hydroxyl (OH) radicals produced by the hemolytic cleavage of H₂O₂, with a wavelength shorter than 280 nm [18–20]. Compared with other AOPs, the UV/H₂O₂ reaction forms two OH radicals due to a single-step dissociation of H₂O₂. Mineralization of organic compounds can be achieved under appropriate conditions without phase transition problems [21].

Recent studies have examined the removal of anatoxin-a using vacuum-UV photolysis and UV/H₂O₂ [22], and UV-C LED and UV-C LED/ H₂O₂ reactions [23]. Afzal et al. [22] reported that low pressure UV lamp was not effective in degrading anatoxin-a through direct photolysis; however medium pressure UV lamp was able to degrade 88% of anatoxin-a. Use of H₂O₂ along with low pressure UV lamp resulted in above 70% degradation of anatoxin-a. Verma and Sillanpää [23] observed that 96% removal of anatoxin-a was achieved by photolysis using UV-C LED, and 79% removal was achieved with UV-C/H₂O₂ process in raw water sample. Verma et al. [24] also found that UV-C/ peroxylmonosulfate (PMS) were efficient in the degradation of anatoxin-a, even in a lake water. However, these UV related AOP processes and the UV/H₂O₂ reaction only observed degradation kinetics of anatoxin-a. Although anatoxin-a can be removed from water, the major concern is the formation of unknown by-products that can lead to the secondary contamination of drinking water. Until now, no studies have examined anatoxin-a degradation by-products or pathways during the UV/H₂O₂ reaction or any AOP process.

In this study, we investigated the kinetics and removal mechanism of anatoxin-a during a UV-C/H₂O₂ reaction, and compared it with UV photolysis and a H₂O₂ only reaction. To investigate the degradation mechanism of anatoxin-a during the UV-C/H₂O₂ reaction, we examined the mineralization of anatoxin-a by measuring various parameters such as total organic carbon (TOC), acetate, NH_4^+ , NO_2^- , and NO_3^- ions, and identified the organic by-products formed during the UV-C/H₂O₂ reaction using liquid chromatography-electrospray ionization-tandem mass spectrometry (LC-ESI-MS/MS) and ACD/MS Fragmenter software. Finally, using the by-products identified, we proposed a possible degradation pathway of anatoxin-a during the UV-C/H₂O₂ reaction.

2. Materials and methods

2.1. Chemicals

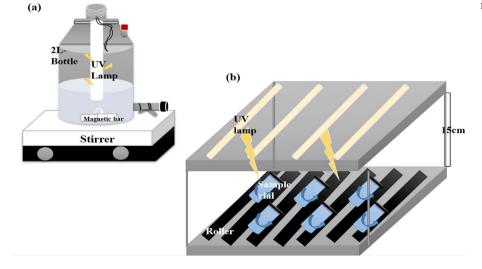
Anatoxin-a ($C_{10}H_{15}NO$) was purchased from Enzo Life Sciences (Farmingdale, NY, USA). Table S1 shows the physicochemical properties of anatoxin-a. A stock solution of anatoxin-a was prepared by dissolving 1 mg anatoxin-a in 1 mL Milli-Q water deionized water (Millipore, Billerica, MA, USA). A 10 mg/L stock solution was prepared by diluting with deionized water. The stock solution was kept in darkness at -20 °C to inhibit degradation.

Hydrogen peroxide (30%, v/v), formic acid (FA: 88%, v/v), and catalase (10,000-40,000 units/mg protein) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Acetonitrile and distilled water (J.T. Baker, Waltham, MA, USA) were used as mobile phase eluents for LC-ESI-MS/MS.

2.2. Experimental procedures

To investigate the rate of anatoxin-a removal, UV photolysis and UV-C/H₂O₂ reactions were performed using a UV photoreactor. A schematic diagram of the photolysis reactor system is shown in Fig. 1(a). The reactor consisted of a 2-L glass bottle, a stirrer, and a UV-C lamp (6 W, 254 nm, Sankyo Electronics Co., Ltd., Inazawa, Japan). UV intensity was measured with a radiometer (VLX-3W Radiometer 9811–50, Cole-Parmer, Vernon Hills, IL, USA). The single UV lamp had

Fig. 1. Schematic diagram of the photolytic reactors.



Download English Version:

https://daneshyari.com/en/article/6581079

Download Persian Version:

https://daneshyari.com/article/6581079

Daneshyari.com