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Simple immobilization of mercury ion chemosensors to solid substrate

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ABSTRACT

To develop selective and disposable mercury chemosensors, four different mercury chelators were synthesized and grafted onto cellulose filter paper. Three colorimetric sensors were azobenzene derivatives bearing a dithiaazadioxocrown ether moiety as a core chelator for Hg^{2+} ions. The fourth chelator was a rhodamine-based *turn-on* mercury binder that responded to both methylmercury and Hg^{2+} ions. This sensor develops the typical pink color of rhodamine (colorimetric) and exhibits strong fluorescence (fluorometric). For paper immobilization, the cellulose filter paper was first primed with 3-(aminopropyl)trimethoxysilane (APTMS) so that all the four ligands were covalently immobilized, simply by soaking them in each solution.

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Introduction

Because of the high toxicity of mercury ions (Hg^{2+}), sensitive detection methods have been sought to prevent human exposure, which causes health problems affecting the cardiovascular and central nervous systems [1]. Considering the low limit for mercury in drinking water specified by Environmental Protection Agency (EPA) regulations (i.e., 2 ppb), very sensitive detection schemes, such as electrochemical voltammetry (or amperometry) [2,3], atomic absorption spectroscopy (AAS) [4], and inductively coupled plasma mass spectroscopy (ICP-MS) [5], have been developed in recent decades. These methods usually require sophisticated and expensive instrumentation and specially trained operators. The use of mercury has been extensively banned from many industrial processes, and related regulations have been tightened in most countries. However, the accumulated concentrations in marine food chains and the relatively loose regulations in developing countries still necessitate simple and cost-effective monitoring systems for mercury ions. Therefore, metal-chelating chemosensors [6,7] that change their fluorescent characteristics or exhibit unique

colors upon binding with metal ions are promising alternatives to expensive instrumentation. In a previous study, we synthesized an azobenzene-based colorimetric sensor with a dithiaazadioxocrown ether as a specific binder of Hg^{2+} ions that showed selective chelating capability and typical solvatochromic behavior in aqueous and organic solvents [8]. Here, as a practical extension of this previous investigation, mercury-binding ligands were tethered to cellulose paper to create a disposable mercury sensor based on a concept similar to that of ordinary pH paper. In addition to the dithiaazadioxocrown ether, a rhodamine-based dye with a thiosemicarbazide group [9] was newly synthesized and tethered, and its performance was compared to that of the azobenzene-based chelators.

Experimental

Instruments

1H nuclear magnetic resonance (NMR) and ^{13}C NMR spectra were collected for the structural analysis of the ligands using Varian Inova 600 NB spectrometers (USA). Mass spectra were obtained using a mass spectrometer coupling electrospray ionization and a time-of-flight analyzer (ESI-MS) from Agilent Technologies Inc. 1100 + G1958 (USA). Ultraviolet/visible (UV/Vis) and fluorescence spectra were collected using a PerkinElmer VT 3000 spectrophotometer (USA) and a Scinco Fluoromate FS-2 spectrometer (South Korea), respectively.

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Materials

2,4-Dinitro-5-fluoroaniline (DNFA) (Japan), 4,5-difluoro-2-nitroaniline (DFNA) (Japan), *N*-(3-dimethylaminopropyl)-*N*-ethylcarbodiimide hydrochloride (EDC-HCl) (China) and triethylamine (TEA) (Japan) were purchased from TCI. Sodium nitrite (Japan), rhodamine B (India), hydrazine hydrate (South Korea), 1,4-phenylene diisothiocyanate (Poland), *N*-hydroxysuccinimide (NHS) (USA), (3-aminopropyl)trimethoxysilane (APTMS) (USA), *N,N'*-dimethylformamide (DMF) (Germany), and all metal perchlorates (USA) were purchased from Sigma-Aldrich and used without further purification. Qualitative cellulose filter paper was purchased from Whatman (No. 2) (China), and solvents for NMR experiments, including chloroform-*d* (USA) and dimethylsulfoxide-*d*₆ (USA), were purchased from Cambridge Isotope Laboratories.

10-(4-((4,5-Difluoro-2-nitrophenyl)diazanyl)phenyl)-1,4-dioxo-7,13-dithia-10-azacyclo-pentadecane (**L2**) and 10-(4-((5-fluoro-2,4-dinitrophenyl)diazanyl)phenyl)-1,4-dioxo-7,13-dithia-10-azacyclopentadecane (**L3**)

Sodium nitrite (69.0 mg, 1.0 mmol) was dissolved in ice-cold water at 0 °C (10 mL) and added dropwise into another ice-cooled 40-mL solution of DFNA (174.1 mg, 1.0 mmol) or DNFA (201.0 mg, 1.0 mmol) in a mixture of H₂SO₄/acetic acid (1/3, v/v). The mixture was stirred for 20 min. The diazonium salt solution was added dropwise to a DMF solution (50 mL) of 1,4-dioxo-7,13-dithia-10-phenyl-10-azacyclopentadecane (327 mg, 1.0 mmol) [8]. After additional stirring at 0 °C for 6 h, 20 mL of CHCl₃ and 10 mL of water were sequentially added to the reaction vessel, and the solution was transferred to a separatory funnel. The organic layer was extracted from the aqueous layer three times, and the organic extracts were combined, dried over anhydrous MgSO₄, and filtered. After evaporating the solvent *in vacuo*, the crude product was chromatographed on silica gel using a CHCl₃/MeOH mixture (20/1, v/v) to yield **L2** as an orange solid (353.8 mg, 69.1% yield) and **L3** as a purple solid (311.5 mg, 57.8% yield). **L2**: ¹H NMR (CDCl₃, 600 MHz): δ 7.85 (d, *J* = 9.2 Hz, 1H), 7.75 (dd, *J* = 9.0, 7.1 Hz, 1H), 7.63 (dd, *J* = 10.7, 7.6 Hz, 1H), 6.70 (d, *J* = 9.3 Hz, 1H), 3.83–3.80 (m, 1H), 3.78–3.75 (m, 1H), 3.66 (s, *J* = 4.2 Hz, 1H), 2.96–2.92 (m, 1H), 2.79–2.77 (m, 1H); ¹³C NMR (CDCl₃, 600 MHz): δ 150.98, 143.58, 143.39, 142.70, 129.46, 126.87, 113.52, 111.61, 111.57, 106.92, 106.72, 74.35, 74.29, 70.70, 70.66, 70.37, 52.21, 52.17, 51.75, 31.50, 31.45, 29.54, 29.49; MS (ESI) *m/z* calcd for C₂₂H₂₆F₂N₄O₄S₂ [M + H]⁺ 513.14, found 513.0. **L3**: ¹H NMR (CDCl₃, 600 MHz): δ 8.65 (d, *J* = 6.8 Hz, 1H), 7.89 (d, *J* = 9.3 Hz, 3H), 7.66 (d, *J* = 11.5 Hz, 2H), 6.73 (d, *J* = 9.4 Hz, 4H), 3.84–3.79 (m, 16H), 3.66 (s, *J* = 5.6 Hz, 8H), 2.97–2.91 (m, 8H), 2.81–2.76 (m, 8H); ¹³C NMR (CDCl₃, 600 MHz): δ 160.03, 152.46, 144.23, 127.38, 126.31, 123.07, 111.98, 110.39, 107.67, 107.51, 105.74, 105.57, 74.21, 70.60, 52.33, 31.60, 29.55; MS (ESI) *m/z* calcd for C₂₂H₂₆FN₅O₆S₂ [M + H]⁺ 540.13, found 540.1.

1-(3',6'-Bis(diethylamino)-3-oxospiro[isindoline-1,9'-xanthen]-2-yl)-3-(4-isothiocyanatophenyl)thiourea (**L4**)

Rhodamine B hydrazide **1** (457.2 mg, 1.0 mmol) [11,12] was dissolved in 50 mL of DMF, and then 1,4-phenylene diisothiocyanate (959.9 g, 5.0 mmol) was added. After stirring overnight at room temperature (rt), the solvent was evaporated *in vacuo*. The crude residue was chromatographed on silica gel using a CHCl₃/MeOH mixture (50/1, v/v) to yield **L4** as a white solid (588.3 mg, 91% yield). ¹H NMR (CDCl₃, 600 MHz): δ 7.98 (d, *J* = 7.6 Hz, 1H), 7.65 (td, *J* = 7.6, 1.2 Hz, 1H), 6.60–7.56 (m, 1H), 7.56 (s, 1H), 7.27 (d, *J* = 7.7 Hz, 1H), 7.12–7.06 (m, 2H), 7.04–6.99 (m, 2H), 6.97 (s, *J* = 6.0 Hz, 1H), 6.47 (d, *J* = 8.8 Hz, 2H), 6.42 (d, *J* = 2.6 Hz, 2H), 6.27 (dd, *J* = 8.9, 2.6 Hz, 2H), 3.44–3.21 (m, 10H), 1.15 (t, *J* = 7.1 Hz, 16H); ¹³C NMR (CDCl₃, 600 MHz): δ 182.46, 167.27, 154.30, 149.98, 149.34,

136.79, 134.38, 129.12, 128.99, 128.28, 127.57, 125.72, 125.52, 124.77, 123.89, 108.30, 104.07, 98.29, 67.36, 44.40, 12.56; MS (ESI) *m/z* calcd for C₃₆H₃₆N₆O₂S₂ [M + H]⁺ 649.23, found 649.2.

Immobilization of mercury binder molecules (**L1–L4**) onto amine-modified cellulose paper

Cellulose filter paper with a diameter of 90 mm was soaked in an ethanolic APTMS solution (1 wt%) for 1 h. The amine-primed cellulose paper was washed with copious ethanol three times and dried under vacuum at 40 °C for 6 h. To tether **L1** onto the amine-modified cellulose paper, **L1** was activated with the NHS ester in advance by dissolving **L1** (47.5 mg, 0.1 mmol), EDC-HCl (22.9 mg, 0.12 mmol) and NHS (13.8 mg, 0.12 mmol) in DMF (20 mL) [8]. After stirring at rt for 3 h, TEA (30.3 mg, 0.3 mmol) was added to the reaction mixture, and the amine-modified cellulose paper was soaked in it overnight. After washing with copious ethanol three times and drying under vacuum at 40 °C for 6 h, **L1** was immobilized on amine-modified cellulose paper (**L1-AP**). For the immobilization of **L2** and **L3**, a piece of amine-modified cellulose paper was simply dipped into the above-described DMF solution (0.1 mmol in 20 mL) containing TEA (0.3 mmol) overnight. After incubation at rt, the cellulose paper was washed with copious ethanol three times and dried for 6 h *in vacuo* at 40 °C to yield the corresponding **L2-AP** and **L3-AP**. We were unable to tether **L2** under these conditions based on the color of the resultant **L2-AP** compared to that of **L3-AP**, which exhibited a color characteristic of **L3**. For **L4** immobilization, a piece of amine-modified cellulose paper was simply dipped into a DMF solution of **L4** (0.1 mmol in 20 mL) overnight (without TEA). The resultant **L4-AP** was washed with copious ethanol three times and dried *in vacuo* at 40 °C for 6 h.

Results and discussion

Synthesis of **L1**, **L2**, and **L3** and the estimation of the selective Hg²⁺-binding capability

In addition to the previous azobenzene-based chelator (compound **L1** in Fig. 1(a)) that bears a carboxylate group in the *para*-position [8], we also synthesized mercury chelators (**L2** and **L3**) with backbones similar to that of **L1** but different terminal functionalities, as shown in Fig. 1(a). The introduction of electron-withdrawing nitro- or fluoro-groups was intended to tune the spectral characteristics by modulating the internal charge-transfer (ICT) dipoles [11]. Specifically, two fluoronitroaniline compounds—DFNA and DNFA—were diazotized in acidic conditions and coupled to a dithiaazadioxo crown ether moiety. Both fluoronitroanilines increase the electron withdrawal capability, yielding a fortified *push–pull* dipole for the azobenzene [12]. The diazotization reactions were moderately successful, yielding **L2** (~69% yield) and **L3** (~58% yield) after silica gel column chromatography. Because of the nitro- (or fluoro-) group in the azobenzene moiety, which increases the ICT dipole moment, the maximum absorption wavelengths of **L2** (468 nm) and **L3** (533 nm) were red-shifted compared to that of **L1** (Δ*λ*_{max}: **L2** ~30 nm, **L3** ~95 nm), yielding deep yellow and magenta colors for **L2** and **L3**, respectively. The binding behaviors of **L2** and **L3** with Hg²⁺ were considered to be the same as that of **L1** in a mixed solvent of acetonitrile and water, and the coordination of a single Hg²⁺ ion in the dithiaazadioxo ring of **L1** was established by NMR titration [8]. Notably, the newly synthesized **L2** and **L3** have additional advantages for solid immobilization because the highly negative aromatic fluoride may be prone to aromatic nucleophilic substitution by suitable nucleophiles, such as amines or alkoxides (see below) [14].

As shown in Fig. 2(b), the colorimetric changes of **L2** in response to various metal ions were first examined by dissolving it in solutions containing excess metal (5 eq.) in a mixed solvent of

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