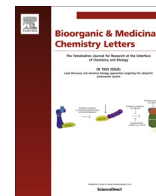




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## Structure-activity relationship study of $\beta$ -oxidation resistant indole-based 5-oxo-6,8,11,14-eicosatetraenoic acid (5-oxo-ETE) receptor antagonists

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### ABSTRACT

5-Oxo-6,8,11,14-eicosatetraenoic acid (5-oxo-ETE) is formed from 5S-hydroxy-6,8,11,14-eicosatetraenoic acid (5-HETE) by the 5-lipoxygenase (5-LO) pathway under conditions associated with oxidative stress. 5-Oxo-ETE is an important pro-inflammatory mediator, which stimulates the migration of eosinophils via a selective G-protein coupled receptor, known as the OXE receptor (OXE-R). Previously, we designed and synthesized structural mimics of 5-oxo-ETE such as **1** using an indole scaffold. In the present work, we added various substituents at C-3 of this moiety to block potential  $\beta$ -oxidation of the 5-oxo-valerate side chain, and investigated the structure-activity relationships of the resulting novel  $\beta$ -oxidation-resistant antagonists. Cyclopropyl and cyclobutyl substituents were well tolerated in this position, but were less potent as the highly active 3S-methyl compound. It seems likely that 3-alkyl substituents can affect the conformation of the 5-oxovalerate side chain containing the critical keto and carboxyl groups, thereby affecting interaction with the OXE-receptor.

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Arachidonic acid (AA, **2**) is an  $\omega$ -6 polyunsaturated fatty acid that is a key precursor of a large number of biologically active metabolites formed by different oxygenation pathways, one of the most important of which is the 5-lipoxygenase (5-LO) pathway.<sup>1,2</sup> 5-LO converts AA (**2**) in the presence of 5-LO activating protein (FLAP) to 5S-hydroperoxy-6,8,11,14-eicosatetraenoic acid (5S-HpETE, **3**) that is further metabolized to 5S-hydroxy-6,8,11,14-eicosatetraenoic acid (5S-HETE, **4**) and leukotrienes (LTs), Fig. 1. In the presence of nicotinamide adenine dinucleotide phosphate (NADP<sup>+</sup>), 5-HETE (**4**) is oxidized by the enzyme 5-hydroxyeicosanoid dehydrogenase (5-HEDH) to 5-oxo-6,8,11,14-eicosatetraenoic acid (5-oxo-ETE, **5**), which stimulates a variety of responses in human eosinophils and neutrophils.<sup>2</sup> Notably, 5-oxo-ETE (**5**) is the most potent chemoattractant for human

**Abbreviations:** AA, arachidonic acid; 5-HETE, 5-hydroxy-6E,8Z,11Z,14Z-eicosatetraenoic acid; 5-oxo-ETE, 5-oxo-6E,8Z,11Z,14Z-eicosatetraenoic acid; 5-HEDH, 5-hydroxyeicosanoid dehydrogenase; 5-HpETE, 5-hydroperoxyeicosatetraenoic acid; LT, leukotriene; NADP<sup>+</sup>, nicotinamide adenine dinucleotide phosphate; HPLC, high-performance liquid chromatography; TLC, thin layer chromatography; Hex, hexane; EtOAc, ethyl acetate; Rt, room temperature.

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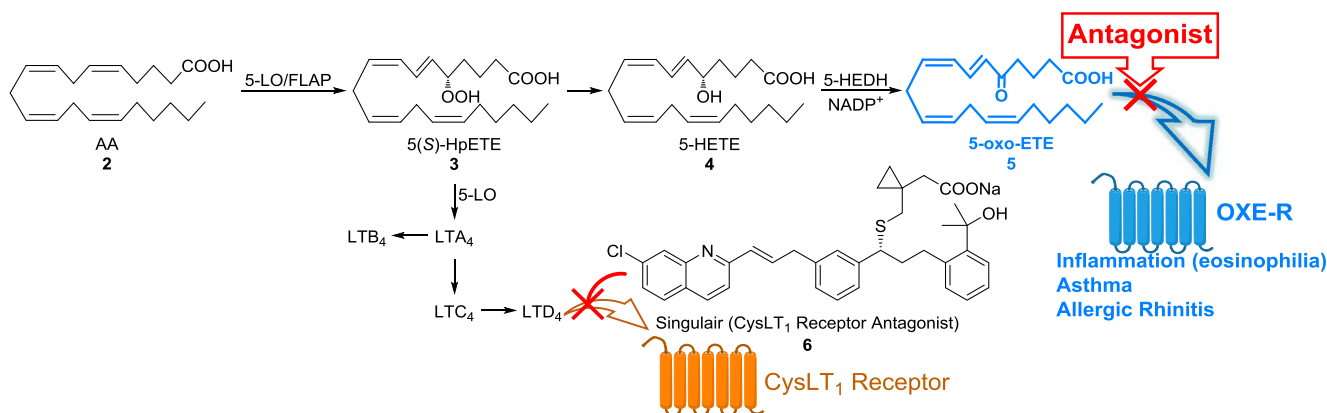
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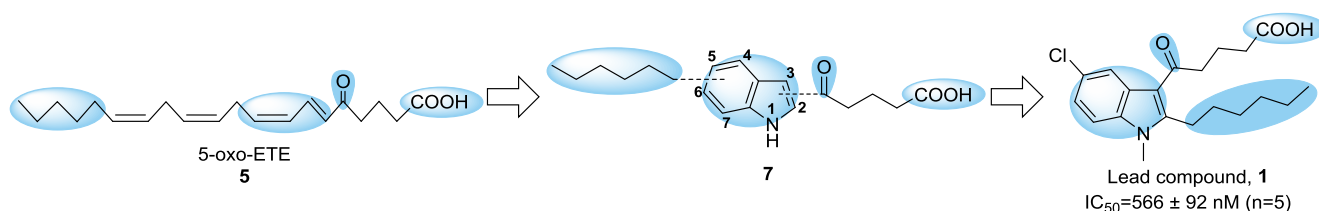
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eosinophils among lipid mediators.<sup>3</sup> The actions of 5-oxo-ETE are regulated by a selective G<sub>i</sub> protein-coupled receptor known as the OXE receptor (OXE-R), which is most highly expressed in eosinophils compared to other cell types. Because of these effects, activation of the OXE-R by 5-oxo-ETE could be a key event in the progression of eosinophilic diseases such as asthma.<sup>2,3</sup> Unlike cysteinyl leukotriene receptor (CysLT<sub>1</sub>) antagonists, such as Singulair (**6**, Fig. 1), at present there is no drug targeting the OXE receptor. For this reason, we embarked on a search for selective OXE-R antagonists that could block the potent proinflammatory effects of 5-oxo-ETE and thereby be useful therapeutic agents in eosinophilic diseases such as asthma. To this end, we designed 5-oxo-ETE receptor antagonists based on the structural mimics of 5-oxo-ETE (Fig. 2).<sup>5,6</sup>

To design OXE receptor antagonists, we first identified the essential structural elements of 5-oxo-ETE (**5**) required for the activation of the OXE-R. Structure-activity relationship (SAR) studies revealed that the C1-carboxyl group, the 5-keto group coupled to an adjacent conjugated system, and the hydrophobic region at the  $\omega$ -end of the molecule were all required for biological activity (**5**, Fig. 2).<sup>4</sup> Based on these results, we replaced the tetraene conjugated system of 5-oxo-ETE with an aromatic system such as



**Fig. 1.** Biochemical formation of eicosanoids by the 5-lipoxygenase (5-LO) pathway. The development of OXE receptor antagonist can stop eosinophil-induced inflammation.



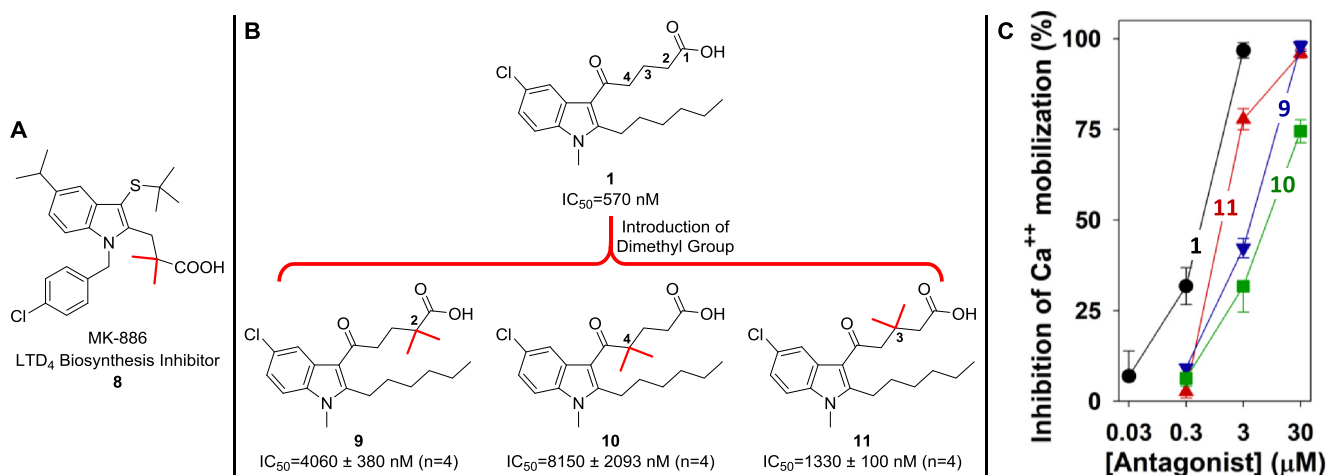
**Fig. 2.** The design of indole-based 5-oxo-EETE receptor antagonists as structural mimics of 5-oxo-EETE. Regions required for activation of the OXE-R are highlighted in blue.

naphthalene, benzofuran, and indole, to which we attached the carboxyl end and the alkyl ( $\omega$ ) end of 5-oxo-EETE as two separate groups. Among these aromatic scaffolds, indole (**7**, Fig. 2) gave the most promising results.<sup>5</sup> Using our  $\text{Ca}^{2+}$  mobilization assay we screened a series of indoles containing 5-oxovalerate and hexyl groups in different positions and found that an indole containing a hexyl group in the 2-position and a 5-oxovalerate group in the 3-position was the most potent. Further modification of this compound by the addition of a chloro group in the 5-position resulted in a 4-fold increase in antagonist potency and led to the lead compound **1**, which has an  $\text{IC}_{50}$  of 566 nM in inhibiting 5-oxo-EETE-induced  $\text{Ca}^{2+}$  mobilization in neutrophils.<sup>6</sup>

Because the 5-oxovalery side chain of the lead compound **1** can potentially undergo  $\beta$ -oxidation,<sup>7,8</sup> we decided to modify the carboxyl side chain of **1**, to block this metabolism. To accomplish this,

we introduced various substituents at the C-2 and C-3 positions of **1** (such as **9**, and **11** respectively, Fig. 3). In this paper, the syntheses and systematic SAR of these novel indole derivatives as OXE receptor antagonists are reported.

*Introduction of a dimethyl group at different positions of the lead compound (1).* In the development of a potent and specific leukotriene biosynthesis inhibitor, MK-886 (**8**, Fig. 3A), a dimethyl group was incorporated in an attempt to prevent  $\beta$ -oxidation of the propionic acid side chain (Fig. 3A).<sup>9</sup> We first substituted a dimethyl group at C-2 position of the lead compound **1** and prepared **9** (Fig. 3B). In the preparation of **9**, a small amount of the 4,4-dimethyl substituted compound **10** was generated as a side product (Scheme 2). We separated the two isomers by column chromatography and measured their antagonist activities. Both **9** and **10** showed substantial decreases in potency of seven and



**Fig. 3.** Panel A shows the structure of MK-886 (**8**), a potent leukotriene biosynthesis inhibitor. Panel B shows the structures of the dimethylated compounds (**9–11**). Panel C shows the concentration-response curves for the indole compounds (**1** and **9–11**).

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