



Synthesis of novel 6-substituted amino-9-(β -D-ribofuranosyl)purine analogs and their bioactivities on human epithelial cancer cells

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ABSTRACT

New nucleoside derivatives with nitrogen substitution at the C-6 position were prepared and screened initially for their *in vitro* anticancer bioactivity against human epithelial cancer cells (liver Huh7, colon HCT116, breast MCF7) by the NCI-sulforhodamine B assay. N⁶-(4-trifluoromethylphenyl)piperazine analog (**27**) exhibited promising cytotoxic activity. The compound **27** was more cytotoxic (IC₅₀ = 1–4 μ M) than 5-FU, fludarabine on Huh7, HCT116 and MCF7 cell lines. The most potent nucleosides (**11**, **13**, **16**, **18**, **19**, **21**, **27**, **28**) were further screened for their cytotoxicity in hepatocellular cancer cell lines. The compound **27** demonstrated the highest cytotoxic activity against Huh7, Mahlavu and FOCUS cells (IC₅₀ = 1, 3 and 1 μ M respectively). Physicochemical properties, drug-likeness, and drug score profiles of the molecules showed that they are estimated to be orally bioavailable. The results pointed that the novel derivatives would be potential drug candidates.

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Cancer is one of the most important causes of death in the world, with almost 14 million new patients and 8.2 million deaths from cancer in 2012.¹ Therefore, development of new potent and selective anticancer agents is of high interest to medicinal chemistry. Nucleobase and nucleoside analogs are often exploited as chemotherapeutic agents in both hematologic malignancies and solid cancers. Nucleobases and nucleosides are the nucleotide precursors; therefore, they are considered as antimetabolites. Nucleotide compounds of similar structure, are involved in many cell processes such as cell growth and division, hence nucleobase and nucleosides have often been exploited as antineoplastic agents.^{2,3} The mechanism of action of nucleobase analogs is through induction of apoptosis.⁴ 5-Fluorouracil which is a nucleobase derivative with fluorine atom, is a frequently preferred anticancer agent for a variety of malignancies in clinics.⁵ Similarly, other pyrimidine nucleosides like cytarabine and gemcitabine have been described as antimetabolite anticancer drugs.⁶ For the last six decades, 6-mercaptopurine and 6-thioguanine have been used as a nucleic acid metabolism inhibitor for the treatment of paediatric acute lymphoblastic leukaemia.⁷ Furthermore, purine nucleosides such as fludarabine, cladribine, and pentostatin, have become estab-

lished to be effective against haematological malignancies.⁸ These analogs achieve an unbalance in dNTP pool via inhibition of the ribonucleotide reductase enzyme that induces degradation in DNA synthesis.⁹ Therefore, nucleosides with anticancer bioactivities induce apoptotic cell death in general.⁶

Primary liver cancer, hepatocellular carcinoma (HCC) is second deadly cancer worldwide (GLOBOCAN 2012). It is the fifth most common cancer in men and seventh in women, accounting for 7% of all cancer cases, worldwide with around 700,000 new cases each year.^{10–12} Ethological factors for primary liver cancer are mainly HBV or HCV infection, chronic alcohol consumption, obesity and environmental toxins (aflatoxin B).^{10,13} Prognosis of HCC patients is usually very poor due to the resistance against conventional chemotherapeutic agents. Sorafenib and regorafenib are FDA approved multikinase inhibitors, which extend patient survival only 3 months with liver cancer.^{14–17} Therefore, it is essential to identify new candidate therapeutic agents for hepatocellular carcinoma.^{18,19}

We have previously exploited purine and purine nucleoside derivatives, which have displayed promising cytotoxic activities in liver cancer cells. The molecules from those studies had significant bioactivities on liver cancer cells. The compound N⁶-(4-trifluoromethylphenyl)piperazine nucleoside (IC₅₀ = 5.2–9.2 μ M) induced senescence and purine analogs (IC₅₀ = 0.1–0.8 μ M) lead to apoptotic cell death in HCC cell lines.^{20,21} Therefore, we designed

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novel compounds with amine and chlorine electronegative substituents at the C-2 position of the purine ring. These molecules were then synthesized as a new series of 6-substituted amino-9-(β -D-ribofuranosyl)purine derivatives (**9–22**, **27**, **28**) and their cytotoxic activities were screened in human epithelial cancer cells (liver Huh7, colon HCT116, breast MCF7). The bioactivities of the most potent nucleoside derivatives (**11**, **13**, **16**, **18**, **19**, **21**, **27**, **28**) were further analyzed in hepatocellular cancer cell lines (see Figure 1).

The piperazine-containing nucleoside analogs (**9–16**) were synthesized as shown in Scheme 1. In the first transformation, inosine and guanosine are converted to the 6-chloro nucleoside (**5**, **6**) with the trifluoroacetic acid anhydride, thionyl chloride method developed by Robins for 2'-deoxyinosine.²² Trifluoroacetyl groups were used for transient hydroxyl protection instead of stability of the glycosidic bond. These groups were readily removed by methanolysis after the chlorination reaction. The inosine and guanosine derivatives (**9–16**) were prepared via nucleophilic aromatic substitution of compounds **7**, **8** with 4-substituted piperazines.

Nucleosides substituted with 4-substituted anilines/2-substituted ethyl amines at the position C-6 (**17–22**), were obtained with nucleophilic aromatic substitution reaction of 6-chloro-9-(β -D-ribofuranosyl)purine (**7**) with the suitable anilines and amines under basic conditions (Scheme 1).

The 2-chloro-6-(4-substituted piperazine)/6-(2-substituted ethyl amino) purine analogs (**27**, **28**) were prepared as shown in Scheme 1. 2,6-Dichloropurine (**23**) was condensed with the acetylated ribofuranose under microwave irradiation for 30 min to get 2,6-dichloro-nucleoside derivative (**24**) in good yield of 79%. The yield obtained as a result of this reaction was significantly higher than the yield in the previously reported method.^{23,24} Displacement of the 6-chloro group was made by nucleophilic aromatic substitution by the substituted piperazine or ethyl amine. Removal of the acetyl groups as the protecting group was made by NaOMe to obtain purine nucleoside analogs **27**, **28**. The structures of the all compounds were confirmed by ¹H, ¹³C NMR mass spectral data and elemental analysis.

The *in vitro* cytotoxicity of the compounds **9–22**, **27**, **28** were initially analyzed on Huh7 (liver), HCT116 (colon) and MCF7 (breast) cancer cells, using a sulforhodamine B (SRB) assay.²⁵ The IC₅₀ values for each compound also were calculated in comparison with the known nucleobase analog 5-fluorouracil (5-FU), nucleoside analogs fludarabine and cladribine and the results were shown in Table 1. Among the synthesized compounds, analogs accommodating substituted piperazine moiety at their C-6 position (**9–16**, **27**), the one with promising IC₅₀ values against Huh7 (2 μ M), HCT116 (1 μ M) and MCF7 (4 μ M) is trifluoromethylphenyl substituted piperazine analogs (**27**). Nucleoside **27** displayed

significant cytotoxic activity for all the cell lines screened. When, IC₅₀ values compared with 5-FU, fludarabine, the compound **27** had displayed lower values, which are in micromolar concentrations. Compound **27** established a better cytotoxic activity on Huh7 cell (2 vs. 30 and 30 for 5-FU, fludarabine respectively), HCT116 (1 vs. 4 and 8 for 5-FU and fludarabine) and MCF7 cells (4 vs. 3 and 15 for 5-FU and fludarabine). Also compound **16**, bearing a diphenylmethyl substituent at piperazine moiety of the nucleoside, had higher cytotoxic activities when compared to 5-FU and known nucleoside drug fludarabine, on Huh7 cells. The substitution of (2-cyclohexenylethyl)amino at C-6 position improved the cytotoxic activity of compound **28** and the IC₅₀ values for 72 h of treatment were comparable to those of 5-FU and fludarabine on Huh7 cell line.

We then analyzed the cytotoxic activities of the most potent nucleoside derivatives (**11**, **13**, **16**, **18**, **19**, **21**, **27**, **28**) in a panel of HCC cells: Huh7, HepG2, Mahlavu, and FOCUS (Table 2). N⁶-Trifluoromethyl nucleoside analog **27** demonstrated the best cytotoxic activity, with IC₅₀ values of 1–3 μ M against Huh7, Mahlavu and FOCUS cells (Table 2). The 2-Cyclohexenylethyl amino derivative **21** was also found to be significantly bioactive (IC₅₀ 1 μ M) on HepG2 cell line. Compounds **27** and **21** had a better cytotoxic activity than the known cytotoxic drugs 5-FU and fludarabine on HepG2 cells. When there was a bigger diphenylmethyl group at the piperazine (**16**), we observed that compound **16** had displayed lower values in micromolar concentrations. Furthermore, nucleoside **13**, which had no substitution at the phenyl ring, were cytotoxic to FOCUS cell line with an IC₅₀ values of 9 μ M.

Nucleoside **21** being one of the most active compound, was showed noteworthy IC₅₀ values (IC₅₀ = 6 μ M) on HCT116 which were comparable to that of 5-FU (IC₅₀ = 4 μ M) and to that of nucleoside analog fludarabine (IC₅₀ = 8 μ M) (Table 1). Similarly, the cytotoxic activity on MCF7 cancer cells was significantly low with nucleoside **21** (IC₅₀ = 3 μ M), which was five times more than the known cytotoxic drug fludarabine.

In silico ADME parameters of the new nucleoside analogs **9–22**, **27–28**, were used to calculate Lipinski's rules, solubility, percentage of absorption (%ABS) and topological polar surface area (TPSA) (Table 3) (see Supplementary documentation). All compounds have molecular weights smaller than 500 (377.44 > MW < 447.47), with the exception of the compounds **16** and **27**. The % ABS values were between the range of 49.72% and 68.73%, predicting that the synthesized nucleosides might penetrate through cell membrane.²⁶ Majority of the synthesized compounds possess the values of TPSA theoretically compatible with acceptable passive oral absorption. The results pointed that the novel derivatives would be potential drug candidates. To further support our *in silico* predictions, we calculated drug-likeness and drug-scores of these

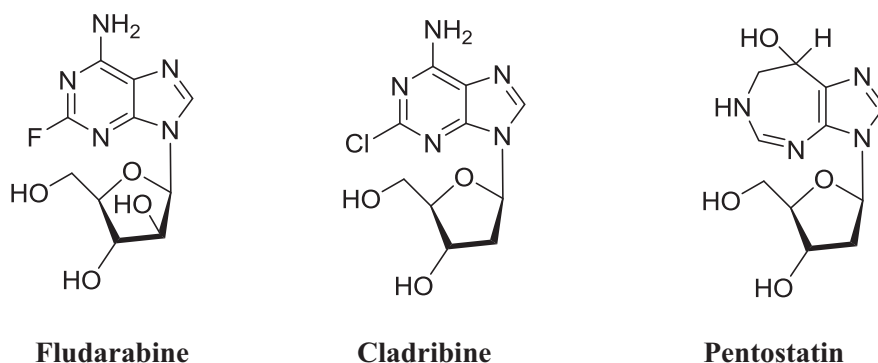


Fig. 1. Structures of fludarabine, cladribine and pentostatin.

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