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A swelling-based chemiresistor for a biogenic odour

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ABSTRACT

Escherichia coli bacteria release 1-decanol as a byproduct of their metabolism. We demonstrate the detection of 1-decanol odour at a partial pressure in the order 100 ppb by the resistance change of a swelling-based sensor, consisting of Langmuir–Schäfer deposited Au core/organic ligand shell nano-particle films. This is an exceptionally low limit of detection for swelling-based sensors, and relies firstly, in the careful matching of the CSNPs ligands to the targeted odour, and secondly, in the very low volatility of this odour. Sensor response can be substantially increased further when films are cooled below the freezing point of 1-decanol. We observe unexpected quantitative behaviour of our sensors: response is only weakly dependent on the odour's partial pressure, and scales differently with temperature than the response of other Au-CSNP odours to more volatile odours. This may be related to their unusually strong thermal resistance drift, the difficulties in delivering very low partial pressure odour atmospheres, and the proximity to the analyte's freezing point.

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1. Introduction

'Swelling-based' chemiresistor sensors use composites (typically, thin films) of an electrically insulating matrix, filled with conductive particles. Sensitivity and selectivity to analytes result from the selective swelling of the matrix in some analyte odours, a consequential increase in the separation of conductive particles, and a resulting increase of electrical resistance, R (or, decrease of conductance, *G*), which is monitored readily. The classic examples of swelling-based sensors are insulating polymers filled with carbon black (CB) particles, e.g. [1]. The same concept has been downscaled to the nanoscale, using films of core-shell nanoparticles (CSNPs), typically with Au cores decorated with thiolcoupled, insulating organic ligand shells [2–8]. Typically, Au CSNP sensors have been used to detect odours of solvents or fuels. The relevant concentration benchmark for flammable odours is the 'lower explosive limit' (LEL), which typically is a few 1000 or 10,000 ppm of atmospheric pressure [9] (we understand 'ppm' as partial atmospheric pressure throughout this contribution). For example, the LEL of iso-octane, the main component of petrol, is 7900 ppm. Biologically relevant odours often occur at much lower concentrations e.g. [10], and their sensing traditionally relies in specific chemical 'lock/key' recognition, which often is inspired by their biological functioning rather than the more generic swelling.

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However, recent progress in the understanding of swellingbased Au CSNP sensors [8,11] has encouraged us to attempt the sensing of a biologically relevant odour at sub-ppm concentration by swelling, without specific molecular recognition. Assuming suitable ligands are chosen (e.g., alkanethiol ligands for alkane or aromatic odours), Lewis et al. [8] have shown that the sensitivity, s_R , of Au CSNP sensors is only weakly dependent on the length of ligands, and the identity of the odour, if s_R is defined as the slope of the sensors' relative resistance change, $\Delta R/R$, plotted against the vapour's partial pressure expressed as a fraction of the same odour's saturated vapour pressure, p/p_{sat} . Under this unusual pressure normalisation convention, s_R for a variety of ligands, and hydrocarbon vapours, fall into a small range (0.8-2). s_R somewhat increases for longer ligands, and for odours chemically similar to the ligands, but it remains confined to this rather narrow interval. This implies that odours with low volatility, i.e. low p_{sat} , can be detected at much lower partial pressures (vapour pressure expressed as fraction of atmospheric pressure, p/p_{atm}) than highly volatile odours. We have since directly confirmed the link between volatility and sensitivity by showing that sensitivity of swelling-based Au CSNP sensors increases manifold when sensors are cooled with respect to ambient temperature, thus reducing the volatility of the odour in the swollen matrix [11].

However, this does not imply that there is a sensitivity advantage for the detection of low-volatility odours with swelling-based sensors, when the source of the odour is e.g. an accidental spillage, as it would be likely for explosive or



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poisonous odours: volatility controls both, the build-up of vapour atmosphere from the spillage, and the degree of swelling of the sensor matrix. Hence, there is no overall advantage (nor disadvantage) for sensing low volatility odours. The situation is different when instead the odour is biogenic, i.e. its source are living organisms. Life forms are not in thermodynamic equilibrium, therefore vapour build-up is controlled by the organism's rate of metabolism, rather than the vapour's volatility. Matrix swelling, on the other hand, still is controlled by volatility. We therefore expect a sensitivity advantage for 'heavy' (low volatility) biogenic odours, because vapour build-up is no longer limited by low volatility, but low volatility still enhances swelling.

Here, we report on the sensing of 1-decanol, a biogenic odour with low saturated vapour pressure, by its swelling of an Au-CSNP at less than 1 ppm odour concentration. 1-Decanol is released by strains of Escherichia coli (E. coli) [12]. Since some serotypes of E. coli are associated with serious food- and water-borne infections [13-16], the sensing of 1-decanol is relevant for food health and safety monitoring. Previous attempts at sensing 1-decanol released by E. coli involved the pumping of headspace air above an E. coli culture through a filter, subsequent extraction of the 1-decanol from the filter by a solvent, and chromatographic determination of the concentration of 1-decanol in the extraction solvent. Results ranged from 23.6 ng/mL to 148 ng/mL [12], however, concentration in ng of 1-decanol per mL extraction solvent does not allow a direct conclusion on the 1-decanol partial pressure in the original atmosphere. Hence, currently, neither convenient 1-decanol sensors nor typical partial pressures of 1-decanol from biological sources are available.

Interestingly, 1-decanol freezes at 6.4 °C, which we can easily access with a Peltier cooler [11]. This allows us to investigate the behaviour of a swelling-based sensor when temperature drops below the analyte's freezing point.

2. Experimental

2.1. Materials

As the material for our swelling-based sensors, we used monodisperse Au CSNPs with self-assembled 11-mercapto-1undecanol ('undecanolthiol') ligands, sourced from PlasmaChem [17]. Ligands were selected for their similarity to the target analyte.

2.2. Sample preparation

Nanoparticles were dissolved in methanol (1 mg/mL), rather than in chloroform as used for alkanethiol CSNPs [7,11], because a good dispersion could not be achieved in chloroform even after sonication. 400 μ L of such solutions were spread on a Nima Langmuir trough and compressed to 11 mN/m, a Langmuir isotherm is shown in Fig. 1. We used the Langmuir–Schäfer (LS) technique for 5 deposition cycles on glass substrates, previously cleaned and silanised with hexamethyldisilazane (HMDS). For a control experiment, Au-dodecanethiol CSNP films were prepared similarly, as reported previously [7].

2.3. Resistance measurements

The electric resistance of resulting films was measured with an AlphaLab Teraohm meter (HR2 Model). The baseline resistance prior to exposure was averaged over 3 min. Although we kept film deposition procedure as constant as we possibly could, we observed baseline resistances for different Au-undecanolthiol samples ranged from $\sim 250 \text{ M}\Omega$ to 1 G Ω prior to any vapour



Fig. 1. Langmuir isotherm of Au-undecanolthiol CSNP film spread on water from methanol dispersion. LS deposition was at 11 mN/m.

exposure. We believe this variation results from the difficulty in ensuring that Au-undecanolthiol Langmuir films are strictly monolayers. Unlike the Langmuir isotherms for alkanethiol CSNPs [7,11], the isotherm in Fig. 1 shows no defined monolayer collapse. The differences between alkane- and alkanol-ligand Au CSNPs with regards to both, suitable dispersion solvent and isotherms, probably result from the hydrophilic terminal hydroxyl groups present in the alkanol ligands.

2.4. Odour exposure

For odour sensing tests, we sourced 1-decanol from Aldrich and generated saturated 1-decanol odour by bubbling inert carrier gas (N₂) through a sparger in a phial that was held at 25 °C in a thermostatted water bath. Saturated odour was then diluted by mixing with carrier gas as required, e.g. down to 1% or 10% p_{sat} . The saturated vapour pressure of 1-decanol is quoted as 11.2 ppm at 25 °C [18,19]. When we quote 1-decanol vapour pressures as 112 ppb or 1.1 ppm, these are to be understood as 1% or 10% p_{sat} of 1-decanol at 25 °C. 1-decanol odour was fed into a Teflon-lined exposure chamber where samples were located. Samples could be cooled with a Peltier element, which was heat-sinked into an ice bath. The exposure set-up is sketched in Fig. 2.

3. Results and discussion

Fig. 2 shows the response of a Au-undecanolthiol CSNP sensor film under exposure/recovery cycles to 112 ppb 1-decanol odours. We find a small, but clearly observable resistance increase $\Delta R/R$ of approximately 0.4% under exposure, which is more than 10 times larger than the noise in $\Delta R/R$. Under purge, the resistance recovers fully to its previous value. We thus observe a response at a partial pressure that is remarkably low for a swelling-based sensor, e.g., Lewis et al. exposed Au CSNP sensors to various analytes (e.g., alkanes, alcohols, toluene) at odour concentrations in the order 100–1000 ppm for a resistance change in the order 2% [8]. However, all their analytes were significantly more volatile than 1-decanol. This confirms our premise that swelling-based sensors can detect 'heavy' (i.e., low volatility) odours at remarkably low partial pressure.

For comparison, we also exposed a Au-dodecanethiol CSNP film to 1-decanol up to 10% p_{sat} = 1.1 ppm. We have used such films in previous work, and found good sensitivity to aromatic and alkane odours, e.g. toluene and decane [7]. However, even at 10% p_{sat} 1-decanol, there was no measurable resistance change, while the same Au-dodecanethiol film did clearly respond e.g. to toluene, when a soaked cotton bud was placed nearby. Lewis et al. [8] have

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