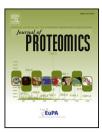
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JOURNAL OF PROTEOMICS XX (2013) XXX-XXX



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Dracula's children: Molecular evolution of vampire bat venom

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16 A R T I C L E I N F O

Article history: 28 Received 29 March 2013 29 Accepted 28 May 2013 20 25 26Keywords: 56Molecular evolution $5\overline{8}$ Vampire bat $\overline{29}$ Venom 50 Positive selection 60 Desmodus rotundus <u>9</u> <u>63</u> 34 35 36 37 38 39 4041 42 43 44 45

ABSTRACT

While vampire bat oral secretions have been the subject of intense research, efforts have concentrated only on two components: DSPA (Desmodus rotundus salivary plasminogen activator) and Draculin. The molecular evolutionary history of DSPA has been elucidated, while conversely draculin has long been known from only a very small fragment and thus even the basic protein class was not even established. Despite the fact that vampire bat venom has a multitude of effects unaccounted by the documented bioactivities of DSPA and draculin, efforts have not been made to establish what other bioactive proteins are secreted by their submaxillary gland. In addition, it has remained unclear whether the anatomically distinct anterior and posterior lobes of the submaxillary gland are evolving on separate gene expression trajectories or if they remain under the shared genetic control. Using a combined proteomic and transcriptomic approach, we show that identical proteins are simultaneously expressed in both lobes. In addition to recovering the known structural classes of DSPA, we recovered a novel DSPA isoform as well as obtained a very large sequence stretch of draculin and thus established that it is a mutated version of the lactotransferrin scaffold. This study reveals a much more complex secretion profile than previously recognised. In addition to obtaining novel versions of scaffolds convergently recruited into other venoms (allergen-like, CRiSP, kallikrein, Kunitz, lysozyme), we also documented novel expression of small peptides related to calcitonin, PACAP, and statherin. Other overexpressed protein types included BPI-fold, lacritin, and secretoglobin. Further, we investigate the molecular evolution of various vampire bat venom-components and highlight the dominant role of positive selection in the evolution of these proteins. Conspicuously many of the proteins identified in the proteome were found to be homologous to proteins with known activities affecting vasodilation and platelet aggregation. We show that vampire bat venom proteins possibly evade host immune response by the mutation of the

1874-3919/\$ – see front matter © 2013 Published by Elsevier B.V. http://dx.doi.org/10.1016/j.jprot.2013.05.034

Please cite this article as: Low DH.W., et al, Dracula's children: Molecular evolution of vampire bat venom, J Prot (2013), http://dx.doi.org/10.1016/j.jprot.2013.05.034

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JOURNAL OF PROTEOMICS XX (2013) XXX-XXX

such neglected taxon for biodiscovery.

and development.

Biological significance

surface chemistry through focal mutagenesis under the guidance of positive Darwinian

selection. These results not only contribute to the body of knowledge regarding haematophagous

venoms but also provide a rich resource for novel lead compounds for use in drug design

These results have direct implications in understanding the molecular evolutionary history

of vampire bat venom. The unusual peptides discovered reinforce the value of studying

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Q468 1. Introduction

Venom is defined as a secretion produced in a specialised gland 60 in an animal, which is delivered to a target animal by inflicting a 70 71 wound (regardless of size). Venom must also contain molecules (toxins) that disrupt normal physiological and/or biochemical 72processes so as to facilitate feeding or defence by the producing 73animal [1]. This definition includes secretions produced by 74 blood feeding (haematophagous) specialists such as fleas, ticks, 75 leeches or vampire bats that disrupt the haemostatic defences 76 of prey/host organisms. These secretions which facilitate 77 haematophagy or blood feeding are considered a particular 78 79subtype of venom.

With the level of infamy that extends far beyond the 80 boundaries of science, blood-sucking vampire bats (Chiroptera, 81 Desmodotinae) have been the subject of folk tales, superstitions 82 and stories associated with the legendary Count Dracula for 83 centuries [2,3]. All the three species of vampire bats are confined 84 to Central and South America and typically live in the caves, 85 tree hollows, and abandoned mines [4]. The relatively rare 86 hairy-legged vampire bat (Diphylla ecaudata) feeds exclusively on 87 avian hosts, while the white-winged vampire bat (Diaemus 88 89 youngi) thrives on both the mammalian and avian blood, but 90 most likely favouring the latter [2,5]. In contrast, the common 91 vampire bat, Desmodus rotundus feeds overwhelmingly on 92 mammals and has established itself in large colonies over an 93 increasingly extensive distribution [2,6,7]. The expanding population of these bats is attributed to the increasing human 94 population and the associated large number of domesticated 95animals and livestock, which provide a constant, high-density 96 97 food supply [4,7].

All three species of vampire bats are highly specialised 98 for a haematophagous lifestyle, especially D. rotundus [4]. 99 100 Modifications of the teeth and limbs of vampire bats facilitate this lifestyle [6]. Reinforcement of limb strength permits 101 D. rotundus and the other vampire bat species to approach 102their prey from the ground via quadrupedal walking and 103 jumping [5,7,8]. D. rotundus is the most specialised of all 104 vampire bats, and possesses specialised sensory capabilities 105for the detection of prey [9]. Close-range thermal and mechan-106ical sensitivity is utilised for locating capillaries during feeding, 107 while long-range vision, olfaction, acute hearing and echoloca-108 109tion are utilised for the discovery of potential prey [9]. They also have razor-like upper and lower incisors [5,7,10] that inflict a 110 crater-shaped wound unique to D. rotundus. While feeding, their 111 tongue darts in and out of the wound, releasing venom from the 112 113 dorsal side of the tongue while drawing in blood via two straw-like ducts located on the ventral side of the tongue [7]. 114

In order to facilitate blood-feeding, D. rotundus must be 115 capable of interfering with their prey's natural haemostatic 116 response to injury during feeding and digesting [11,12]. 117 A typical haemostatic response produces a fibrin clot within 118 minutes of the infliction of a wound, preventing further blood 119 loss. The response commences with the constriction of blood 120 vessels, restricting blood flow to the wound, and is followed 121 by the adhesion of activated platelets to the site of injury and 122 the conversion of fibrinogen to insoluble fibrin, forming a 123 blood clot [12]. In contrast to this normal response to injury, 124 bleeding from a wound induced by vampire bats may be 125 prolonged from minutes to hours, ensuring a constant flow of 126 blood for the bat to feed upon [12].

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The gland predominantly associated with haematophagy in 128 D. rotundus is the principle submaxillary gland, which consists 129 of two separate anterior and posterior lobes [13]. This gland is 130 responsible for the secretion of venom with strong anticoagu-131 lant and proteolytic activities [13]. The venom delays the onset 132 of blood clotting by interfering with fibrin formation or acting 133 upon fibrin as it is converted from fibrinogen. In addition it has a strong proteolytic action that breaks up any blood clots that 135 may be formed. This proteolytic action is accomplished through 136 the activation of the host's fibrinolytic system which converts 137 plasminogen to plasmin, solubilising and removing fibrin clots 138 to prevent excessive fibrin buildup at the site of the wound. 139

The venom of the vampire bat has been documented to 140 disrupt the coagulation cascade via four distinct mechanisms: 141 (i) inhibition of factor IXa, (ii) inhibition of factor Xa, 142 (iii) activation of plasminogen and (iv) inhibition of platelets 143 [1]. A venom component that has been researched consider- 144 ably is draculin, an anticoagulant factor. It is an 88.5 kDa 145 glycoprotein that inhibits activated factors IX (IXa) and X (Xa) 146 of the coagulation mechanism [11,14,15]. The irreversibly 147 bonded complex of Xa-Draculin forms immediately upon 148 contact, inhibiting factors IXa and Xa [14]. The inhibition 149 of IXa and Xa prevents the conversion of prothrombin to 150 thrombin, which in turn prevents the conversion of fibrino- 151 gen to insoluble fibrin. Furthermore, the characteristic of 152 non-competitive inhibition prevents the cleavage of draculin 153 from Xa after binding, thus maintaining the toxin's antico- 154 agulant activity during feeding and digestion [16]. Interest- 155 ingly, a study conducted recently described the capacity of 156 D. rotundus prey to develop immunity to draculin if they were 157 targeted and fed upon over prolonged periods [17]. This kind 158 of predator-prey arms race scenario is similar to that observed 159 between other venomous animals and their prey. 160

The plasminogen activators or Desmokinase [12], better 161 known as DSPA (Desmodus rotundus salivary plasminogen 162

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