



## Effect of anoxic/aerobic phase fraction on N<sub>2</sub>O emission in a sequencing batch reactor under low temperature

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### ABSTRACT

Laboratory scale anoxic/aerobic sequencing batch reactor (A/O SBR) was operated around 15 °C to evaluate the effect of anoxic/aerobic phase fraction (PF) on N<sub>2</sub>O emission. The ammonia removal exhibited a decrease trend with the increase of PF, while the highest total nitrogen removal was achieved at PF = 0.5. Almost all the N<sub>2</sub>O was emitted during the aerobic phase, despite of the PF value. However, the net emission of N<sub>2</sub>O was affected by PF. Under the premise of completely aerobic nitrification, the lowest N<sub>2</sub>O emission was achieved at PF = 0.5, with a N<sub>2</sub>O–N conversion rate of 9.8%. At lower PF (PF = 0.2), N<sub>2</sub>O emission was stimulated by residual nitrite caused by uncompleted denitrification during the anoxic phase. On the other hand, the exhaustion of the easily degradable carbon was the major cause for the high N<sub>2</sub>O emission at higher PF (PF = 0.5). The N<sub>2</sub>O emission increased with the decreasing temperature. The time-weighted N<sub>2</sub>O emission quantity at 15 °C was 2.9 times higher than that at 25 °C.

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### 1. Introduction

The general degradation of water quality in rivers and lakes has led to more stringent effluent quality standards, in order to prevent eutrophication by reducing nutrient level in wastewater discharged into local waterways. Many modifications and novel processes have been developed and implemented for nitrogen removal from wastewater (Tchobanoglous et al., 2003).

In recent years, sequencing batch reactor (SBR) system has attracted a great deal of interest because SBR can carry out biological nitrogen removal in a single reactor by maintaining anoxic and aerobic stages sequentially (Andreottola et al., 2001; Wilderer et al., 2001). Anoxic/aerobic activated sludge process is one of the most widely used biological nitrogen processes. The simultaneous removal of organic and nitrogen can be accomplished through scheduling of the anoxic and aerobic phase and the allocation of reaction time can be changed by varying the phase lengths (Hong et al., 2008). The pre-denitrification phase can degrade the influent organic anaerobically, reduced the oxygen demanded in the following aerobic phase. More importantly, the alkalinity produced during the denitrification phase provide an optimum circumstance for the subsequential aerobic nitrification process, thus reduced the operation cost (Muller et al., 2003).

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It is generally accepted that the biological treatment process of domestic wastewater occupies an extremely important position among the many sources of nitrous oxide (N<sub>2</sub>O) (Peter et al., 1995; Kong et al., 2002). N<sub>2</sub>O is an important greenhouse gas. It can cause the greenhouse effect and ozone depletion in the stratosphere. Its 100-year global warming potential is 298 times higher than that of carbon dioxide (CO<sub>2</sub>) and it has a lifetime of 114 years (IPCC, 2007). The concentration of atmospheric N<sub>2</sub>O is estimated to be approximately 314 ppbv, which is approximately 16% higher than that during the preindustrial era, and it continues to increase at a rate of 0.25%/year (Kishida et al., 2004). It is therefore of great importance to develop technologies that can suppress N<sub>2</sub>O emission from wastewater treatment processes.

N<sub>2</sub>O has been shown to be produced during both nitrification and denitrification in activated sludge and released to the atmosphere (Zheng et al., 1994; Tallec et al., 2008). Nitrification is the oxidation of NH<sub>4</sub><sup>+</sup> to NO<sub>3</sub><sup>-</sup> via NO<sub>2</sub><sup>-</sup>, which is achieved by two steps. Ammonia is firstly oxidized to nitrite by ammonia-oxidizing bacteria (AOB) and then converted to nitrate by nitrite-oxidizing bacteria (NOB) (Bock et al., 1986). N<sub>2</sub>O is produced as one of the by-products during nitrification and there are two possible mechanisms of N<sub>2</sub>O production (Prosser, 1989). Certain nitrifying bacteria generate N<sub>2</sub>O from the reduction of NO<sub>2</sub><sup>-</sup> under oxygen limiting conditions. Alternatively, N<sub>2</sub>O can be produced by chemical decomposition of NO<sub>2</sub><sup>-</sup> or various reactions of the intermediates formed during NH<sub>4</sub><sup>+</sup> oxidation (Ritchie and Nicholas, 1972).

On the other hand,  $N_2O$  is one of the obligatory intermediates in denitrification. Through denitrification,  $NO_3^-$ -N is reduced to  $NO_2^-$ -N by nitrate reductase, which is then reduced to  $N_2O$  by nitrite reductase.  $N_2O$  is finally reduced to  $N_2$  gas by  $N_2O$  reductase (Kimochi et al., 1998). Note that denitrifier can use  $NO_2^-$  or  $NO_3^-$  produced during nitrification. This coupling of nitrification and denitrification can occur when the conditions are favorable for both processes, especially in aggregates such as activated sludge (Zeng et al., 2003).

It has been reported that low DO concentration, low C/N ratio, low pH, short SRT or high nitrite concentration can result in  $N_2O$  accumulation during the biological nitrogen removal process (Zheng et al., 1994; Gejlsbjerg et al., 1998; Kampschreur et al., 2009). However, few studies have been reported on the effect of the anoxic/aerobic phase fraction (PF) on  $N_2O$  emission. In addition, most of the studies are preformed under the optimum temperature for nitrifier and denitrifier, around 25 °C, while many wastewater treatment plants are operated below this value for most of the year.

The aim of this paper is to evaluate the effect of anoxic/aerobic phase fraction on  $N_2O$  emission in biological nitrogen removal process under low temperature. For this purpose, a lab scale anoxic/aerobic sequencing batch reactor (A/O SBR) was operated under  $15 \pm 1$  °C. Experiments were conducted at different phase fraction after proper acclimation. In addition, the effect of temperature on  $N_2O$  emission was investigated after the optimum phase fraction was selected.

## 2. Methods

### 2.1. Synthetic wastewater

The synthetic wastewater with COD concentration of 300 mg/L and  $NH_4^+$ -N concentration of 60 mg/L was comprised of (per liter): 195 mg  $C_6H_{12}O_6$ ; 195 mg  $CH_3COONa \cdot 3H_2O$ ; 230 mg  $NH_4Cl$ ; 200 mg  $NaHCO_3$ ; 11 mg  $KH_2PO_4$ ; 18 mg  $K_2HPO_4 \cdot 3H_2O$ ; 10 mg  $MgSO_4 \cdot 7H_2O$ ; 10 mg  $FeSO_4 \cdot 7H_2O$ ; 10 mg  $CaCl_2 \cdot 2H_2O$ ; and 1 mL nutrient solution. One liter of nutrient solution contained: 0.15 g  $H_3BO_3$ ; 0.03 g  $CuSO_4 \cdot 5H_2O$ ; 0.18 g KI; 0.12 g  $MnCl_2 \cdot 4H_2O$ ; 0.06 g  $Na_2MoO_4 \cdot 2H_2O$ ; 0.12 g  $ZnSO_4 \cdot 7H_2O$ ; 0.15 g  $CoCl_2 \cdot 6H_2O$ ; and 10 g ethylene diamine tetraacetic acid (EDTA) (Zeng et al., 2003).

### 2.2. Reactor setup and operation

All experiments were conducted in a gastight sequencing batch reactor, constructed using a transparent, rigid plexiglas cylinder, with an effective volume of 24 L. A peristaltic pump was used to feed the SBR with the synthetic wastewater from the water tank during the feeding phase. An electric agitator with a rectangular paddle, used to keep the suspension of the sludge during the anoxic phase, was installed over the reactor. The air needed during the aerobic phase was supplied by an air pump through a porous stone diffuser located at the bottoms of the reactor. A relatively low aeration rate, which was  $7.5 L_{air}/(L_{reactor} h)$ , was used throughout the aerobic phase to mimic the actual wastewater treatment plant and achieve high nitrogen removal (Smith and Evans, 1982; Pochana and Keller, 1999). The off-gases were collected into gas sampling bags at intervals of 0.5 h. At the same time, liquid phase samples were taken to measure the water quality. Fig. 1 shows the schematic diagram of the experimental system.

The SBR was inoculated with microorganism from the Second Wastewater Treatment Plant of Everbright Water (Jinan) Ltd. (Jinan, China). During the whole experiments, the temperature was maintained at  $15 \pm 1$  °C. Each cycle of SBR consisted of five steps: 10 min feeding,  $x$  h anoxic reaction,  $(6 - x)$  h aerobic reac-

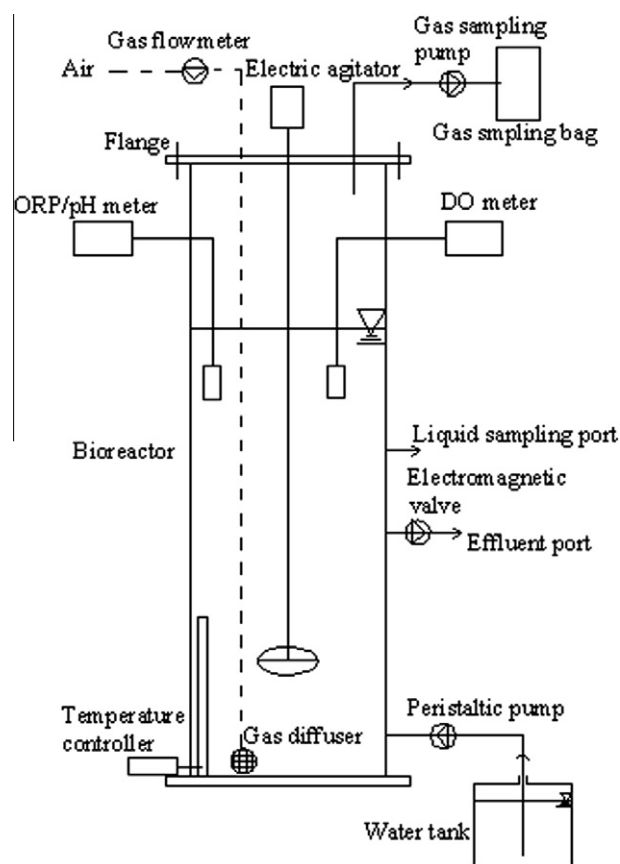


Fig. 1. Schematic description of the experiment system.

tion, 40 min setting, and 10 min decanting. The value of  $x$  depends on the PF investigated. Five different PF were investigated in this study, as listed in Table 1. The mixed liquor suspended solid (MLSS) was maintained at approximately 4000 mg/L and certain amount of excess sludge was disposed at the end of aerobic phase to control the SRT at approximately 20 d.

### 2.3. Analytical methods

COD,  $NH_4^+$ -N,  $NO_3^-$ -N,  $NO_2^-$ -N, TN, TP and MLSS were measured according to the standard methods (APHA, 1998). Dissolved oxygen was measured with a DO meter (HQ30d53LDO™, USA) and pH was measured with a pH meter (PHS-3C, China), every 10 min. The  $N_2O$  concentration was determined by a gas chromatography (SP-3410, China) with an electron capture detector (ECD) and a Poropak Q column, using 30 mL/min high-purity nitrogen as the carrier gas. The temperature of the detector and oven were set at 360 °C and 50 °C, respectively (Wu et al., 2009).

### 2.4. Calculation of $N_2O$ emission rate and emission quantity

During the anoxic phase, the whole reactor was gas tight, and the  $N_2O$  emission rate was calculated through the increase of

Table 1  
The operation conditions of SBR.

Scheduling	I	II	III	IV	V
PF	0.5	0.2	1	2	5
$x$ (h)	2	1	3	4	5

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