



# Triterpenoid saponins from *Albizia lebbeck* (L.) Benth and their inhibitory effect on the survival of high grade human brain tumor cells



Olivier Placide Noté<sup>a,b,\*</sup>, Dong Jihu<sup>c</sup>, Cyril Antheaume<sup>d</sup>, Maria Zeniou<sup>c</sup>, Dieudonné Emmanuel Pegnyemb<sup>b</sup>, Dominique Guillaume<sup>e</sup>, Hervé Chneiweiss<sup>f</sup>, Marie Claude Kilhoffer<sup>c</sup>, Annelise Lobstein<sup>a</sup>

<sup>a</sup> Pharmacognosie et Molécules Naturelles Bioactives, Laboratoire d'Innovation Thérapeutique, UMR 7200, CNRS-Université de Strasbourg, Faculté de Pharmacie, 74 route du Rhin, F-67401 Illkirch Cedex, France

<sup>b</sup> Laboratoire de Pharmacochimie des Substances Naturelles, Département de Chimie Organique, Faculté de Sciences, Université de Yaoundé, BP 812 Yaoundé, Cameroon

<sup>c</sup> Chimie-Biologie Intégrative, Laboratoire d'Innovation Thérapeutique, UMR 7200, CNRS-Université de Strasbourg, Faculté de Pharmacie, 74 route du Rhin, F-67401 Illkirch Cedex, France

<sup>d</sup> Service Commun d'Analyse, UMR 7200, CNRS-Université de Strasbourg, Faculté de Pharmacie, 74 route du Rhin, F-67401 Illkirch Cedex, France

<sup>e</sup> UFR Médecine-Pharmacie, CNRS-UMR7312, 51 rue Cognacq Jay, 51100 Reims, France

<sup>f</sup> Neuroscience Paris Seine, IBPS, CNRS-UMR 8246, Inserm U1130, UPMC, 7 quai Saint Bernard, 75005 Paris, France

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## ABSTRACT

As part of our search of new bioactive triterpenoid saponins from Cameroonian Mimosaceae plants, phytochemical investigation of the roots of *Albizia lebbeck* led to the isolation of two new oleanane-type saponins, named lebbeckosides A–B (**1–2**). Their structures were established on the basis of extensive 1D and 2D NMR (<sup>1</sup>H, <sup>13</sup>C NMR, DEPT, COSY, TOCSY, ROESY, HSQC, and HMBC) and HRESIMS studies, and by chemical evidence. Compounds **1–2** were evaluated for their inhibitory effect on the metabolism of high grade human brain tumor cells, the human glioblastoma U-87 MG cell lines and the glioblastoma stem-like TG1 cells isolated from a patient tumor, and known to be particularly resistant to standard therapies. The isolated saponins showed significant cytotoxic activity against U-87 MG and TG1 cancer cells with IC<sub>50</sub> values of 3.46 μM and 1.36 μM for **1**, and 2.10 μM and 2.24 μM for **2**, respectively.

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## 1. Introduction

The genus *Albizia* comprises about 150 species widely distributed in the tropics, with the greatest diversity in Africa and South America.<sup>1</sup> Triterpenoid saponins are commonly described in this genus. Adiantifoliosides, grandibracteosides, gummiiferaosides, julibrosides, coriariosides, and albizosides are complex triterpenoid saponins isolated from the *Albizia* genus.<sup>2</sup> These glycosides, represent a class of very complex glycosides possessing a common aglycon unit, acacic acid, having various oligosaccharide moieties at C-3 and C-28 and an acyl group at C-21. They have been reported to inhibit the growth of tumor cells, and thus appear as a new potential class of anticancer natural triterpenoid saponins.<sup>2</sup> In order to discover new bioactive acacic acid glycosides, we screened the saponin content of Cameroonian Mimosaceae including *Albizia*, *Acacia*, and *Entada* genera. Three *Albizia* species, among which

*Albizia lebbeck*, were selected on the basis of their HPLC-DAD, LC-NMR, and LC-MS profiles.

*Albizia lebbeck* (L.) Benth is a pantropical species distributed in Africa, Asia, America, and Australia. In West Africa, it is traditionally used against diarrhea, dysentery, hemorrhoids, bronchitis, asthma, eczema, and leprosy.<sup>3</sup> In South-East Asia and Australia, the stem bark is used as a folk remedy to treat abdominal tumors, boils, cough, eye disorders, and lung ailments. It is also reported to be astringent, pectoral, rejuvenating, and tonic.<sup>4</sup> Nootropic and anxiolytic activities of a saponin fraction isolated from *A. lebbeck* leaves have been reported.<sup>5</sup> Oral administration of the saponin fraction isolated from *A. lebbeck* bark to male rats has been reported to significantly reduce fertility through reduction of sperm mobility and density.<sup>6</sup> Previous phytochemical studies of *A. lebbeck* stem bark reported the presence of glycosides of acacic acid lactones.<sup>7</sup>

In the present investigation on *A. lebbeck* roots, we report the isolation and structural characterization of two new acacic acid glycosides, named lebbeckosides A–B (**1–2**). The isolated compounds were evaluated for their inhibitory effect on the

\* Corresponding author. Tel.: +237 727200015; fax: +237 22239588.

E-mail address: [olivernote1@yahoo.fr](mailto:olivernote1@yahoo.fr) (O.P. Noté).

metabolism of high grade human brain tumor cells, namely the human glioblastoma U-87 MG cell lines and temozolomide-resistant glioblastoma stem-like cells isolated from a patient tumor, results are reported herein.

## 2. Results and discussion

The air-dried powdered roots of *A. lebbeck* (300 g) were extracted with aq-EtOH 70% using a soxhlet apparatus. After evaporation of the solvent, the resulting brown residue was partitioned between water and water-saturated *n*-BuOH. The *n*-BuOH fraction was then submitted to vacuum-liquid chromatography (VLC) on reversed-phase silica gel yielding three main fractions that were subjected to VLC on silica gel. Purification of the eluted subfractions by Semprep-HPLC afforded two new triterpenoid saponins (Chart 1).

Lebbeckoside A (**1**) was obtained as a white, amorphous powder. Its high-resolution electrospray ionization mass spectrometry (HRESIMS) (positive-ion mode) exhibited a pseudo-molecular ion peak at  $m/z$  2486.2146  $[M+NH_4]^+$  (calcd 2485.2147), consistent with a molecular formula of  $C_{118}H_{186}O_{54}$ . Upon acid hydrolysis with 2.0 M HCl, **1** gave an acacic acid lactone unit, which was identified with an authentic sample, and compound **1** also gave glucose (Glc), xylose (Xyl), fucose (Fuc), rhamnose (Rha), arabinose (Ara), and quinovose (Qui), which were identified by co-TLC with authentic samples. The absolute configuration of these sugar residues was determined to be *D* for Glc, Xyl, and Fuc, and *L* for Ara and Rha based on GC analysis of their trimethylsilyl thiazolidine derivatives.<sup>8</sup> Its  $^1H$  NMR spectrum showed seven angular methyl groups as singlets at  $\delta_H$  0.90, 1.01, 1.08, 1.10, 1.12, 1.31, and 1.84 (each 3H, s), one olefinic proton at  $\delta_H$  5.64 (1H, br s), and sugar proton signals at  $\delta_H$  4.89–6.42.  $^{13}C$  NMR spectrum showed two olefinic carbon signals at  $\delta_C$  123.5 and 143.8, suggesting that **1** was an oleanane type triterpenoid saponin. 1D ( $^1H$ ,  $^{13}C$  NMR, DEPT) and 2D (COSY, HSQC and HMBC) NMR techniques permitted assignments of all  $^1H$  and  $^{13}C$  NMR signals of the aglycone of **1**. This aglycone was thus recog-

nized to be acacic acid (3 $\beta$ ,16 $\alpha$ ,21 $\beta$ -trihydroxyolean-12-ene-28-oic acid) by comparison of its  $^1H$  and  $^{13}C$  NMR signals with those reported in the literature.<sup>9–21</sup> The downfield position of the axial group at C-14 (Me-27,  $\delta$  1.84) in the  $^1H$  NMR spectrum, implied an additional axial ( $\alpha$ ) hydroxyl group at C-16. The ROESY correlations observed between H-21 ( $\delta$  5.37) and H-29 ( $\delta$  1.01, s), suggested an  $\alpha$ -axial orientation of H-21, as well as between H-3 ( $\delta$  3.49) and H-5 ( $\delta$  0.82) indicated the  $\alpha$ -axial orientation of the two protons. The 3,21-hydroxy groups and 28-carbonyl group of the aglycone carried a sugar moiety, respectively, as evidenced by the glycosylation- and acylation-induced shifts observed at  $\delta_C$  89.0 (deshielded signal for C-3 of the aglycon), 174.9 (shielded signal for C-28 of the aglycon), and 77.6 (deshielded signal for C-21 of the aglycon), indicating that **1** was a 21-acyl 3,28-bidesmosidic acacic acid derivative with sugar chains linked to C-3 and C-28 of the aglycon through an ether and ester bond, respectively, and with an acyl group attached at C-21.

The  $^1H$  NMR spectrum of **1** showed 10 anomeric protons at  $\delta_H$  4.91 [d,  $J$  = 8.0 Hz, glucose (Glc I)], 4.97 [d,  $J$  = 8.0 Hz, fucose (Fuc)], 5.09 [d,  $J$  = 7.1 Hz, xylose (Xyl I)], 6.16 [d,  $J$  = 8.0 Hz, glucose (Glc II)], 6.42 [br s, rhamnose (Rha)], 5.42 [d,  $J$  = 8.1 Hz, glucose (Glc III)], 5.30 [d,  $J$  = 7.1 Hz, xylose (Xyl II)], 4.90 [d,  $J$  = 8.0 Hz, quinovose (Qui I)], 4.89 [d,  $J$  = 8.0 Hz, quinovose (Qui II)], and 4.97 [d,  $J$  = 8.0 Hz, quinovose (Qui III)], which correlated with ten anomeric carbon atom resonances at  $\delta_C$  105.3, 103.7, 107.3, 95.6, 101.6, 106.4, 106.8, 99.7, 99.6, and 97.3, respectively, in the HSQC spectrum (Tables 1 and 2). The  $^1H$  and  $^{13}C$  NMR data (Tables 1 and 2) of the monosaccharide residues were assigned starting, either from the readily identifiable anomeric proton of each hexosyl or pentosyl unit, or from the  $CH_3$ -proton doublet of each 6-deoxyhexosyl unit, by means of TOCSY, HSQC, and HMBC spectra obtained for this compound. The anomeric centers of the *D*-glucose, *D*-fucose, *D*-quinovose, and *D*-xylose units were each determined to have a  $\beta$ -configuration based on large  $^3J_{H-1,H-2}$  values. And the  $\alpha$ -anomeric configuration of the *L*-rhamnose was judged by the broad singlet of the anomeric proton and the chemical shift value of C-5 ( $\delta$  68.8).<sup>22</sup> In

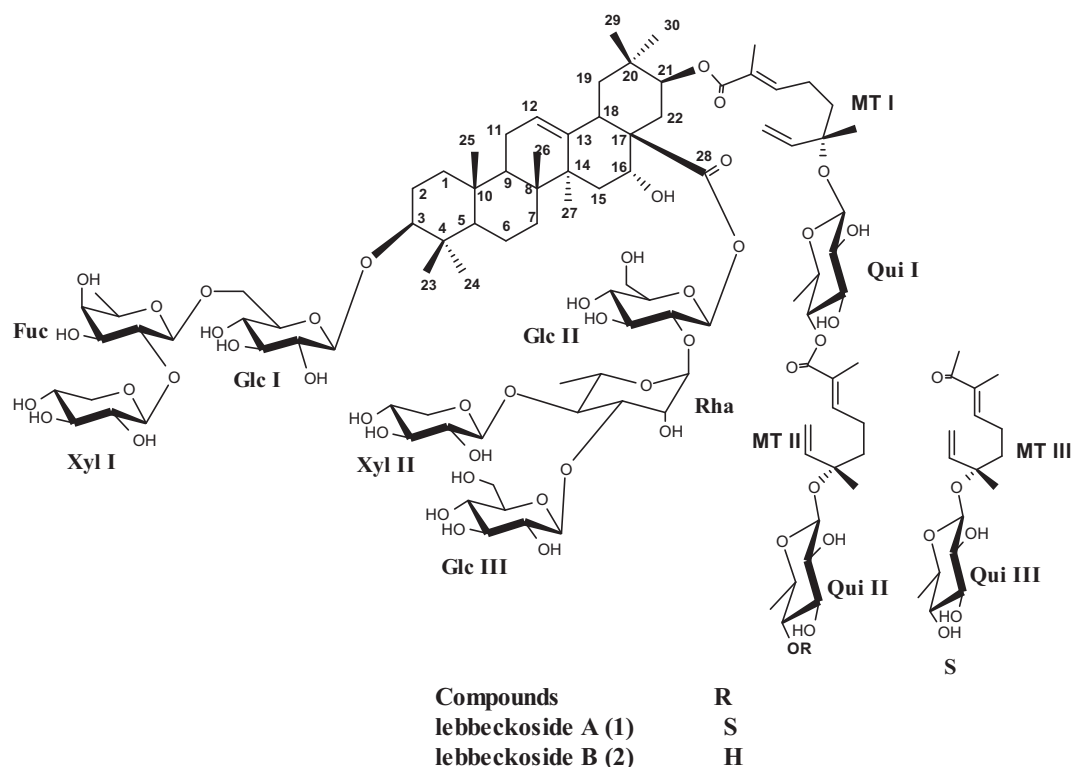


Chart 1. Structures of triterpene saponins **1–2**.

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