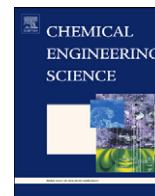




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## Genome-scale *in silico* modeling and analysis for designing synthetic terpenoid-producing microbial cell factories

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### HIGHLIGHTS

- ▶ Developed *in silico* rational framework for synthetic terpenoid production.
- ▶ Updated metabolic model of *A. thaliana* to describe terpenoid biosynthesis.
- ▶ Analyzed heterologous expression of *A. thaliana* pathways in microbial systems.
- ▶ Proposed novel *in silico* method of cofactor modification analysis.
- ▶ Improve terpenoid yield by modifying enzyme cofactor specificity.

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### ABSTRACT

Terpenoids are a large and diverse group of plant secondary metabolites with important applications in the pharmaceutical, cosmetic and food industries. However, low yields obtained from natural plant sources necessitate the search for alternative ways to increase the throughput of terpenoid production. Thus, fast-growing microbial systems, such as *Escherichia coli* and *Saccharomyces cerevisiae*, can be genetically engineered to achieve high productivity by systematically designing terpenoid synthetic routes and improving its precursor, isopentenyl diphosphate (IPP) production. To this end, we develop an *in silico* model-based rational framework where the cellular metabolism in natural terpenoid producer is analyzed to provide a basis for designing its synthetic pathways in microbial hosts. At the outset, we updated the genome-scale *Arabidopsis thaliana* metabolic model to characterize optimal metabolic utilization patterns that were subsequently incorporated into the *in silico* models of microbial hosts for improving terpenoid yield. We also developed a novel computational approach, cofactor modification analysis (CMA), to tackle potential limitations in terpenoid production caused by suboptimal balance of the different redox cofactors. The enzyme targets identified by CMA can potentially lead to better metabolic engineering strategies for enhancing terpenoid production in microbial systems.

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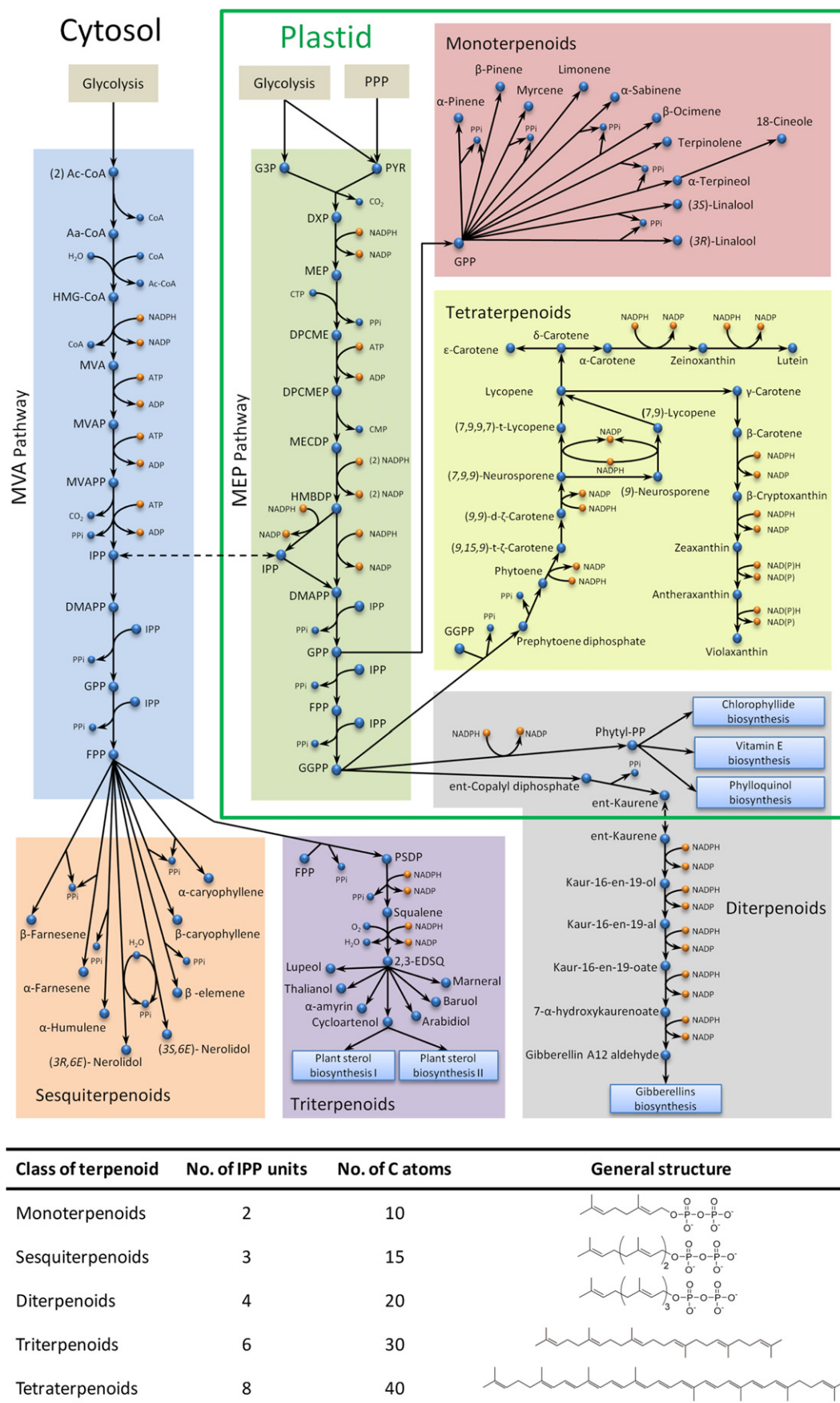
### 1. Introduction

Plant secondary metabolites, such as alkaloids, terpenoids and phenols, play an essential role in the defense against damage by herbivores (Buchanan et al., 2000; Keeling and Bohlmann, 2006). Among these secondary metabolites, terpenoids constitute the largest class of natural organic compounds with about 25,000

chemical structures being reported (Gershenzon and Dudareva, 2007). Such diverse terpenoids can be synthesized from five-carbon isoprene precursors, isopentenyl diphosphate (IPP) and its isomer dimethylallyl diphosphate (DMAPP) through a combination of biosynthetic pathways, thereby forming many multicyclic structures, comprising different carbon skeletons attached with various functional groups (Carretero-Paulet et al., 2002; Rodriguez-Concepcion and Boronat, 2002; Withers and Keasling, 2007). The terpenoids are classified according to the number of isoprene units (Fig. 1). The wide range of biological properties unique to each terpenoid species have been largely exploited for perfume manufacture and therapeutic applications to formulate

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**Fig. 1.** Terpenoid biosynthesis pathways. The biosynthesis of terpenoids occurs via linear pathways which branch out from the precursors formed by the MEP and/or MVA pathways. The pathway information is obtained from online database as described in Section 3.1. Metabolite abbreviations: Aa-CoA, acetoacetyl-CoA; Ac-CoA, acetyl-CoA; DMAPP, dimethylallyl diphosphate; DPCME, 4-diphosphocytidyl-2-C-methylerythritol; DPCMEP, 4-diphosphocytidyl-2-C-methyl-D-erythritol 2-phosphate; DXP, 1-deoxy-D-xylulose 5-phosphate; FPP, farnesyl diphosphate; G3P, glyceraldehyde-3-phosphate; GGPP, geranylgeranyl diphosphate; GPP, geranyl diphosphate; HMBDP, (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate; HMG-CoA, hydroxymethylglutaryl-CoA; IPP, isopentenyl diphosphate; MECDP, 2-C-methyl-D-erythritol 2,4-cyclopyrophosphate; MEP, 2-C-methyl-D-erythritol 4-phosphate; MVA, mevalonic acid; MVAP, mevalonate-5-phosphate; MVAPP, mevalonate-5-diphosphate; and PYR, pyruvate.

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