

Drug design for mPGES-1 from traditional Chinese medicine database: A screening, docking, QSAR, molecular dynamics, and pharmacophore mapping study

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ABSTRACT

To search for new anti-inflammatory that can replace the current COX-1 and COX-2 inhibitors, virtual screening by molecular docking of traditional Chinese medicine (TCM) molecules into microsomal prostaglandin E₂ synthase (mPGES-1) glutathione binding site was performed. To compare the top ranking derivatives with other mPGES-1 inhibitors, we constructed QSAR models using comparative molecular force field analysis (CoMFA) and comparative molecular similarity indices analysis (CoMSIA). The CoMFA model had a non-cross-validated coefficient (r^2) and a cross-validated coefficient (q^2) of 0.960 and 0.597. The r^2 and q^2 for CoMSIA (S + H + D) was 0.931 and 0.719, respectively. The top three TCM derivatives all can map into the respective steric, hydrophobic and hydrogen bond donor force fields. The top ranking TCM molecules were taken for *de novo* design; the top three *de novo* products were further analyzed using molecular dynamics simulation and qualitative structure–activity relationship (QSAR) model. Derivative, 2-O-caffeoyl tartaric acid-Evo_2, glucogallin-Evo_1 and 4-O-feruloylquinic acid-Evo_7, all had conserved hydrogen bond networks to key residues Arg38 and Arg70 during the 20 ns molecular dynamics simulation. In addition, all derivative–protein complexes had total energy lower the control–protein complex. Combining the results from molecular dynamics simulation and CoMFA/CoMSIA, we suggest 2-O-caffeoyl tartaric acid-Evo_2, glucogallin-Evo_1 and 4-O-feruloylquinic acid-Evo_7 as potent mPGES-1 inhibitors.

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1. Introduction

Prostaglandins, products of prostanoids synthesis, are auto-crines that are produced in various parts of human body. Prostaglandin E₂ (PGE₂), the most abundant subtype of prostaglandins, is the product from the action of prostaglandin E synthases on prostaglandin H₂ (Hara *et al.*, 2010). To date, there are three known classes of prostaglandin E synthases, namely cytosolic PGE synthase (cPGES) and two membrane-bound synthases,

mPGES-1 and mPGES-2 (Hara *et al.*, 2010). The mPGES-1 protein, unlike the other PGE synthases, is not constitutively expressed and can be induced in response to inflammatory stimuli.

In the past, non-steroidal anti-inflammatory drugs (NSAIDs) have been designed to target COX-1 and COX-2, the upstream enzyme for producing prostaglandin H₂. However, long-term suppression of prostanoid biosynthesis by using COX-1 and COX-2 inhibitors can have severe side effects, including gastrointestinal injury and renal irritation (Koeberle and Werz, 2009). Current evidences suggest that suppression of mPGES-1 activity can be considered as an alternative anti-inflammatory approach. Since mPGES-1 is functionally coupled to COX-2 and is also being responsible for excessive PGE₂, its inhibitions are, therefore, related to inflammation, pain, fever, atherosclerosis and tumorigenesis.

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To develop novel anti-inflammatory agents, TCM Database@-Taiwan (<http://tcm.cmu.edu.tw>), the current world largest small molecule database on traditional Chinese medicine, was employed in docking. Natural compounds isolated from TCM are gaining attention in the last few, and studies have been done already on compounds, such as emodin or gallic acid and many more, to investigate their potential effects to cellular process (Chen, 2009d; Chen et al., 2010b; Lin et al., 2010; Liu et al., 2010; Lo et al., 2010). In addition to screening of TCM database, we used both structure-based and ligand-based approaches to further validate and support the bioactivity of the lead candidates. Both strategies have been implemented in drug researches in the past, and we have successfully used both techniques in designing anti-viral, anti-inflammatory, and anti-tumor agents (Chang et al., 2010; Chen, 2008, 2009a,b,c,e,f, 2010a,b; Chen and Chen, 2007, 2010; Chen et al., 2009a,b, 2010a; Huang et al., 2010a,b,c).

2. Materials and methods

2.1. Docking

A total of 20,000 traditional Chinese medicine compounds were downloaded from TCM Database@Taiwan (<http://tcm.cmu.edu.tw>) and docked into the glutathione binding site of mPGES-1. All the ligands were pre-treated with force field of CHARMM, and all the missing hydrogen were added. The protein model used for docking was downloaded from Protein Data Bank (PDB: 3DWW (Jegerschold et al., 2008)). The nature substrate for mPGES-1, glutathione (γ -L-glutamyl-L-cysteinyl-glycine), which co-crystallized with mPGES-1 by electron crystallography, was used as the control molecule. The binding location of glutathione found in the protein crystal was set as the docking site.

Table 1
TCM docking results. Only the top 10 candidates and controls are shown.

Compound	DS	-PMF04	Jain	Ludi1	Ludi2	Ludi 3
2-O-caffeoyl tartaric acid	215.079	144.63	5.42	870	698	789
Chicoric acid	206.092	177.22	5.00	915	694	826
Mumefural	201.985	136.75	8.50	1028	822	799
2-O-feruloyl tartaric acid	198.739	145.13	5.16	833	661	693
Rosmarinic acid	148.434	145.59	5.40	971	743	921
Quinic acid	143.961	106.56	1.49	557	487	475
Genipinic acid	142.772	110.31	2.53	530	467	651
Digallic acid	142.547	147.27	1.67	953	708	848
5-O-feruloylquinic acid	142.462	149.26	6.90	1011	784	865
4-O-feruloylquinic acid	140.488	142.14	7.45	946	746	878
Glutathione	66.787	127.28	6.39	564	447	412

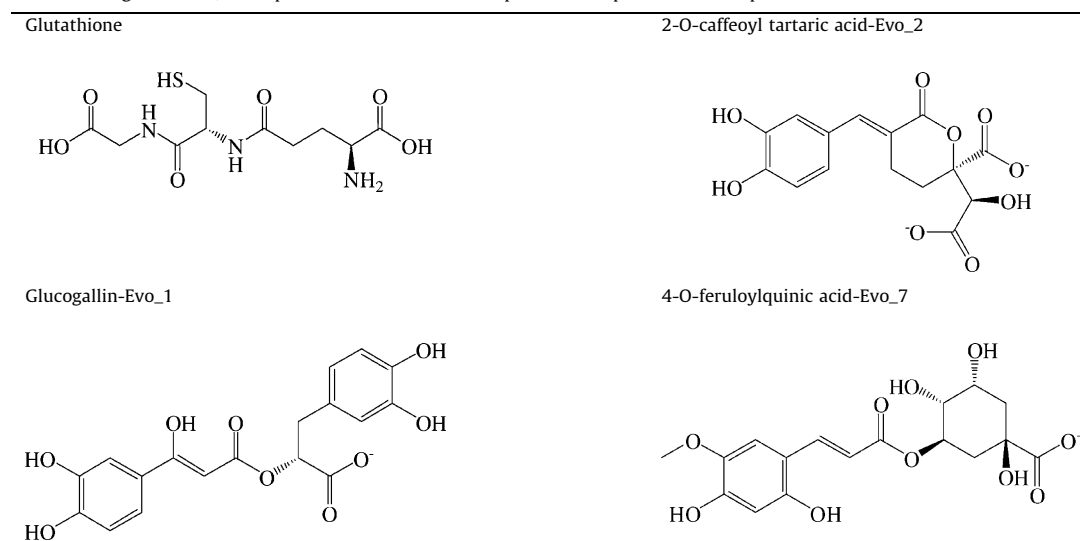
DS: Dock Score; PMF: Potential of Mean Force.

Table 2
Docking results for *de novo* products. The top 5 candidates are shown.

Compound	DS	-PMF04	Jain	Ludi1	Ludi2	Ludi3
2-O-caffeoyl tartaric acid-Evo_2	222.198	136.13	5.65	701	569	656
Glucogallin-Evo_1	169.762	158.84	6.12	1062	795	974
4-O-feruloylquinic acid-Evo_7	167.056	152.93	8.01	999	783	919
1-Caffeoylquinic acid-Evo_3	165.916	150.93	5.90	774	591	674
4-O-feruloylquinic acid-Evo_5	159.929	160.19	7.34	1071	822	950
Glutathione	66.787	127.28	6.39	564	447	412

DS: Dock Score; PMF: Potential of Mean Force.

Table 3
Structures of glutathione, the top three derivatives and the parental compounds of the top three derivatives.



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