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## **Bioorganic Chemistry**

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# Design, synthesis, analgesic, anti-inflammatory activity of novel pyrazolones possessing aminosulfonyl pharmacophore as inhibitors of COX-2/5-LOX enzymes: Histopathological and docking studies



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#### ARTICLE INFO

#### Article history: Received 5 December 2017 Revised 27 January 2018 Accepted 7 March 2018 Available online 8 March 2018

Keywords: Pyrazolone Analgesic Anti-inflammatory COX-2 5-LOX

#### ABSTRACT

A series of newly synthesized 4-aryl-hydrazonopyrazolones were designed and their structures were confirmed by spectral and elemental analyses. All synthesized compounds were evaluated for their *in vitro* COXs, 5-LOX inhibition, *in vivo* analgesic and anti-inflammatory activities. Compounds **5d**, **5f** and **5i** were found to be the most potent COX-2/5-LOX inhibitors with superior COX-2 selectivity index values (SI = 5.29–5.69) to reference standard celecoxib (SI = 3.52). Four compounds; **5b**, **5c**, **5d** and **5f** showed excellent anti-inflammatory activity (% edema inhibition = 72.72–54.54%) and perfect ED<sub>50</sub> values (ED<sub>50</sub> = 0.044–0.104 mmol/kg) relative to celecoxib (ED<sub>50</sub> = 0.032 mmol/kg). To explore the most active compounds, ulcerogenic effect on stomach in comparison with indomethacin and celecoxib in addition to histopathological investigations were performed. Compound **5f** showed better gastric profile (UI = 2.33) than celecoxib (UI = 3.00). Also, **5f** caused 50% increase in thermal pain threshold close to reference drug indomethacin (53.13%). Docking study of all the target compounds into COX-2 and 5-LOX active sites was performed to rational their anti-inflammatory activities.

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#### 1. Introduction

The inflammatory mediators, prostaglandins (PGs), leukotrienes (LTs) and thromboxanes (TXs) are responsible for inflammation, other pathological and physiological processes [1]. They are generated from arachidonic acid (AA), a poly saturated fatty acid released from membrane phospholipids metabolism by the action of cyclooxygenase (COX-1, -2, -3) and lipoxygenase (5-LOX, 8-, 12-, 15-) enzymes [2–4].

COX enzymes are responsible for the production of PGs and TXs. COX-1 is a constitutive enzyme found in the stomach, platelets and kidneys as a "house-keeper" enzyme and involved in gastric protection, platelet aggregation and normal kidney functions. COX-2

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is an inducible enzyme found in macrophages, fibroblasts and leukocytes and stimulated in response to pro-inflammatory mediators. COX-3, the third cyclooxygenase is latterly discovered and present in central nervous system [5–8].

Classical non-steroidal anti-inflammatory drugs (NSAIDs) such as aspirin and indomethacin exert their therapeutic action *via* suppressing PGs bio-synthesis through non-selective inhibition of COX-1 and COX-2 enzymes resulting in serious adverse effects like gastric pain, bleeding, ulcer and kidney complications [9,10].

COX-2 selective NSAIDs (Coxibs) illustrated by celecoxib (Celebrex®), valdecoxib (Bextra®) and rofecoxib (Vioxx®) have no effect on gastric mucosal prostaglandin. But recent studies have shown the risk of some highly selective COX-2 inhibitors to increase the incidence of myocardial infarction leading to cardiac arrest due to alteration in the COX-1/COX-2 biochemical pathway [3,11,12].

5-LOX is a human non-heme enzyme responsible for the production of LTs involved in the inflammatory process. Zileuton, licofelone and meclofenamate sodium (Meclomen®) are examples of orally active 5-LOX inhibitors [13,14].

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The incident of NSAIDs side effects is thought to be due to inhibition of one enzyme pathway (COX) over the other (LOX) pathway leading to shift in AA metabolism [15], hence the development of new anti-inflammatory (AI) agents targeting both metabolic pathways of AA (COX-2 and 5-LOX) inhibition is a worthy rational approach to obtain effective and safe NSAIDs [14,16,17].

Pyrazolone ring system is a core structure in numerous drugs displaying analgesic and AI activities such as aminoantipytine, propyphenazone and famorofazone [18–20].

Several research studies reported the enhanced biological activities of heterocyclic compounds incorporating hydrazono pharmacophore as analgesic and AI agents with improved gastric profile due to its dual COX/5-LOX inhibitory activities [18,21–23].

Furthermore, structure activity relationship studies of selective COX-2 inhibitors demonstrated the importance of aminosulfonyl (SO<sub>2</sub>NH<sub>2</sub>) pharmacophore for COX-2 selectivity [24–26].

On the basis of these findings and in continuation of our previous work [27–32] to develop effective AI agents devoid from adverse effects, we describe the design, synthesis, analgesic and AI activities of novel 4-aryl-hydrazonopyrazolone derivatives incorporating both sulfonamoyl and hydrazone pharmacophores as COX-2 and 5-LOX inhibitors (Fig. 1). Ulcerogenic liability and histopathological screening were performed in order to identify the non-ulcerogenic AI active compounds. Docking studies were also performed to understand the possible binding modes of the synthesized compounds into both COX-2 and 5-LOX active sites in order to explain their AI activities.

#### 2. Results and discussion

#### 2.1. Chemical synthesis

The synthetic pathways adopted for starting materials **2a–e** and target compounds **3**, **4**, **5a–i** are illustrated in Scheme **1**, **2**.

Hydrazones **2a–d** were prepared *via* coupling the diazonium salt of different primary aromatic amines with the active methylene group of ethyl acetoacetate [33].

**Fig. 1.** Structure of aminoantipyrine, CBS 1108, celecoxib, and general structure of targeted pyrazolones (A) and (B).

3-0xo-2-[(4-sulfamoylphenyl)-hydrazono]-butyric acid ethyl ester (**2e**) was prepared through diazotization of sulfanilamide **1e** and coupling the formed diazonium salt with ethyl acetoacetate in presence of sodium acetate. IR spectrum of compound **2e** demonstrated two absorption bands at 1690 and 1676 cm<sup>-1</sup> corresponding to two C=O groups while <sup>1</sup>H NMR spectrum showed a D<sub>2</sub>O exchangeable peak at  $\delta$  11.60 ppm indicating NH proton. Also, the presence of a singlet CH<sub>3</sub>, a triplet CH<sub>3</sub> and a quartet CH<sub>2</sub> peaks at  $\delta$  2.47, 1.28 and 4.29 ppm corresponding to acetyl (COCH<sub>3</sub>) and ethoxy (OCH<sub>2</sub>CH<sub>3</sub>) protons respectively confirmed the structure. These spectral data demonstrated the existence of compounds **2a-f** in hydrazone form (*i*) rather than azo form (*ii*) (Scheme 1).

Cyclo-condensation ethanolic solution of **2e** with an equimolar amount of hydrazine hydrate afforded pyrazolone **3** in 74% yield. 
<sup>1</sup>H NMR spectrum of **3** revealed the disappearance of the signals due to ethoxy protons of the parent ester **2e**. The presence of additional  $D_2O$  exchangeable singlet signal at  $\delta$  13.22 ppm corresponding to pyrazolone NH proton, confirmed the reaction. In addition, the mass spectra of **3** displayed molecular ion peak at m/z 281 (92.94%)

Heating pyrazolone **3** with acetyl chloride afforded the *N*-acetylpyrazolone **4** in a good yield of 63%. IR spectrum of **4** displayed an additional absorption band at 1749 cm<sup>-1</sup> due to the carbonyl group of *N*-acetyl moiety. The absence of pyrazolone NH peak of the precursor **3** and presence of a peak of an additional signal of three protons integration due to acetyl moiety at  $\delta$  2.45 ppm in <sup>1</sup>H NMR spectrum of **4** confirmed the reaction. Also, <sup>13</sup>C NMR spectrum of compound **4** revealed the presence of two peaks at  $\delta$  24.20 and  $\delta$  171.61 ppm corresponding to COCH<sub>3</sub> and COCH<sub>3</sub>, respectively.

Different substituted phenylhydrazine hydrochlorides were heated with hydrazones **2a–e** under reflux in absolute ethanol to give the target compounds **5a–i** in excellent yield (68–85%). The reaction proceeds *via* addition of the more nucleophilic NH<sub>2</sub> hydrazine group to the reactive acetyl carbonyl group (COCH<sub>3</sub>) followed by intra-molecular cyclization through nucleophilic substitution of the good leaving ethoxy group and loss of an ethanol molecule (Scheme 2).

The structure of diarylpyrazolones **5a-i** was investigated by elemental and spectral analyses. The IR spectra of compounds 5a-i indicated the presence of two absorption bands at 3460-3313 cm<sup>-1</sup> and 3279-3247 cm<sup>-1</sup> corresponding to NH<sub>2</sub> and NH groups in addition to an absorption band at 1672-1650 cm<sup>-1</sup> indicating C=O group at C-5 of pyrazolone ring. Carboxy compounds, 5b, 5c, 5d, 5f, 5g, 5h displayed two additional absorption bands at 3440-3416 cm<sup>-1</sup> and 1701-1680 cm<sup>-1</sup> due to carboxylic OH and C=O groups, respectively. <sup>1</sup>H NMR spectra of 5a-i showed the presence of a singlet signal at δ 2.29-2.34 ppm corresponding to methyl protons at C-3 of pyrazolone ring, in addition to two exchangeable singlet signals at δ 7.22-7.42 corresponding to NH<sub>2</sub> protons of aminosulfonyl moiety, and at  $\delta$  13.03–13.66 for NH protons. Carboxy compounds 5b, 5c, 5d, 5f, 5g and 5h displayed carboxyl protons at  $\delta$  13.03–15.05 ppm. Also, <sup>13</sup>C NMR spectra of 5a-i exhibited the methyl carbon at δ 12.14-12.30 ppm and carbonyl carbons at  $\delta$  154.71–156.99 ppm. In Addition, <sup>13</sup>C NMR spectra of carboxy compounds **5b**, **5c**, **5d**, **5g**, **5h** displayed carboxy carbons at δ 167.17–168.67 ppm which confirmed the structure.

#### 2.2. Biological activity

#### 2.2.1. Analgesic activity (hot plate latency test)

Analgesic activity of compounds **3**, **4** and diarylpyrazolones **5a-i** was evaluated applying hot plate latency test [34]. Oral administration of tested compounds produced a significant delay in the latency time relative to basal values except for compounds **3**, **4**,

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