Accepted Manuscript

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PII: S0032-3861(17)31237-5

DOI: 10.1016/j.polymer.2017.12.059

Reference: JPOL 20248

To appear in: *Polymer*

Received Date: 21 November 2017

Revised Date: 19 December 2017

Accepted Date: 23 December 2017

Please cite this article as: Morita S, Takasu A, Adhesion control of Human umbilical vein endothelial cells using clickable poly(2-oxazoline)-grafted biosynthesized extracellular matrix protein, *Polymer* (2018), doi: 10.1016/j.polymer.2017.12.059.

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Revised 12/18/2017

Adhesion Control of Human Umbilical Vein Endothelial Cells Using Clickable Poly(2-oxazoline)-Grafted Biosynthesized Extracellular Matrix Protein

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Key Words: T7 expression system; in vivo incorporation; artificial extracellular matrix protein (aECM); *p*-azidophenylalanine; alkyne-containing poly(2-oxazoline), click reaction, ring-opening polymerization

Abstract: A bacterial expression host was used for *in vivo* incorporation of *p*-azidophenylalanine (p-N₃Phe), a phenylalanine (Phe) analog, into an artificial extracellular matrix protein (aECM-CS5-ELF). The host harbors the mutant phenylalanyl-*t*RNA synthetase (PheRS) which has an enlarged binding pocket, where the Ala294Gly/Thr251Gly mutant PheRS (PheRS**) was expressed under the control of T7 promoters. Biosynthesized aECM-CS5-ELF containing p-N₃Phe (aECM-CS5-ELF-N₃) was coupled with alkyne-containing poly(2-oxazoline) prepared via ring-opening polymerization of 2-oxazolines (2-methyl-2-oxazoline, 2-ethyl-2-oxazoline, and 2-iso-propyl-2-oxazoline) using methyl triflate (TfOMe) and propiolic acid as the initiator and terminator, respectively. A copper-catalyzed alkyne-azide click reaction was used to graft the poly(2-oxazoline)s onto the aECM. Examination of the solubility in organic and aqueous media, and the lower critical transition temperature (LCST), revealed a dependence on the incorporation ratio of *p*-N₃Phe and graft chain species; the LCST behavior was markedly altered when poly(2-oxazoline) moieties were present as side chains. Circular dichroism measurements indicate that grafting was not responsible for conformational changes, because the conformation was retained both at below and above the LCST. Specific adhesion and subsequent temperature-sensitive detachment of human umbilical vein endothelial cells (HUVECs) onto the cross-linked aECM-CS5-ELF-N₃-graft-poly(2-oxazoline) surfaces was also demonstrated.

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